



Taihoro Nukurangi

NZ National Institute for Water & Atmospheric Research

Reproduction of hāpuku, *Polyprion oxygeneios*, in New Zealand



***Matthew J. Wylie**, P. Mark Lokman, Abigail Elizur, Jane E. Symonds,
Alvin N. Setiawan

*Present Institute: The New Zealand
Institute for Plant and Food Research



UNIVERSITY
of
OTAGO
Te Whare Wānanga o Ōtago
NEW ZEALAND



University of the
Sunshine Coast

New Zealand Aquaculture

The industry

- grown from small beginnings to a significant primary industry, sustainably producing three flagship products
- a target goal of \$1 billion in sales by 2025
- a need for species diversification and high value products
 - Requires significant research & development



Greenshell Mussel
(*Perna canaliculus*)



Pacific Oysters
(*Crassostrea gigas*)



Chinook Salmon
(*O. tshawytscha*)

Hapuku

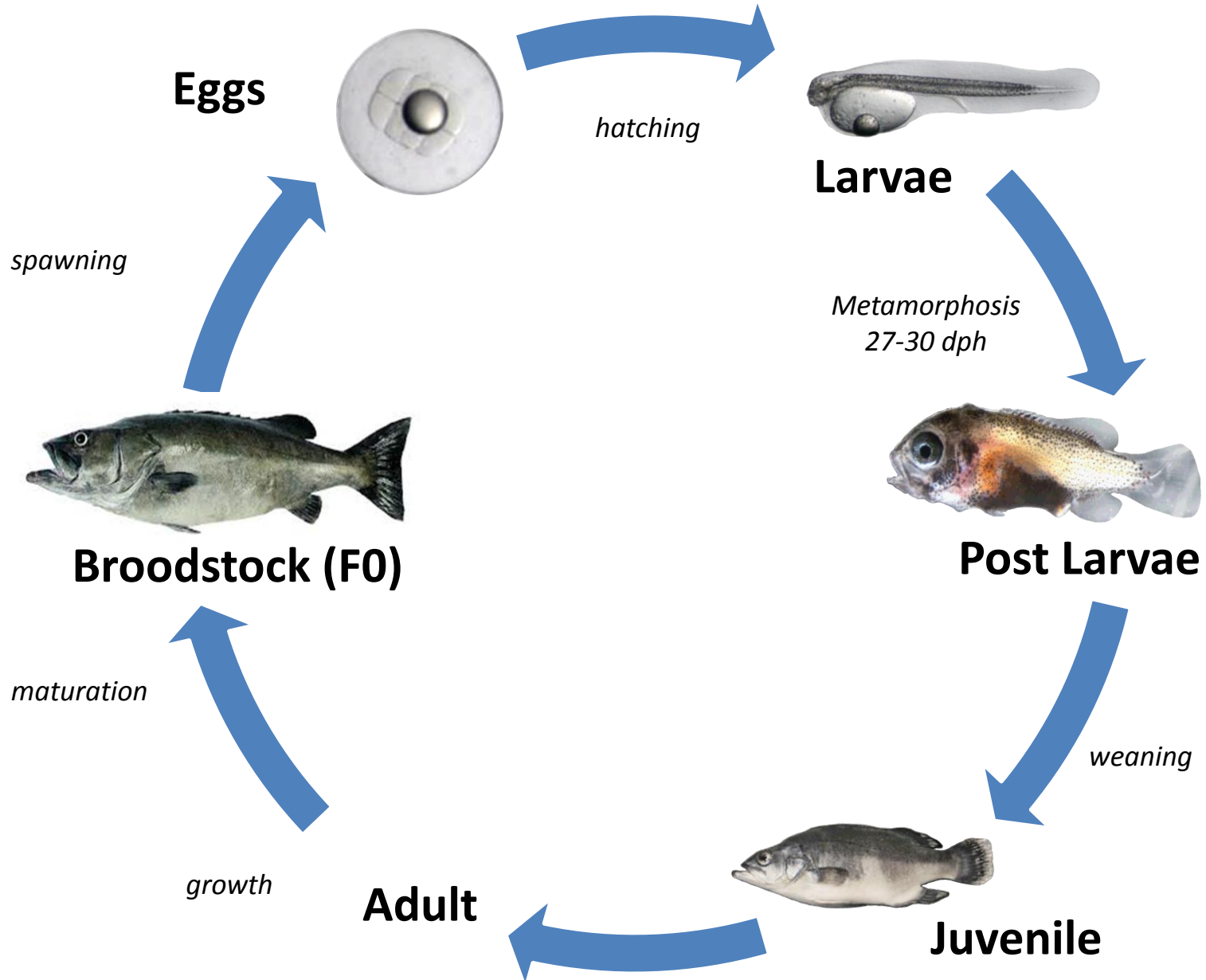
In 2007, NIWA established a broodstock selective breeding programme

- Southern hemisphere species
- Long-lived deep water species
- Sexually mature (10-13 years in wild)
- Grow large
- Growth - very rapid during juvenile stages and up to the first 3 years of their life (~ 4 kg at 2.5 years)
- Recently began to understand the reproductive physiology of this species – due to the availability of captive stocks

