

INSTITUTE OF MARINE RESEARCH
HAVFORSKNINGSINSTITUTTET



Prospects for probiotics with Atlantic halibut larvae

Øivind Bergh

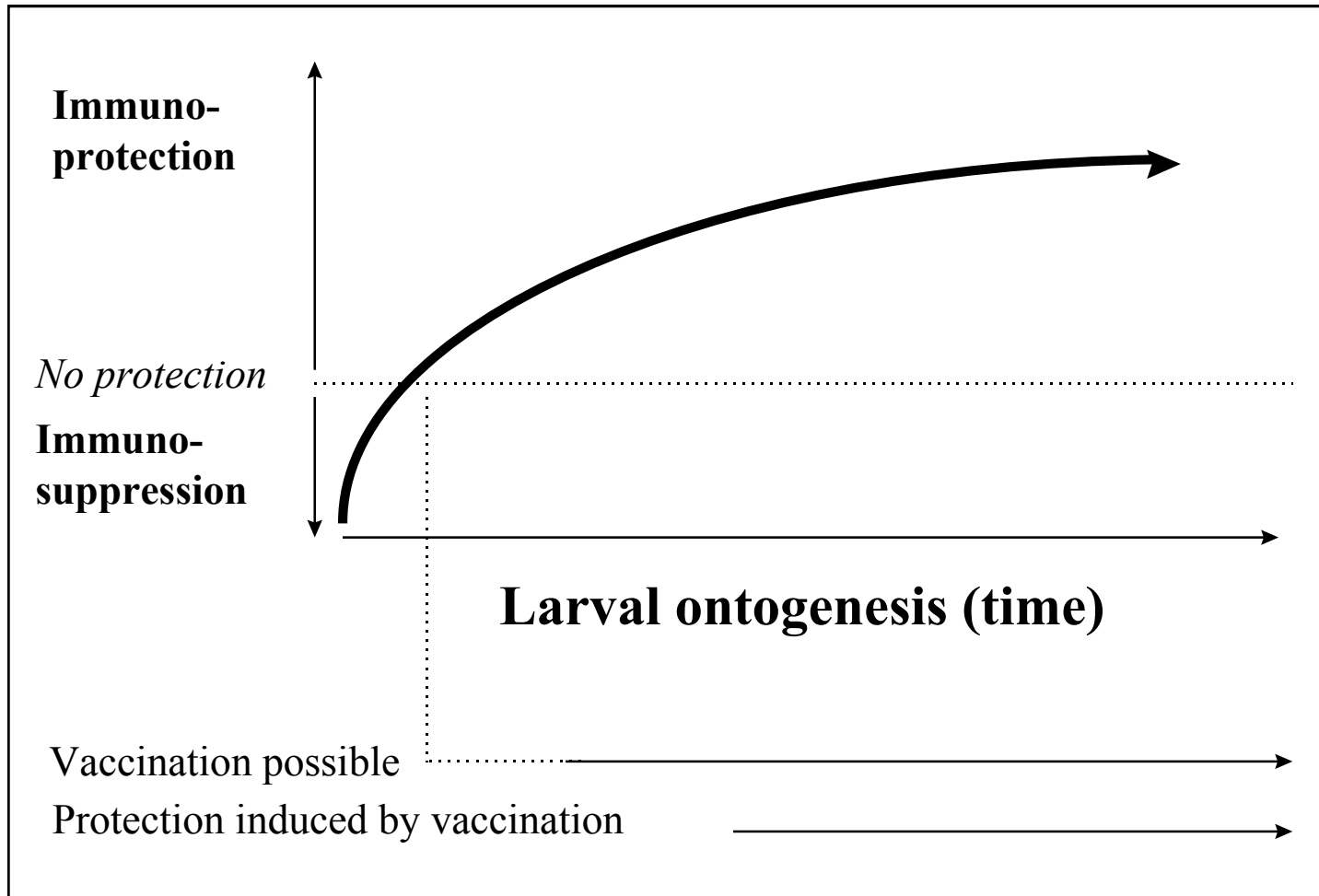
Institute of Marine Research, Bergen, Norway



What is probiotics

- Beneficial bacteria which when added to the water or the food affects the health of the host in a positive way
- Often, but not always related to suppression of pathogens





Vadstein, Mo and Bergh 2004

Why challenge?

- **Basic questions:**
 - Is "the bug" virulent?
 - How does it affect the host?
 - How does it enter the host?
- **Applied:**
 - How can we protect the host
 - How can we verify that the protection is effective?



Collaborators: PROAQUA

- Danish Technological University
- University of Bergen
- HMRC – Crete



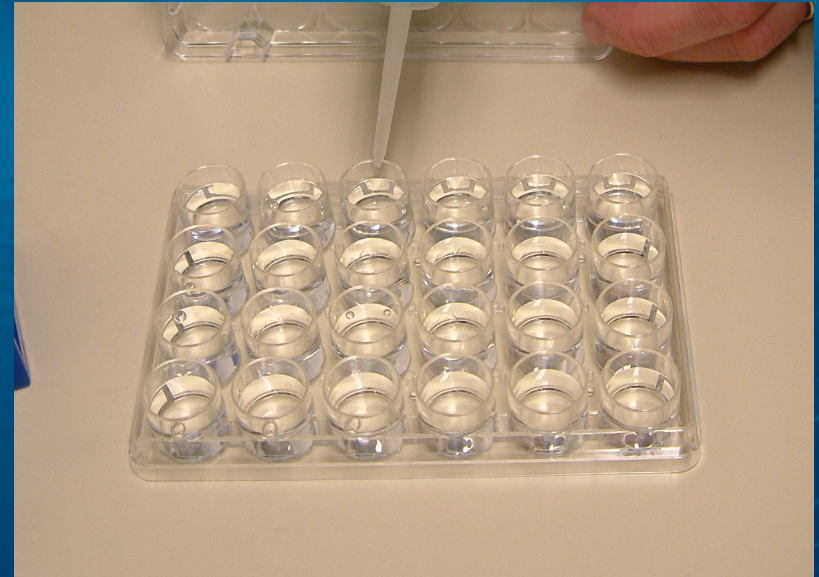
Sequencing of community

- To be performed by Dr. Nina Sandlund

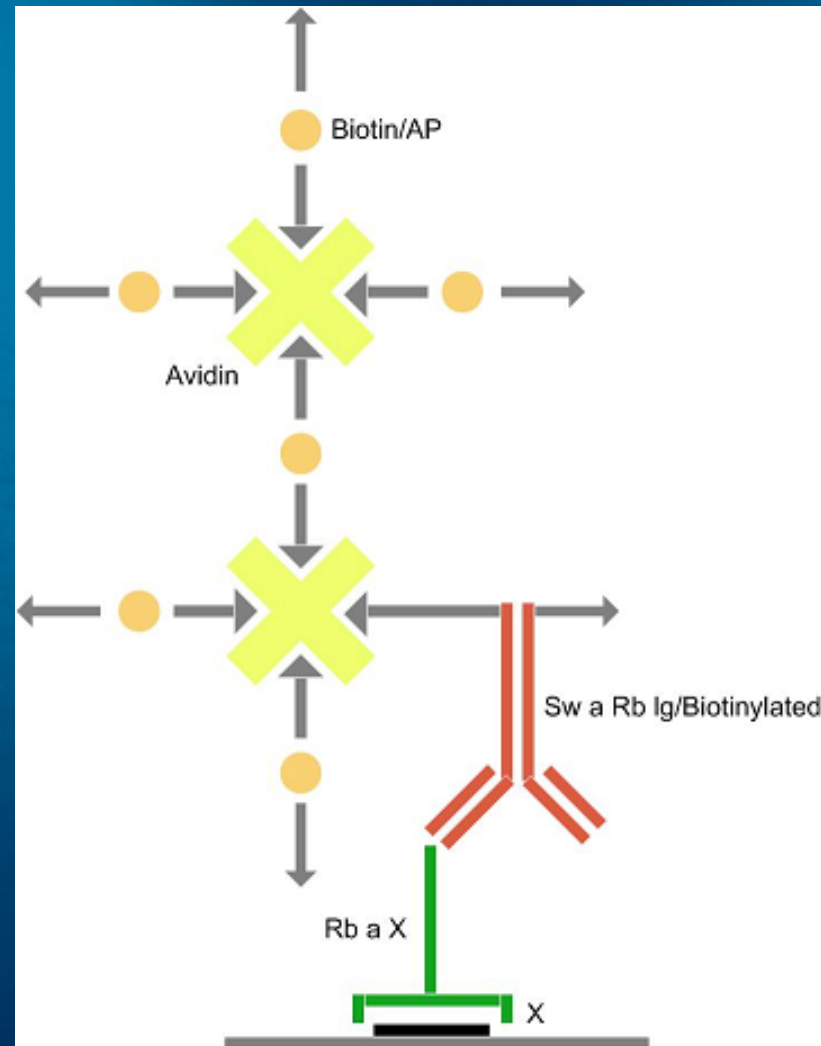


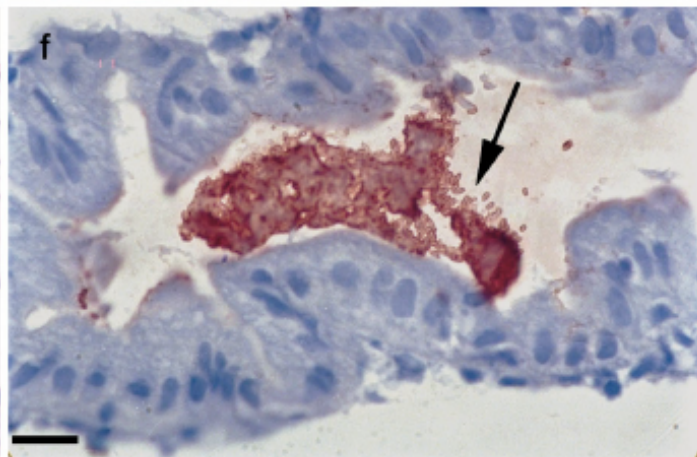
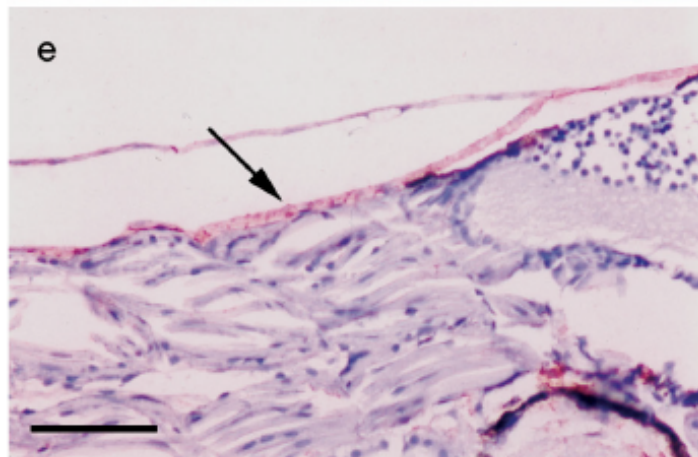
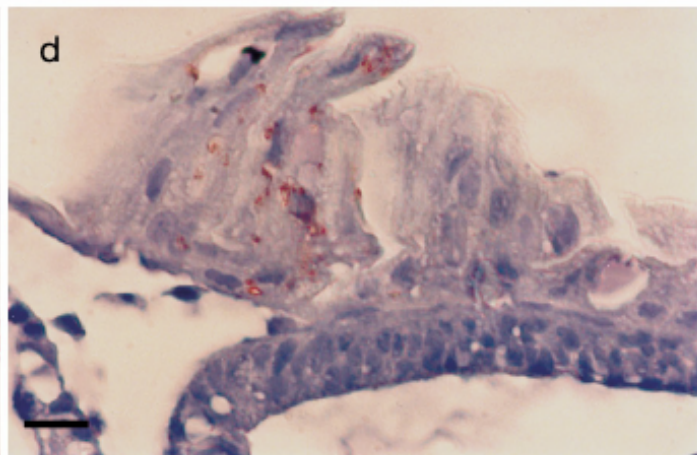
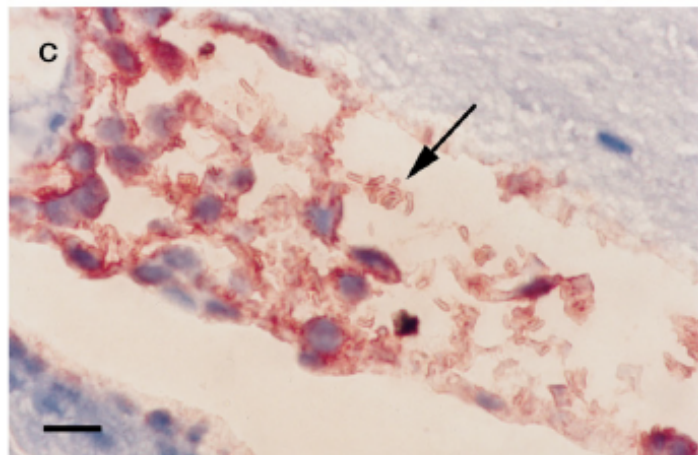
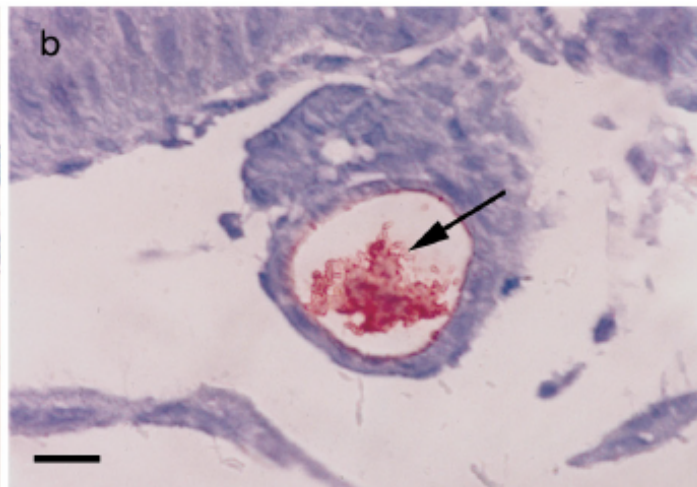
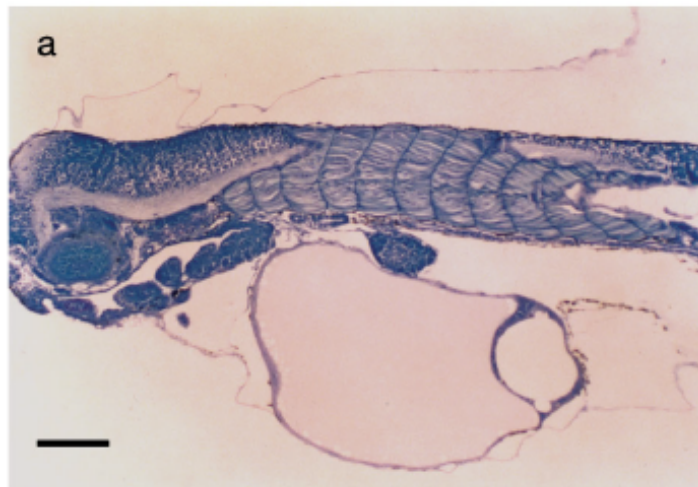
Challenge experiments on yolk sac larvae

- Rearing of larvae in multiwell dishes
- 72 independent parallel wells
- One egg/larvae per well
- Larvae hatches in well, lives until end of yolk sac period
- Protocol developed from various challenge experiments during two decades:
 - Bergh et al. 1991 J. Fish Dis.
 - Sandlund et al. 2010 Dis. Aquat. Org.