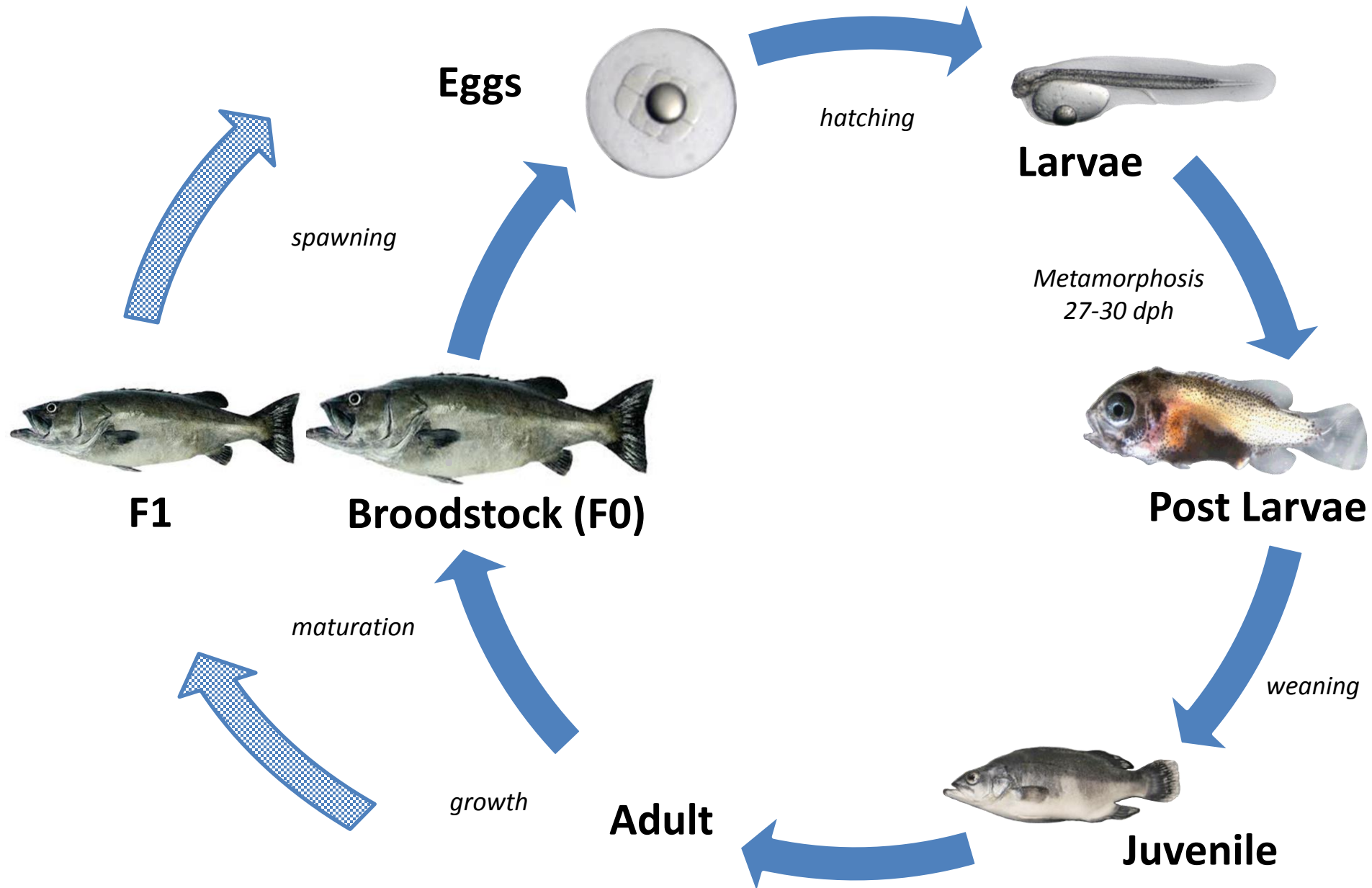


Hāpuku (*Polyprion oxygeneios*)