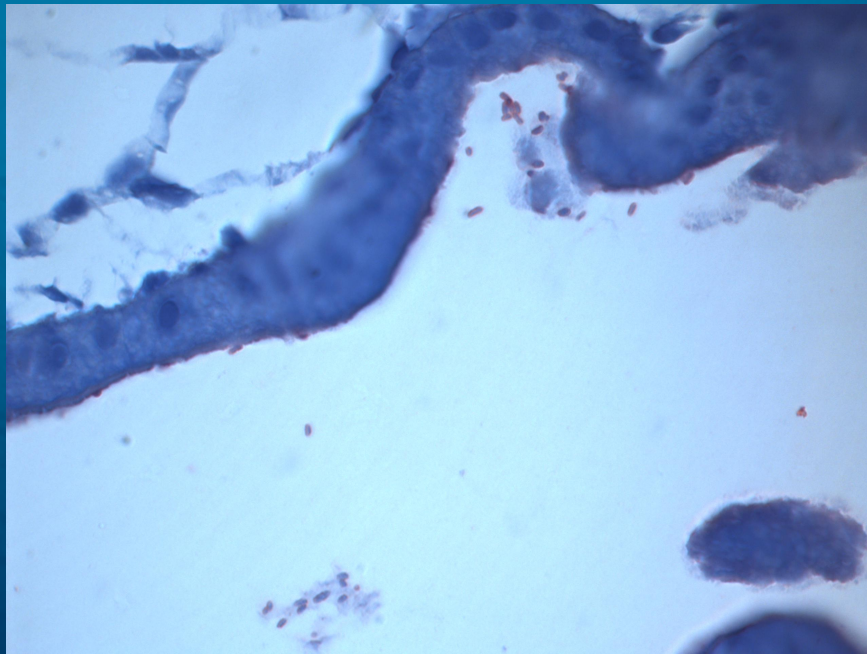
Immunohistochemistry



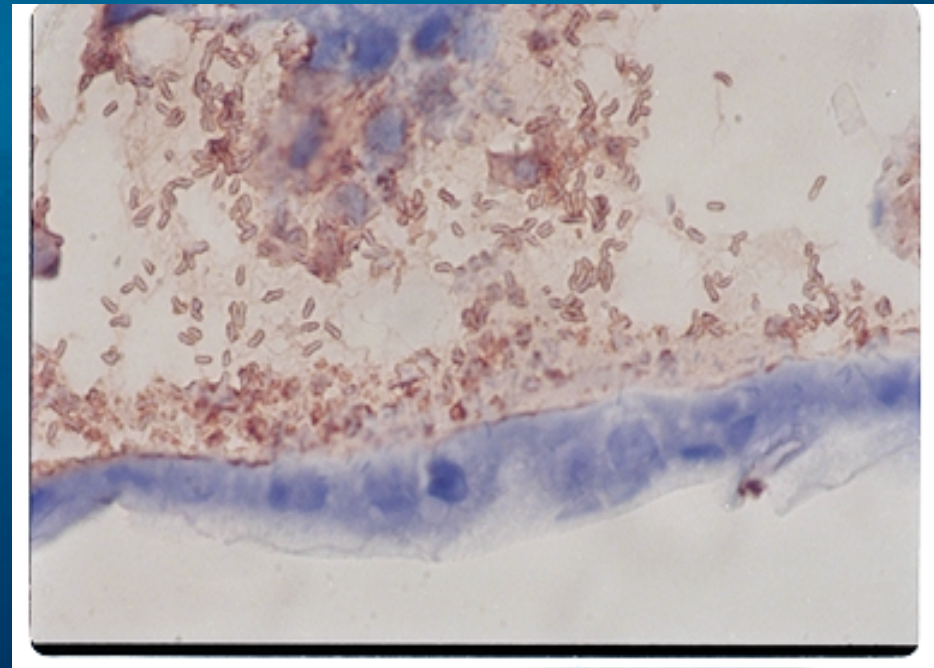


Immunohistochemistry – yolk sac larvae

Intestinal epithelium - cod



Epidermis - halibut





ELSEVIER

Available online at www.sciencedirect.com

SCIENCE @ DIRECT®

Aquaculture 255 (2006) 323–333

Aquaculture

www.elsevier.com/locate/aqua-online

Probiotic effect in vivo of *Roseobacter* strain 27-4 against *Vibrio (Listonella) anguillarum* infections in turbot (*Scophthalmus maximus* L.) larvae

Miquel Planas ^{a,*}, María Pérez-Lorenzo ^a, Mette Hjelm ^b, Lone Gram ^b,
Ingrid Uglenes Fiksdal ^c, Øivind Bergh ^c, José Pintado ^a

^a *Instituto de Investigaciones Marinas (CSIC), Eduardo Cabello 6, 36208 Vigo, Galicia, Spain*

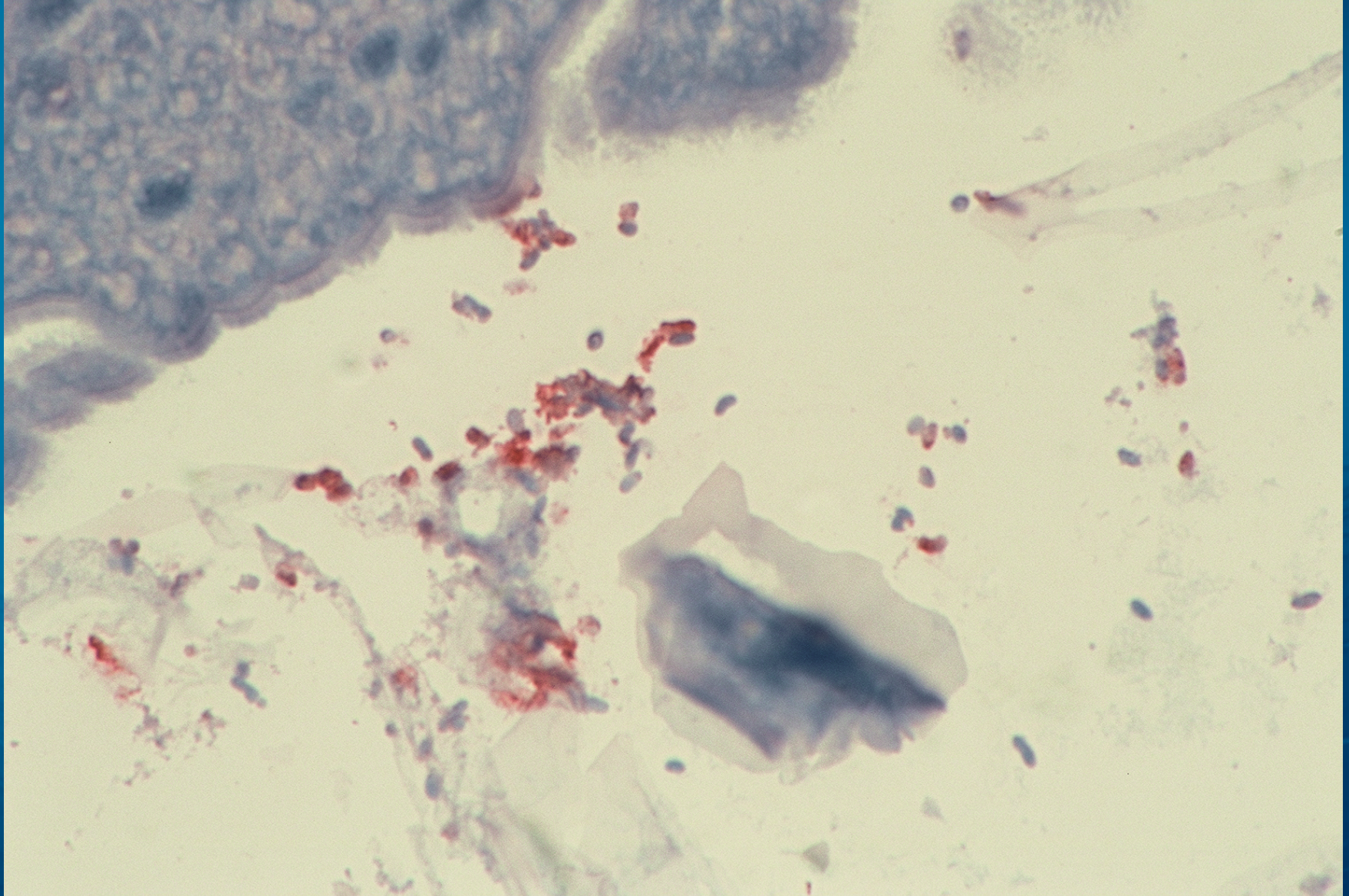
^b *Danish Institute for Fisheries Research, Department of Seafood Research, Søtofts Plads, c/o Technical University of Denmark Bldg. 221, DK-2800 Kgs. Lyngby, Denmark*

^c *Institute of Marine Research, PO Box 1870, N-5817, Bergen, Norway*

Received 3 May 2005; received in revised form 17 November 2005; accepted 19 November 2005

**Addition of *Roseobacter*(*Rugeria*) 27-4
positively affected survival**





**Note: probiotics in lumen
– NOT attached to epithelium**

Comparative mortality pilot

5 different *Vibrio* spp

3 x *V. anguillarum*

V. splendidus

V. salmonicida

3 different fish larvae: turbot, halibut, cod

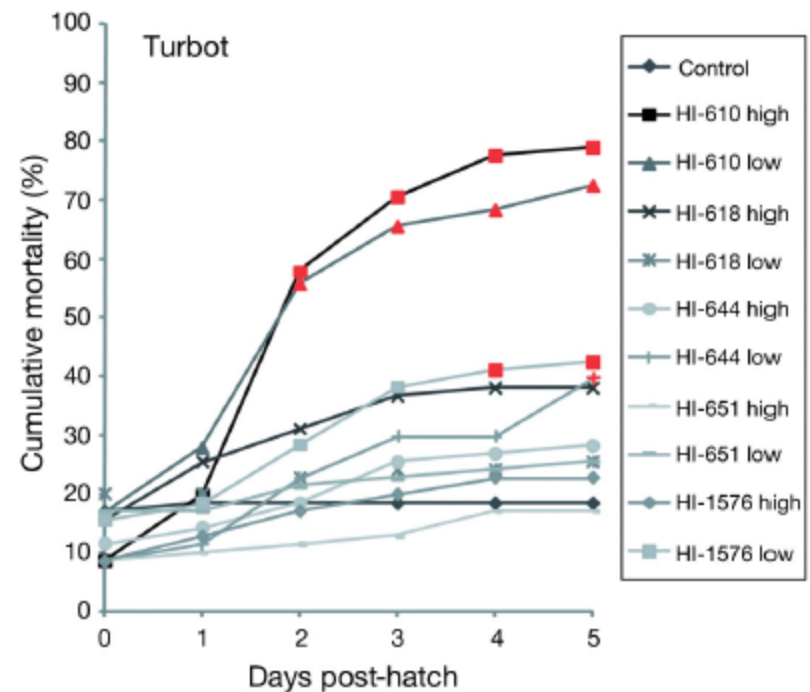


Fig. 1. *Scophthalmus maximus*. Cumulative percentage mortality of turbot larvae challenged with the bacterial strains HI-610, HI-618, HI-644, HI-651, and HI-1576 (see Table 1 for bacterial strains used. High: challenge dose 10^6 CFU ml^{-1} , low: challenge dose 10^4 CFU ml^{-1} , control: unchallenged larvae. Day 0: day of hatching. Red symbols: mortality rates significantly different from the control ($p < 0.01$ Bonferroni correction))



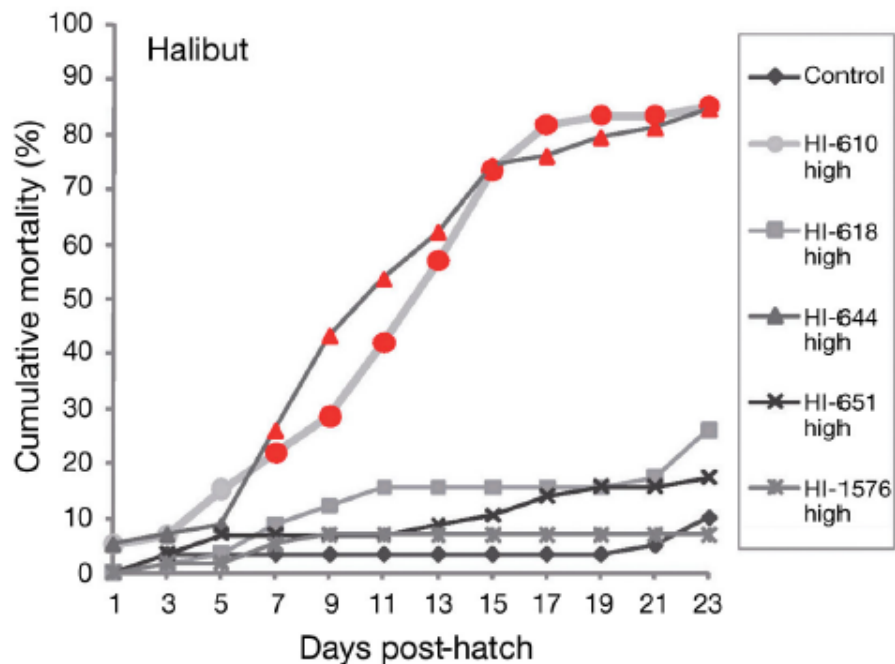


Fig. 2. *Hippoglossus hippoglossus*. Cumulative percentage mortality of halibut larvae challenged with the bacterial strains HI-610, HI-618, HI-644, HI-651, and HI-1576 (see Table 1 for bacterial strains used). High: challenge dose 10^6 CFU ml^{-1} , control: unchallenged larvae. Red symbols: mortality rates significantly different from the control ($p < 0.01$ Bonferroni correction)

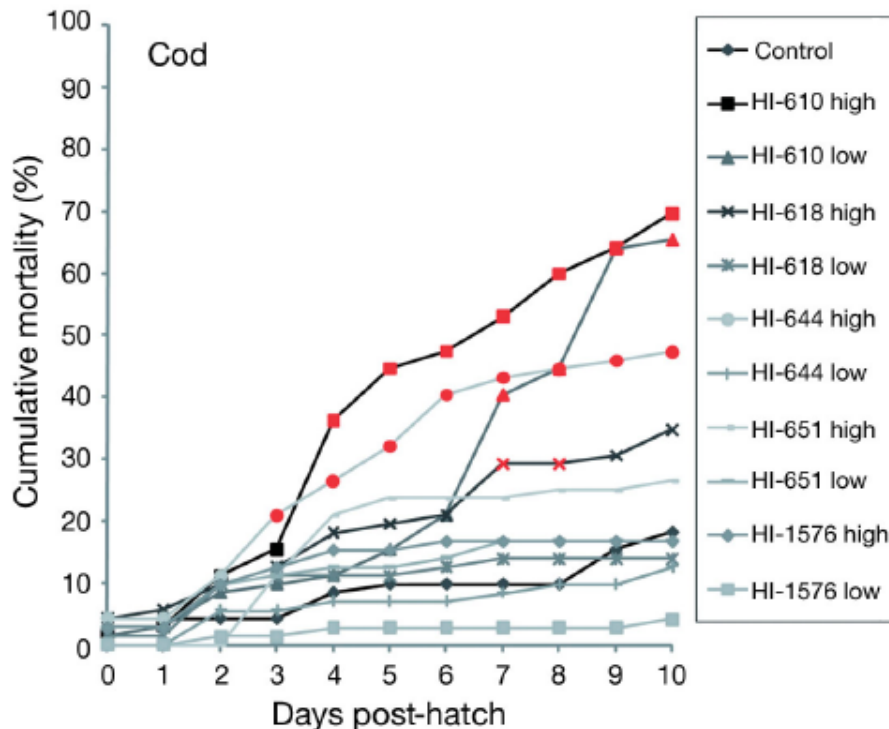
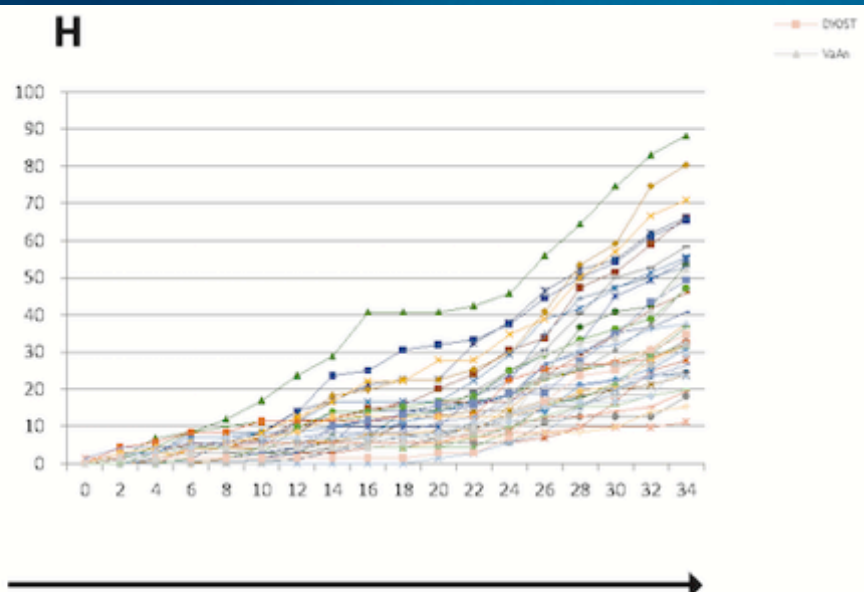
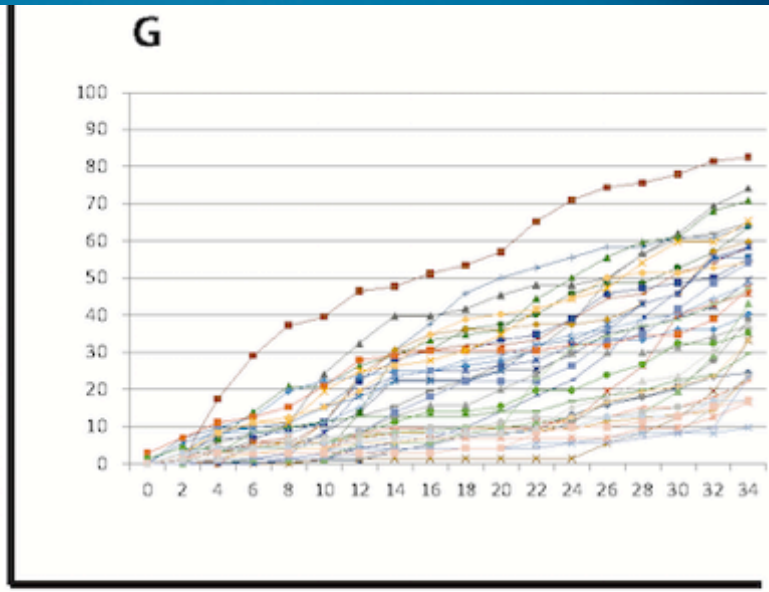