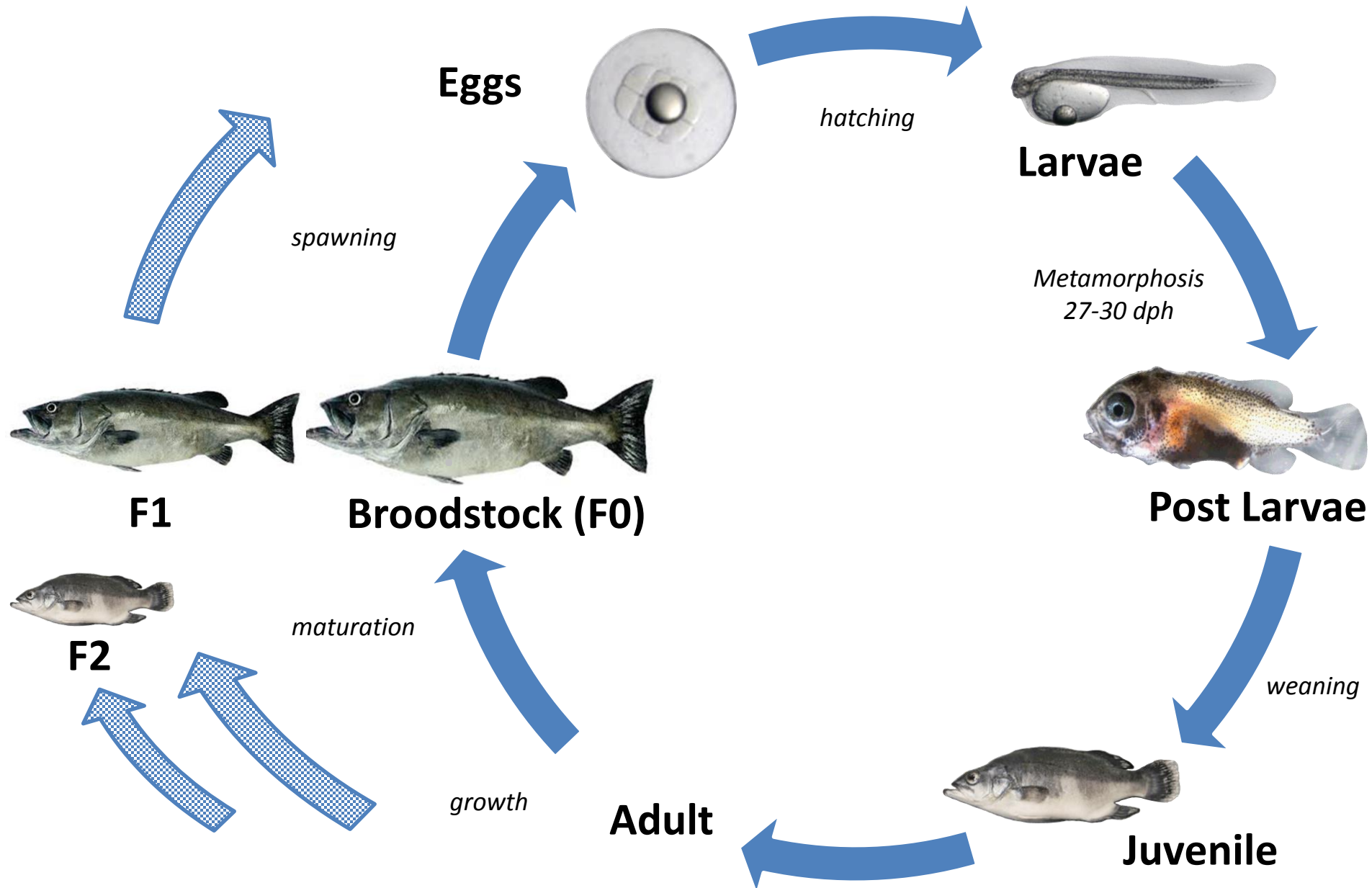
The hapuku production cycle



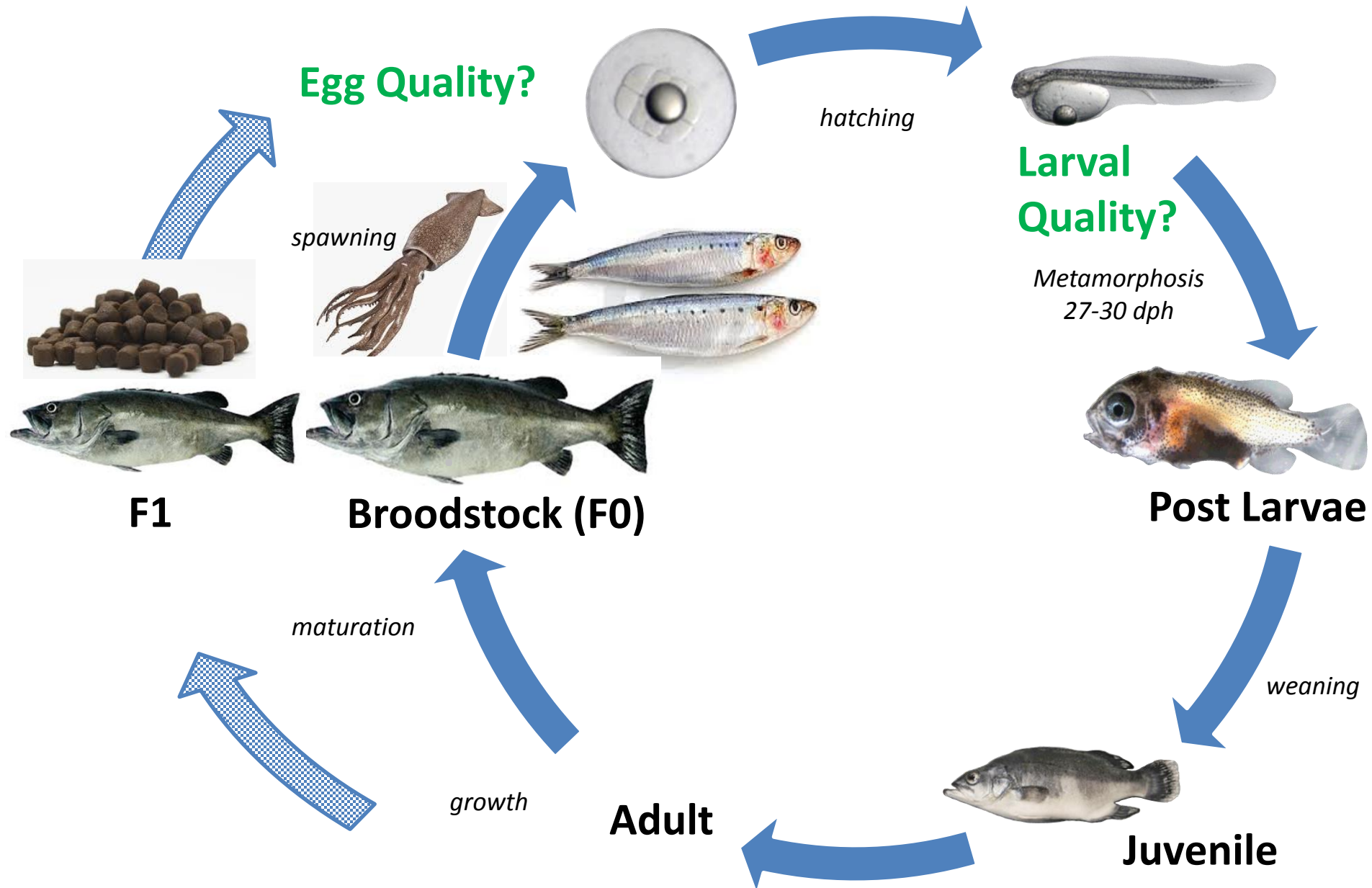
The hapuku production cycle



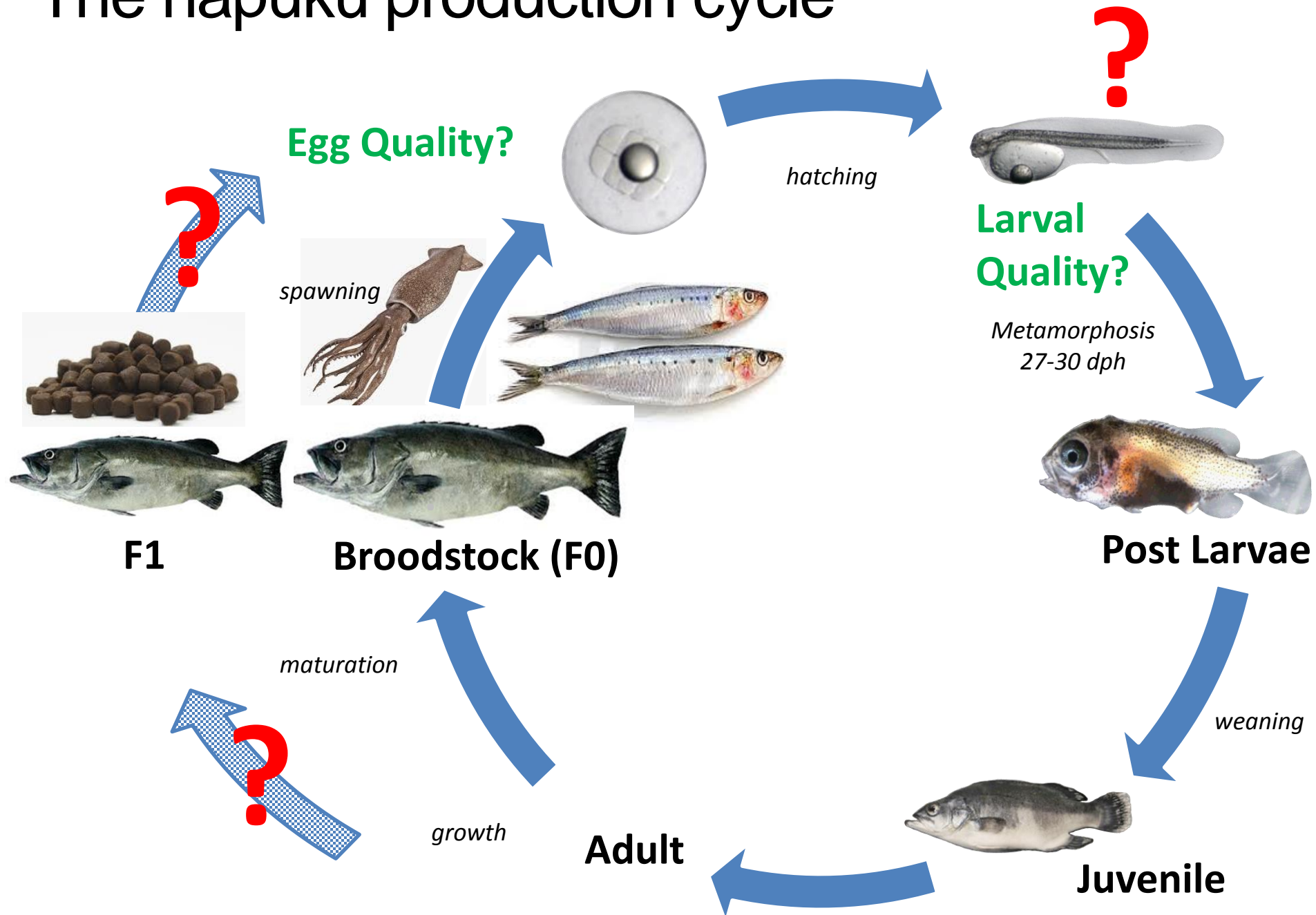
The hapuku production cycle



The hapuku production cycle

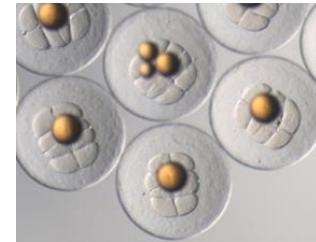
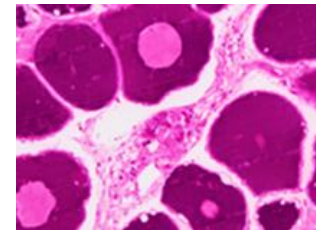


The hapuku production cycle



Core research objectives since 2012

- Examining early oogenesis using next generation sequencing **Wylie et al. *in prep***
- Describing ovarian development of 'first-time spawning' hapuku **Wylie et al. 2018**
- Improving spawning of F1 hapuku with hormonal manipulations **Wylie et al. *in prep***
- Reducing generation intervals (puberty advancement) **Wylie et al. 2018**
- Life-cycle closure **Wylie et al. *in prep***
- Enhancing larval survival



Core research objectives since 2012

- ❑ Examining early oogenesis using next generation sequencing **Wylie et al. *in prep***