Fig. 3. *Gadus morhua*. Cumulative percentage mortality of cod larvae challenged with the same bacterial strains and doses as in Fig. 1. Day 0: day of hatching. Red symbols: mortality rates significantly different from the control group ($p < 0.01$ Bonferroni correction)





—■— DHOST
—▲— VAA

Dph →



Table 1. Bacterial strains and plasmids.

Strain or plasmid	Genotype or relevant markers
Strains	
<i>P. gallaeciensis</i> BS107 (DSM17395)	Wild type
<i>P. gallaeciensis</i> BS107-Pda8	CDS104961::EZ-Tn5, Kan ^R
<i>P. gallaeciensis dsRed</i>	MiniTn7(Gm ^R)P _{A1/04/03} DsRedExpress-a
<i>V. anguillarum</i> NB10	Serotype O1, cm ^R , PA1/04/03-RBSII- <i>gfp</i> mut3*-T1
<i>V. anguillarum</i> HI610	Serotype O2 α
Plasmids	
EZ-Tn5 TM Transposome	EZ-Tn5<R6K γ ori, Kan ^R >Tnp
pAKN132	miniTn7(Gm)P _{A1/04/03} DsRedExpress-a
pUX-BF13	Helper plasmid: Tn7 transposase proteins
pPDA11	<i>tdaCp::gfp</i> ligated into broad host range vector pRK415

doi:10.1371/journal.pone.0043996.t001

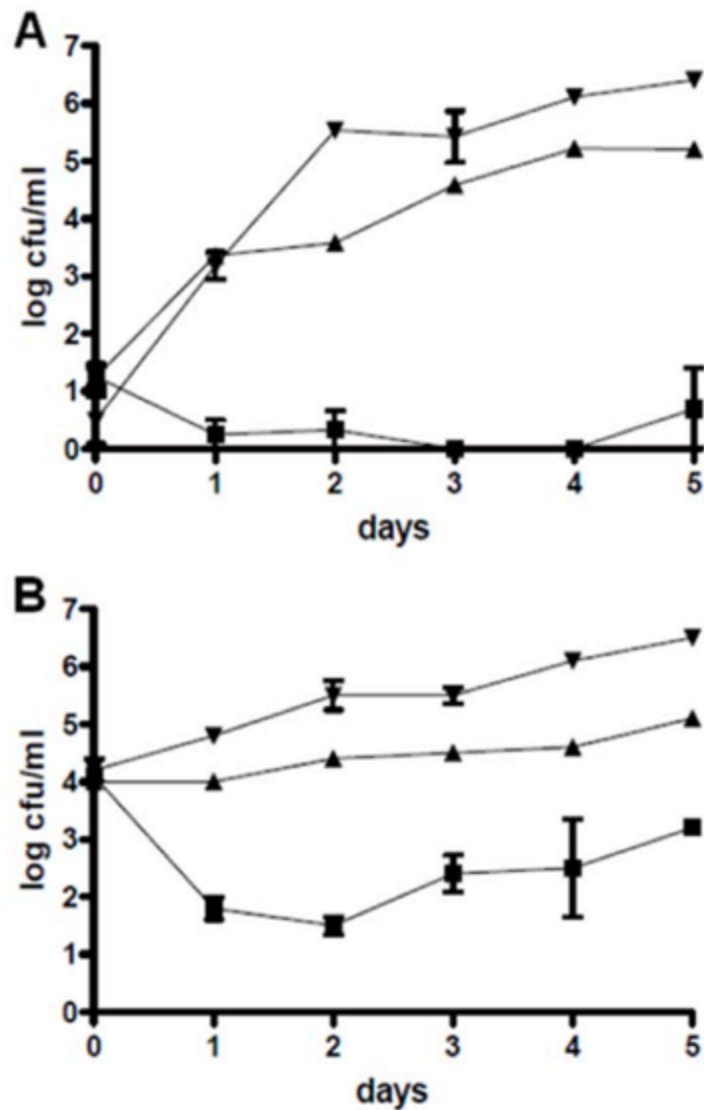


Figure 3. Reduction of *V. anguillarum* in cultures of *Tetraselmis suecica* by *Phaeobacter gallaeciensis*. Colony-forming units of *V. anguillarum* inoculated at 10^1 cfu/ml (A) and at 10^4 cfu/ml (B) in presence of *P. gallaeciensis* wild type (■), in presence of the *P. gallaeciensis* TDA-negative mutant (▲), and in the monoxenic control (▼).

doi:10.1371/journal.pone.0043996.g003



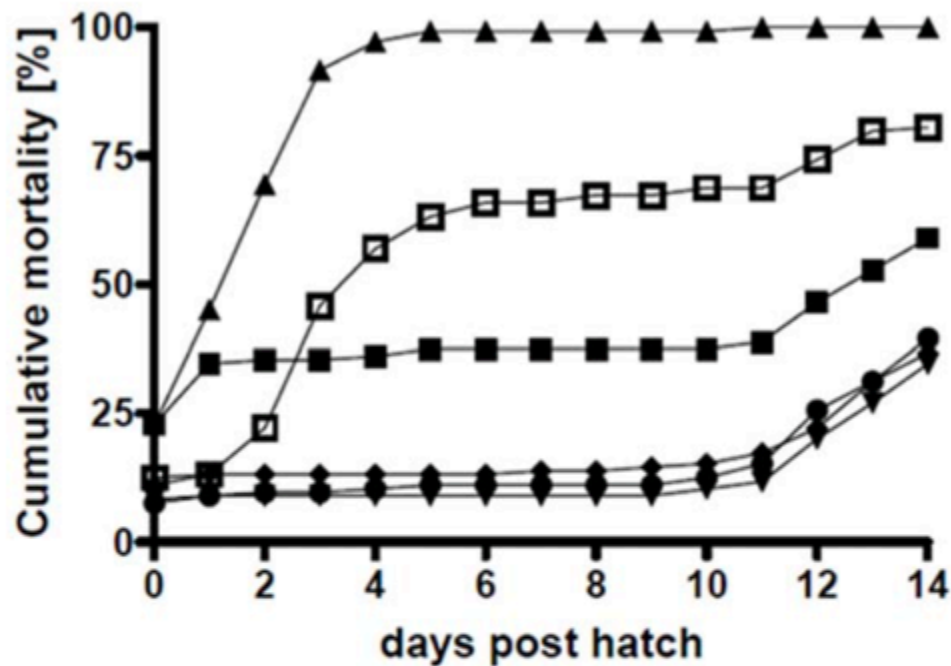


Figure 7. Mortality of cod larvae during the challenge trials. Mean values of two independent triplicate experiments. The single-larvae cultures were simultaneously inoculated with *P. gallaeciensis* wild type and *V. anguillarum* (T5, ●), or with the TDA-negative mutant of *P. gallaeciensis* and *V. anguillarum* (T6, □). Unexposed larvae and larvae exposed to single bacterial strains acted as controls: Negative Control (T1, ■), only *V. anguillarum* (T2, ▲), only *P. gallaeciensis* wild type (T3, ▼), and only *P. gallaeciensis* TDA-negative mutant (T4, ◆).
doi:10.1371/journal.pone.0043996.g007





Screening and characterisation of potentially pathogenic bacteria associated with Atlantic cod *Gadus morhua* larvae: bath challenge trials using a multidish system

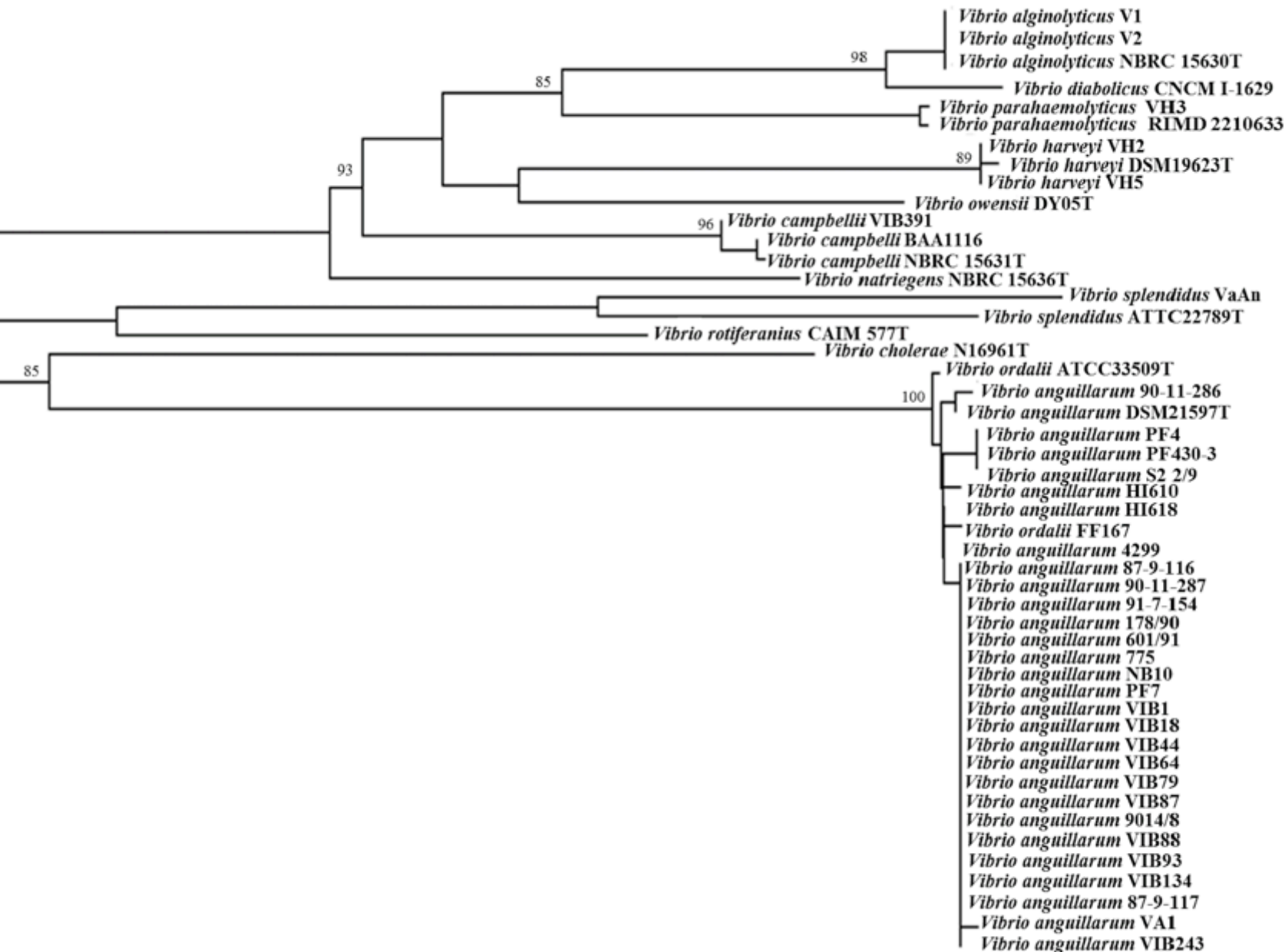
Nina Sandlund*, Øivind Bergh

Institute of Marine Research, PO Box 1870 Nordnes, 5817 Bergen, Norway

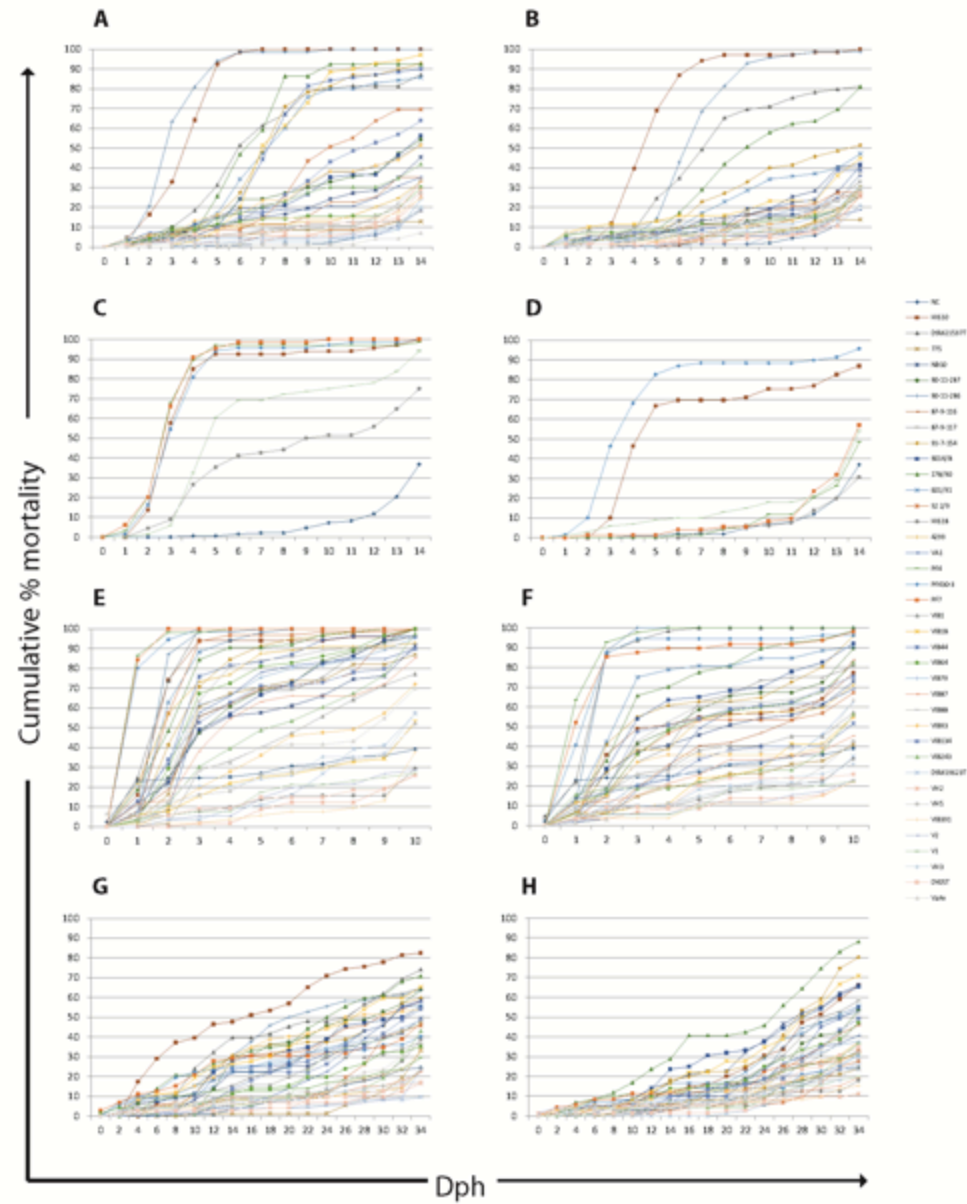
Most bacteria associated with larvae are opportunists, not able to cause mortality alone