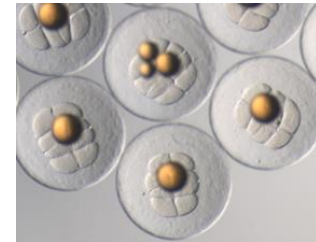
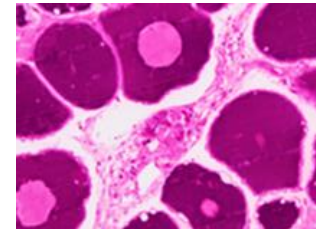
- ❑ Describing ovarian development of 'first-time spawning' hapuku **Wylie et al. 2018**

- ❑ Improving spawning of F1 hapuku with hormonal manipulations **Wylie et al. *in prep***

- ❑ Reducing generation intervals (puberty advancement) **Wylie et al. 2018**

- ❑ Life-cycle closure **Wylie et al. *in prep***

- ❑ Enhancing larval survival



Study sites

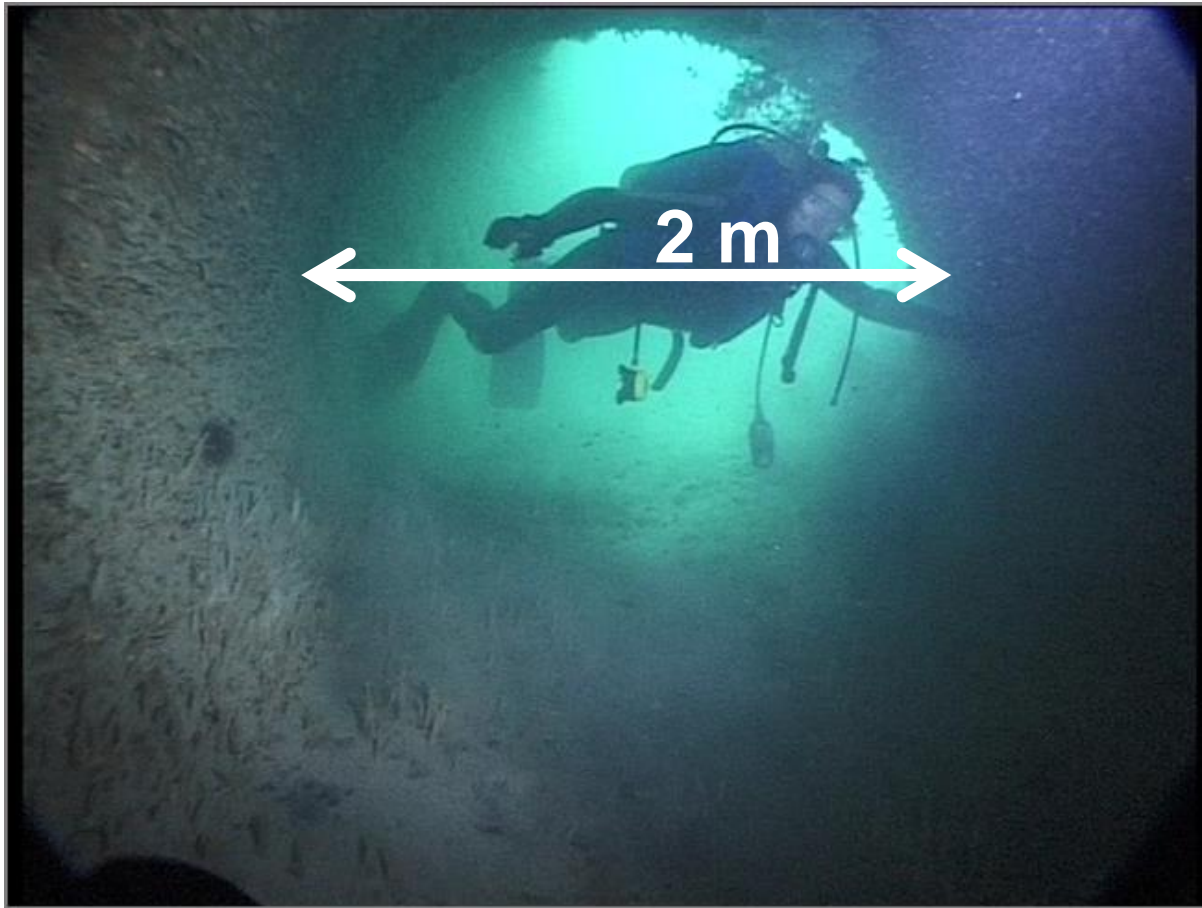


Live animal work conducted at NIWA's Northland Marine Science Centre

Northland Marine Research Centre

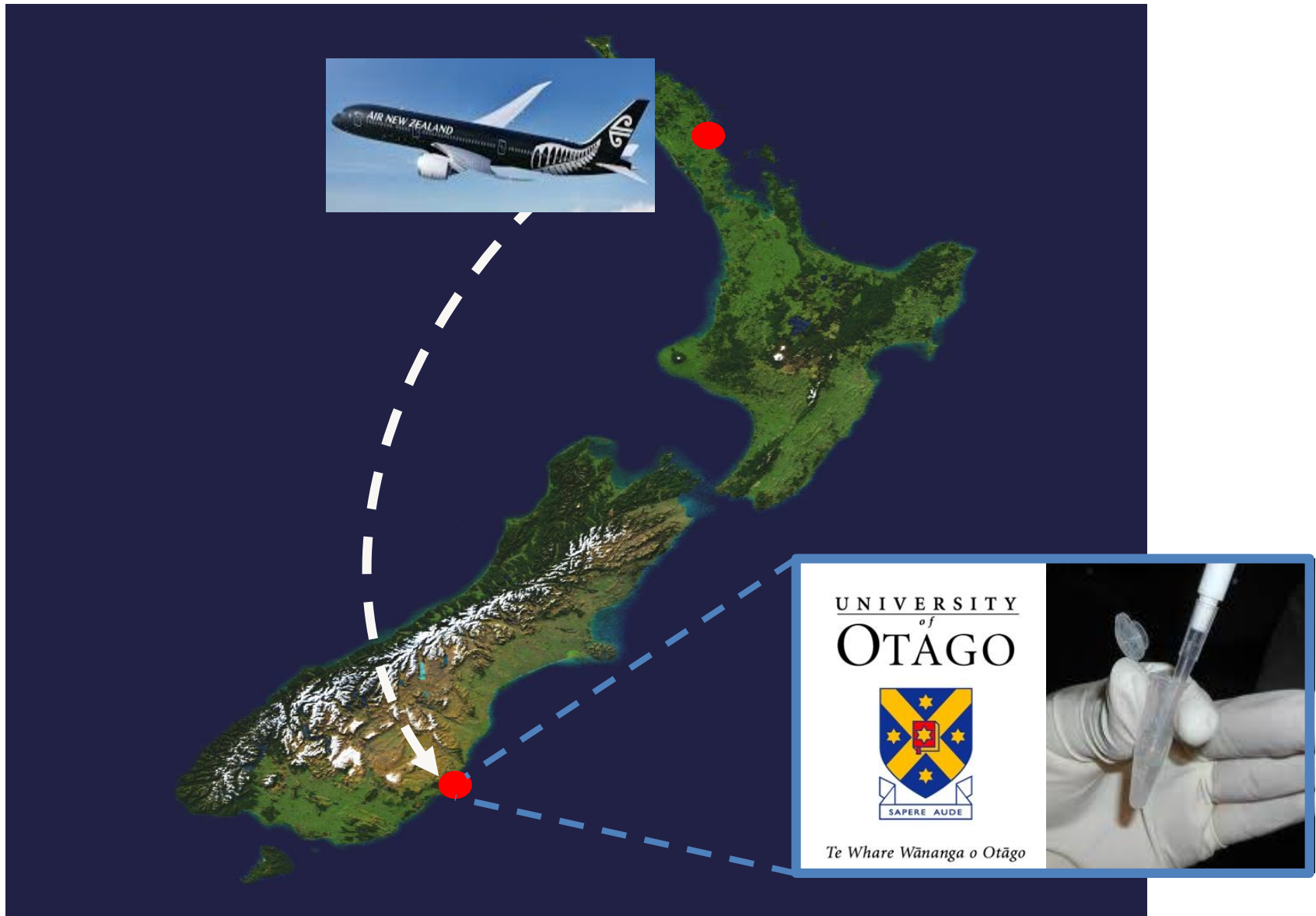


- Land based marine aquaculture facility bordering the Pacific Ocean
- Makes use of power station infrastructure



Water Source:

Existing power station intakes



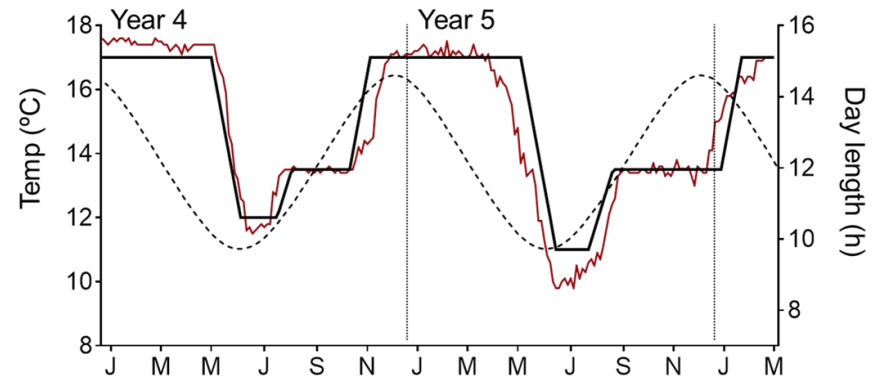
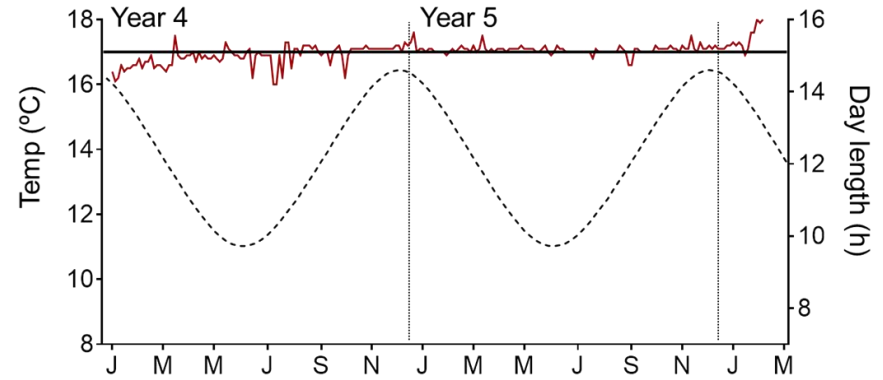
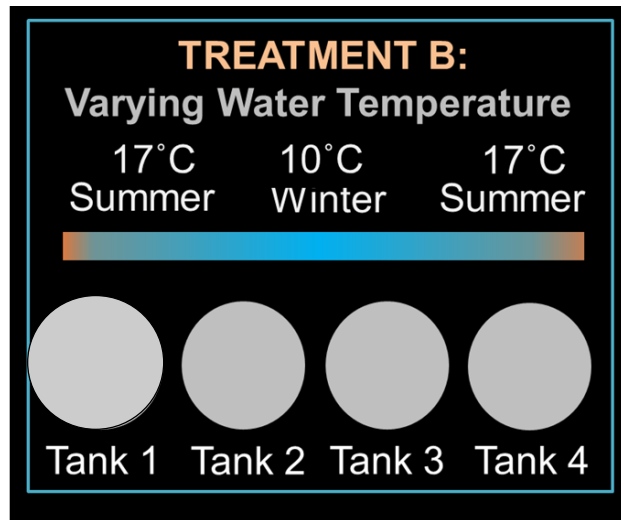
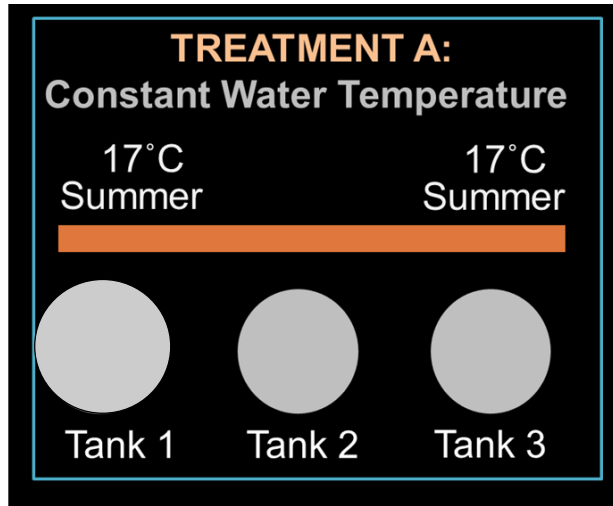
Sample analysis conducted at the Department of Zoology

I. Oogenesis in hapuku:

Aims:

- Assess the effect of thermal regime on ovarian development of first-time spawning F1
 - Improve husbandry conditions
 - Provide a basis for photoperiod manipulations
 - Determine the age at maturity of F1

Oogenesis in first-time spawning hapuku:



- Target temp
- Actual temp
- - - Simulated photoperiod

Oogenesis in first-time spawning hapuku:

TREATMENT A:

Constant Water Temperature

17°C
Summer

17°C
Summer



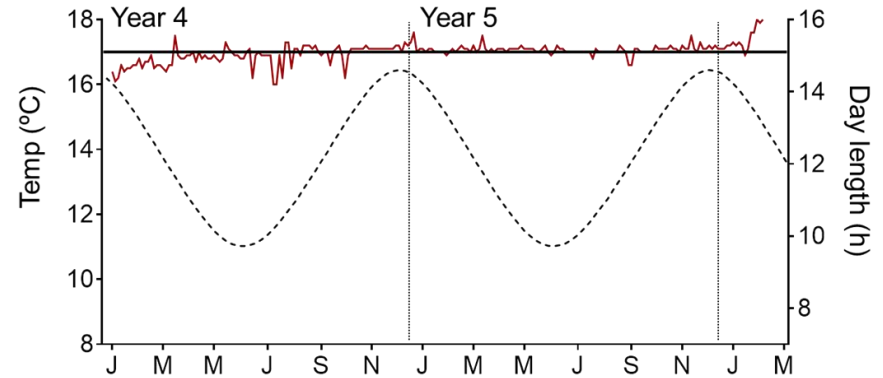
Tank 1



Tank 2



Tank 3



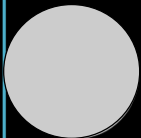
TREATMENT B:

Varying Water Temperature

17°C
Summer

10°C
Winter

17°C
Summer



Tank 1



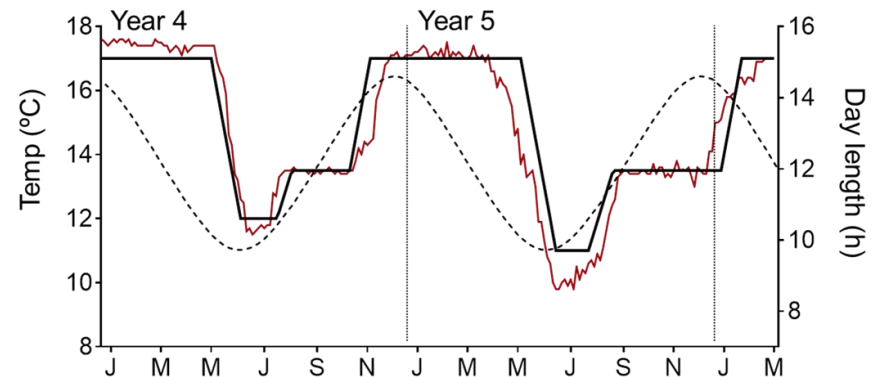
Tank 2



Tank 3

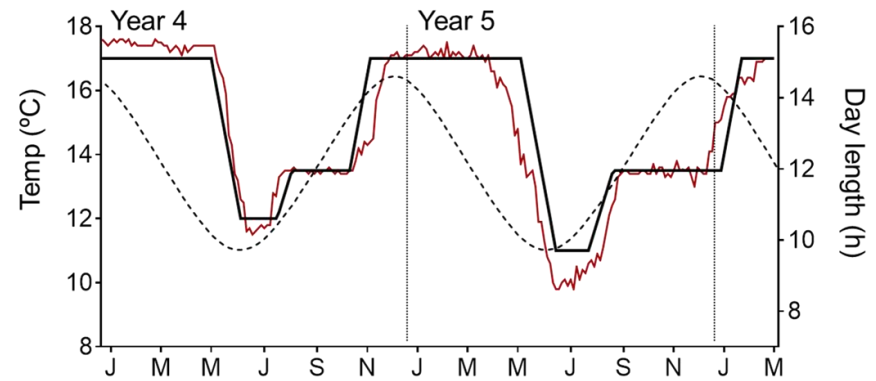
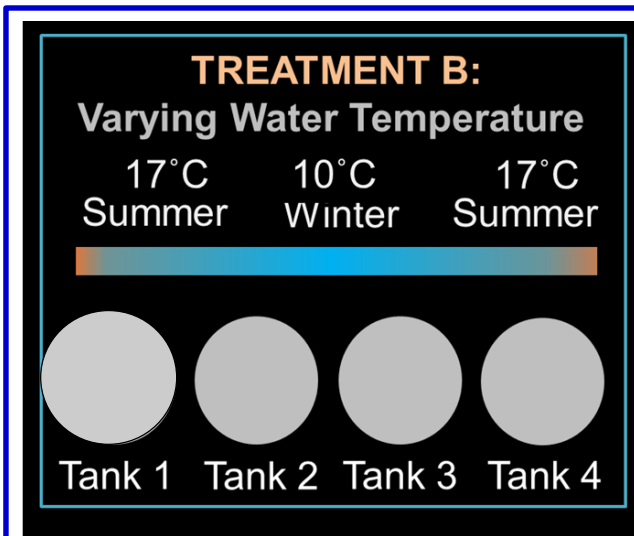
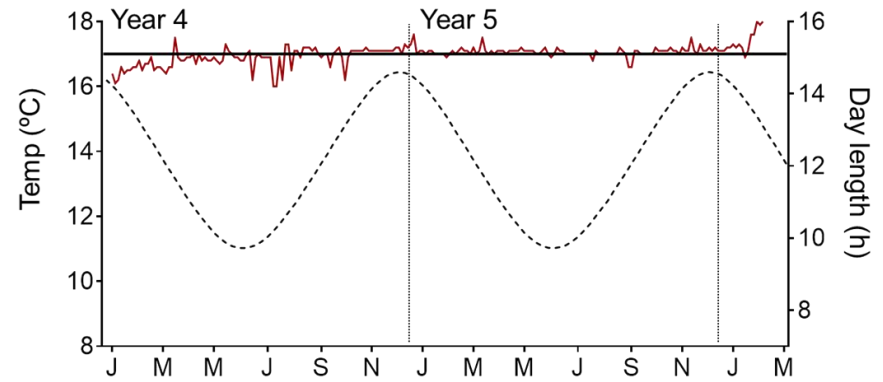
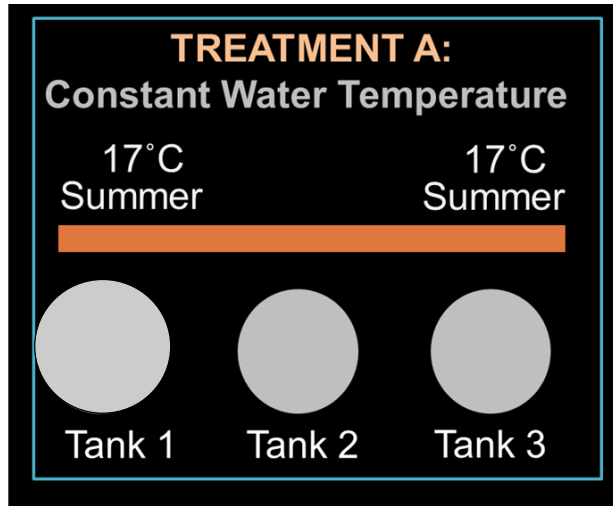