V. anguillarum is a "true" pathogen



0.02



1 **Comparative assessment of *Vibrio* virulence in marine fish larvae**

2

3 Anita Rønneseth¹⁺, Daniel Castillo²⁺, Paul D'Alvise^{3+,6}, Øyvind Tønnesen¹, Gyri Haugland¹,
4 Torben Grotkjær³, Kirsten Engell-Sørensen⁴, Louise Nørremark⁴, Øivind Bergh⁵, Heidrun I.
5 Wergeland¹ and Lone Gram³

6

7 ¹ University of Bergen, Department for Biology, Thormøhlensgaten 55, N-5020 Bergen

8 ² University of Copenhagen, Marine Biology Section, Strandpromenaden 5, DK-3000
9 Helsinore

10 ³ Technical University of Denmark, Department of Biotechnology and Biomedicine,
11 Matematiktorvet 301, DK-2800 Kgs. Lyngby

12 ⁴ Fishlab, Terp Skovvej 107b, DK-8270 Højberg

13 ⁵ Institute for Marine Research, Nordnesgaten 50, N-5005 Bergen

14 ⁶ Present address: University of Hohenheim, Institute for Animal Science, Garbenstraße 17,
15 D-70599 Stuttgart

16

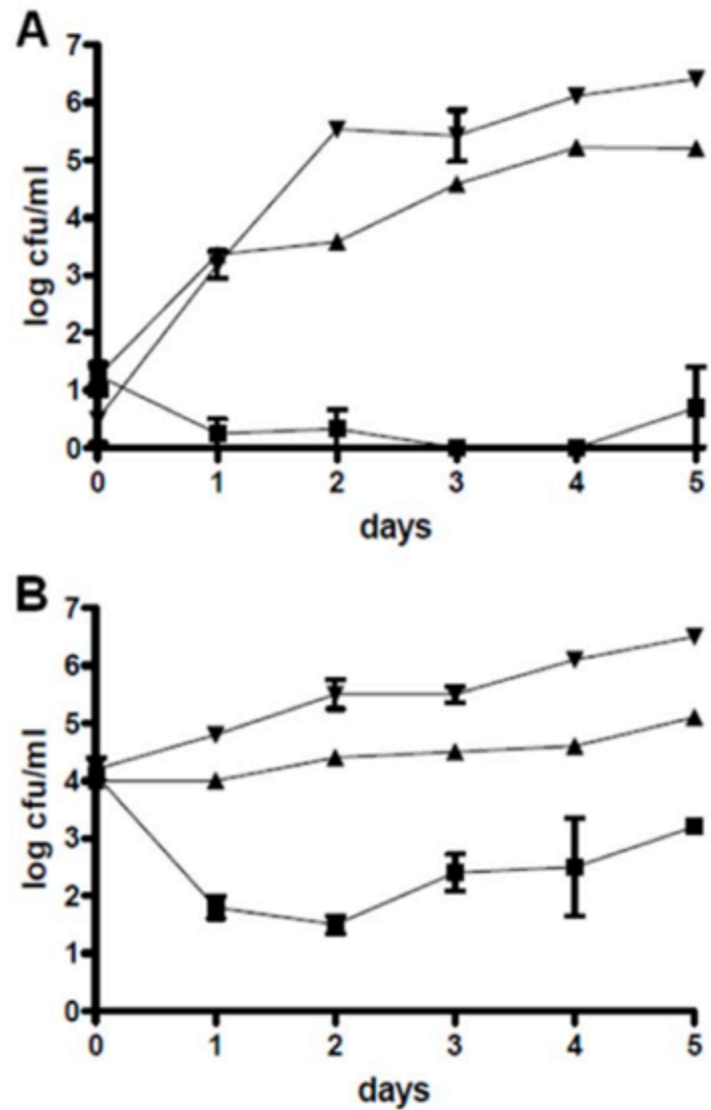


Figure 3. Reduction of *V. anguillarum* in cultures of *Tetraselmis suecica* by *Phaeobacter gallaeciensis*. Colony-forming units of *V. anguillarum* inoculated at 10¹ cfu/ml (A) and at 10⁴ cfu/ml (B) in presence of *P. gallaeciensis* wild type (■), in presence of the *P. gallaeciensis* TDA-negative mutant (▲), and in the monoxenic control (▼).

doi:10.1371/journal.pone.0043996.g003



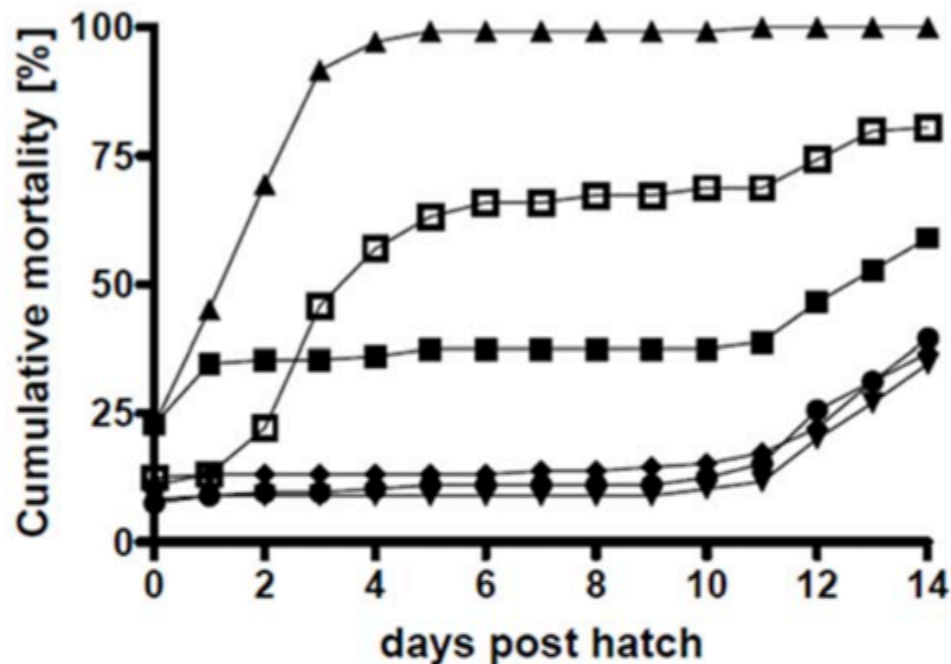
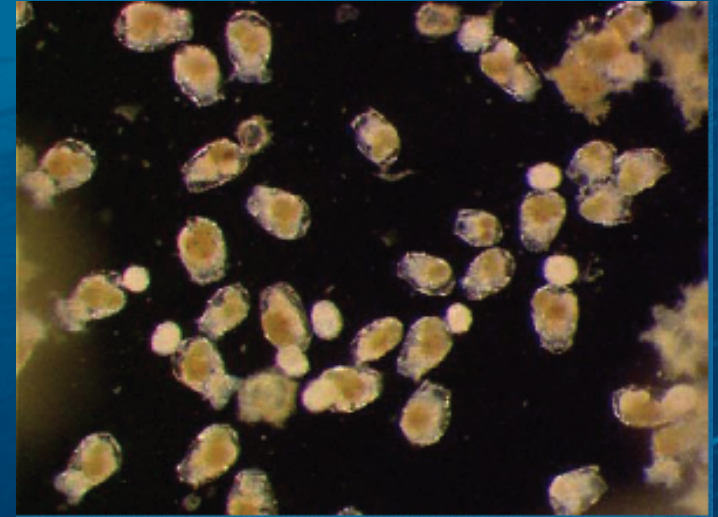


Figure 7. Mortality of cod larvae during the challenge trials. Mean values of two independent triplicate experiments. The single-larvae cultures were simultaneously inoculated with *P. gallaeciensis* wild type and *V. anguillarum* (T5, ●), or with the TDA-negative mutant of *P. gallaeciensis* and *V. anguillarum* (T6, □). Unexposed larvae and larvae exposed to single bacterial strains acted as controls: Negative Control (T1, ■), only *V. anguillarum* (T2, ▲), only *P. gallaeciensis* wild type (T3, ▼), and only *P. gallaeciensis* TDA-negative mutant (T4, ◆).
doi:10.1371/journal.pone.0043996.g007

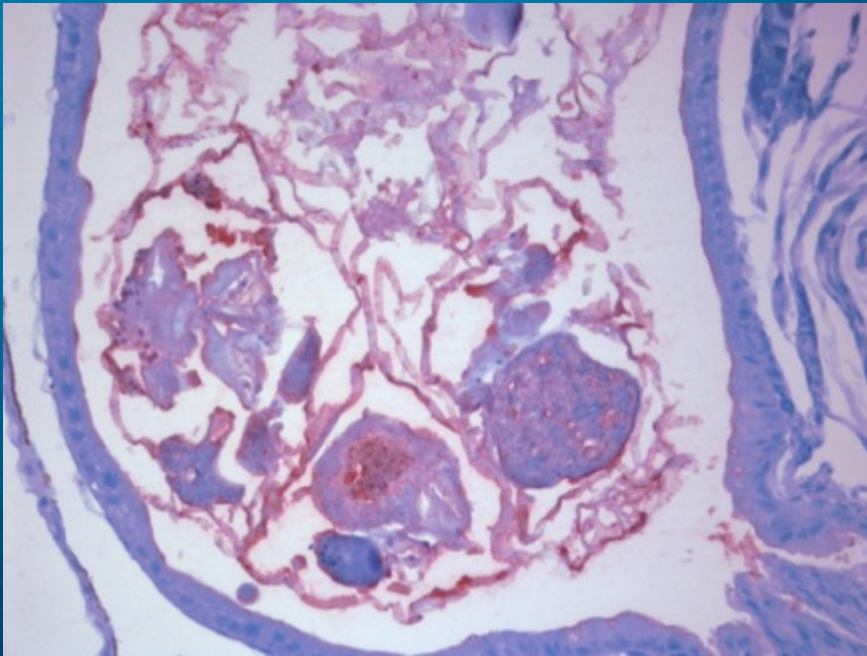


Challenges wia live feed = oral administration

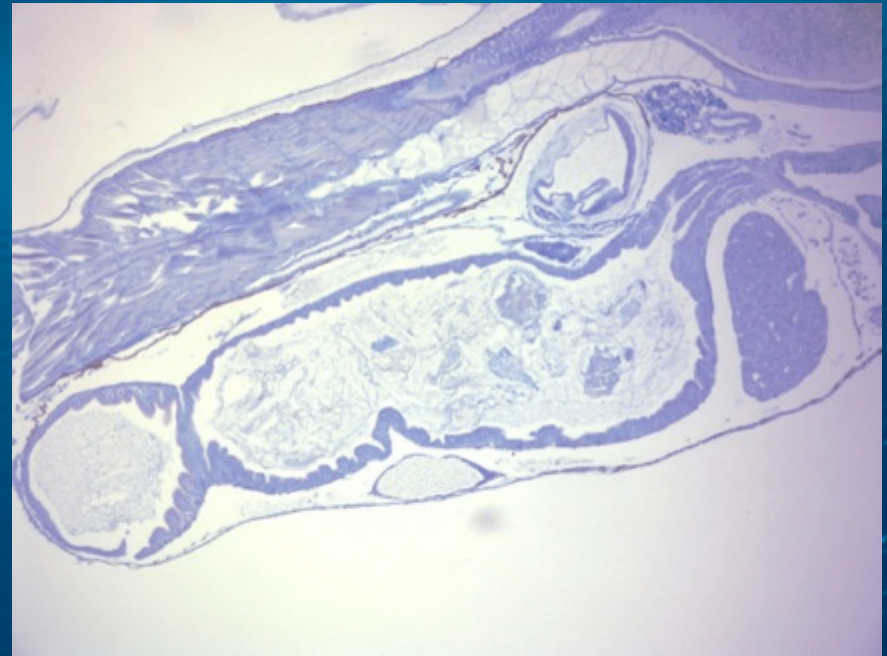


Challenge with *V. logei*

Intestine 24 h
post challenge

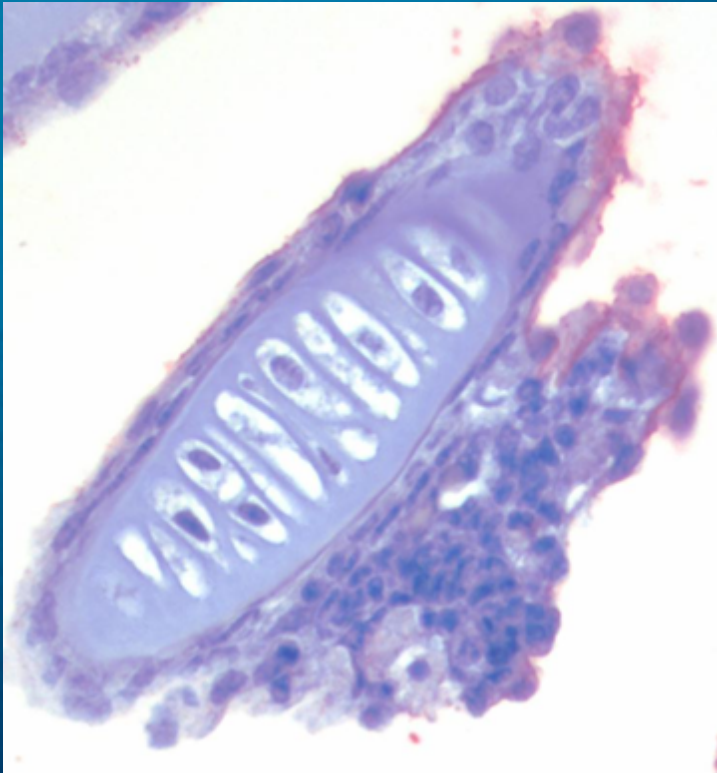


72 h
post challenge

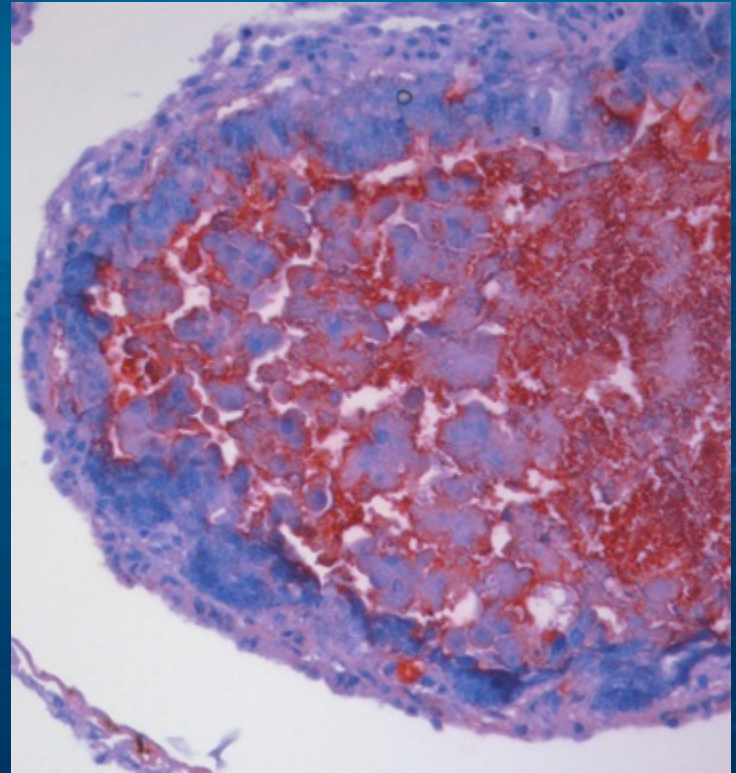


V. splendidus

Gills



Intestine





***Thank you for your
attention!***