Tank 4



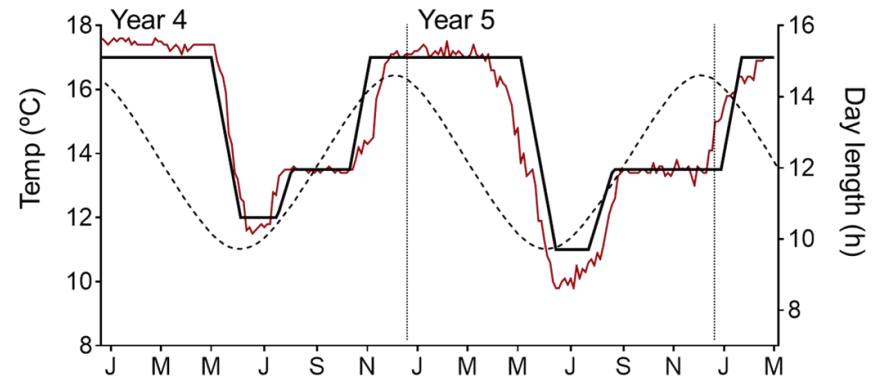
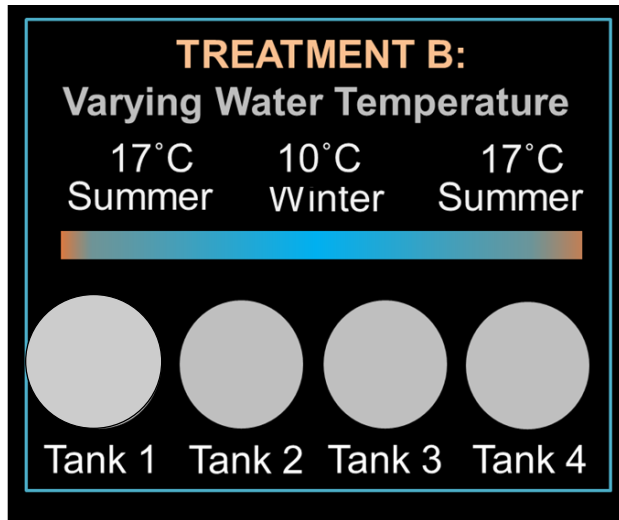
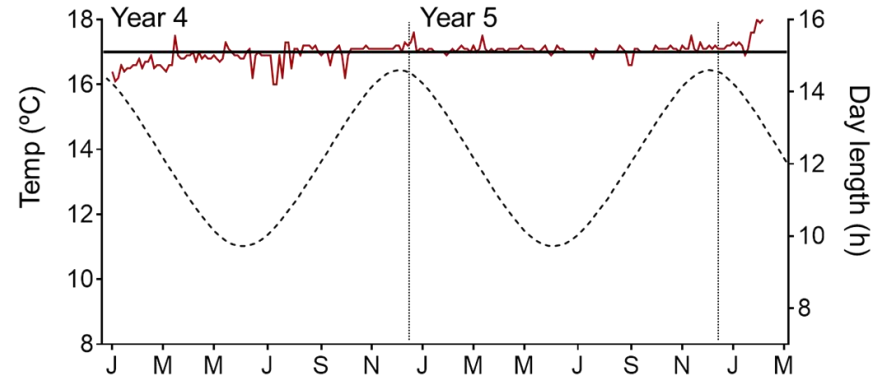
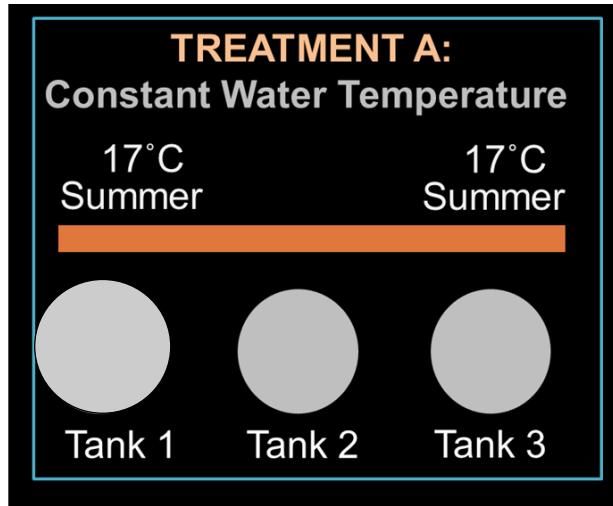
- Target temp
- Actual temp
- - - Simulated photoperiod

Oogenesis in first-time spawning hapuku:



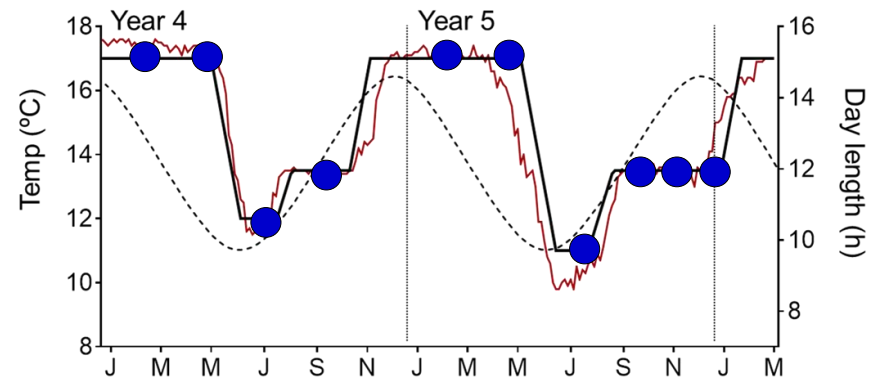
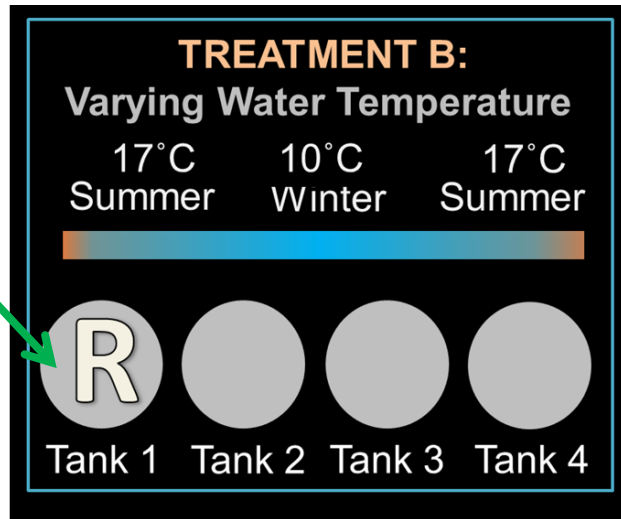
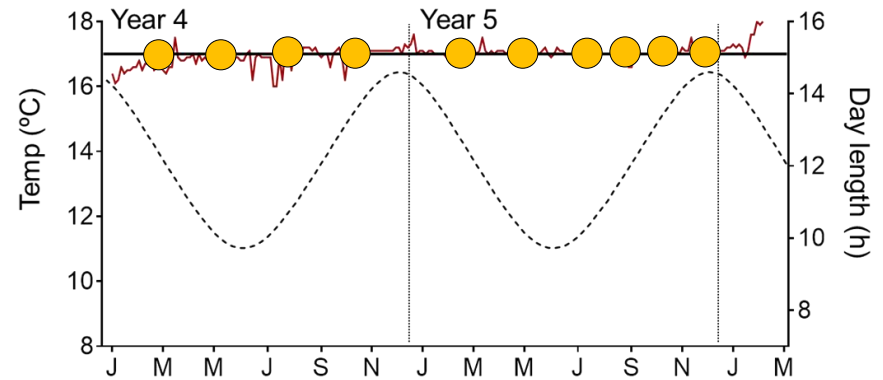
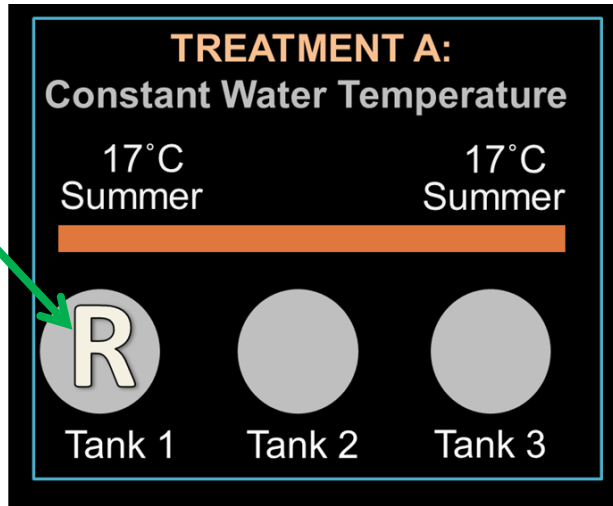
- Target temp
- Actual temp
- - - Simulated photoperiod

Oogenesis in first-time spawning hapuku:

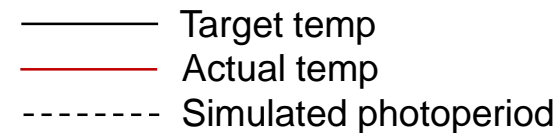


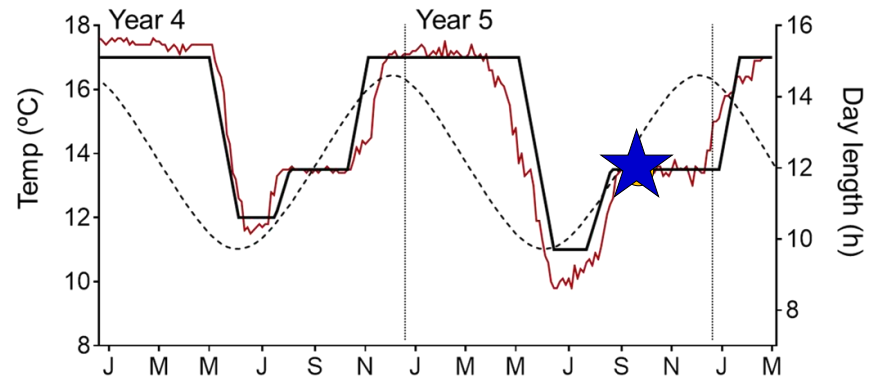
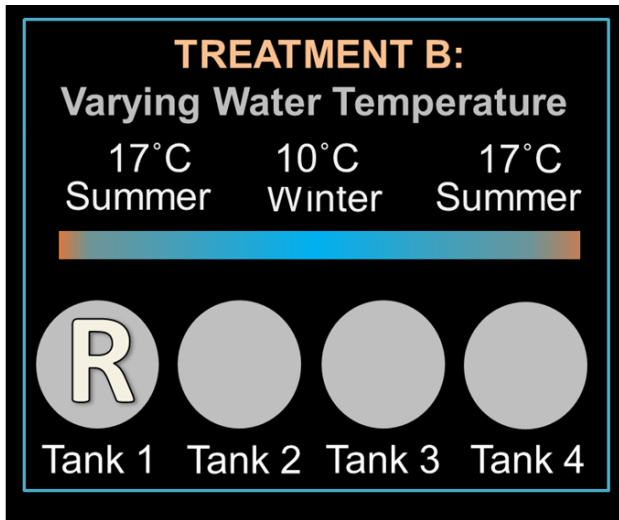
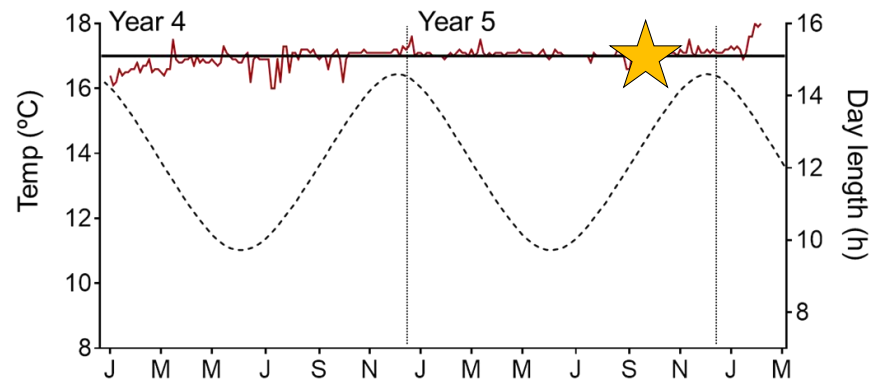
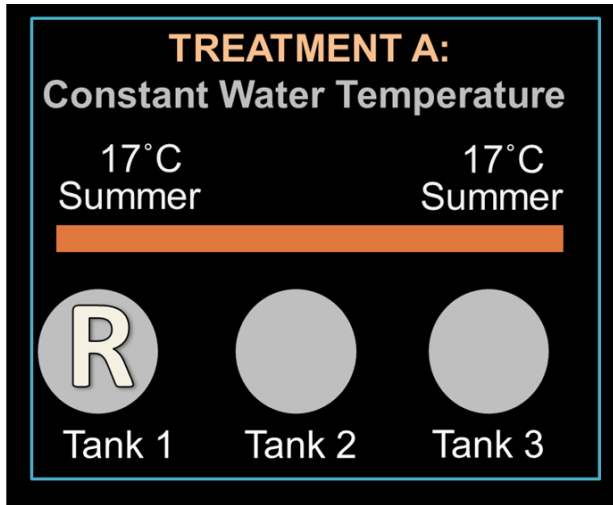
- Photoperiods kept consistent
- Approx. 20 fish per tank (male and female; 4 yr old at start)

— Target temp
— Actual temp
- - - Simulated photoperiod



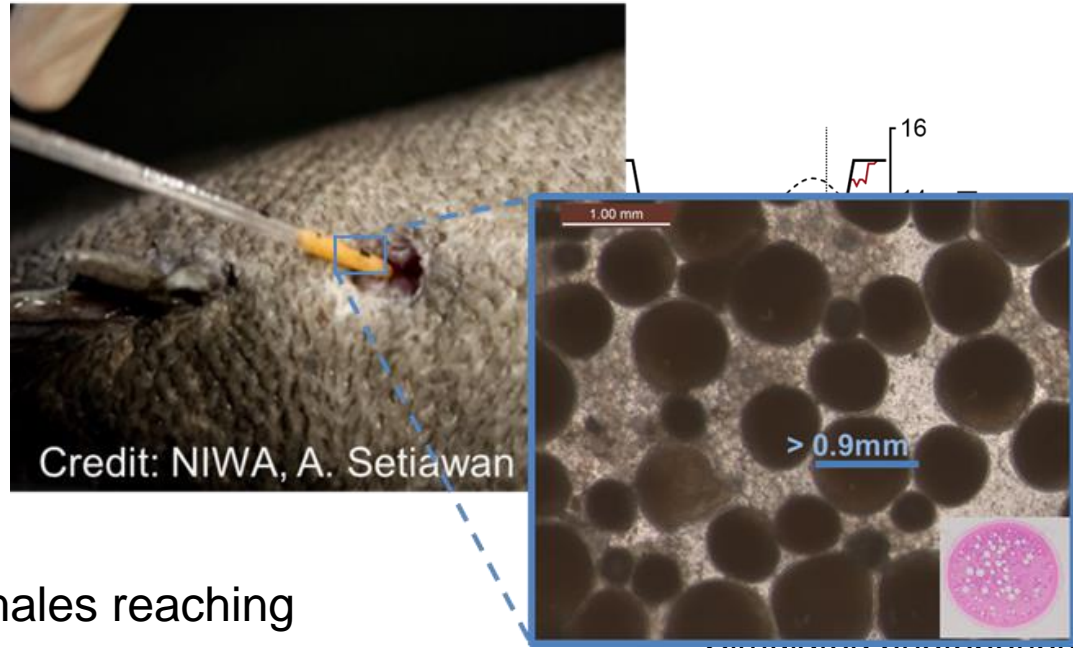
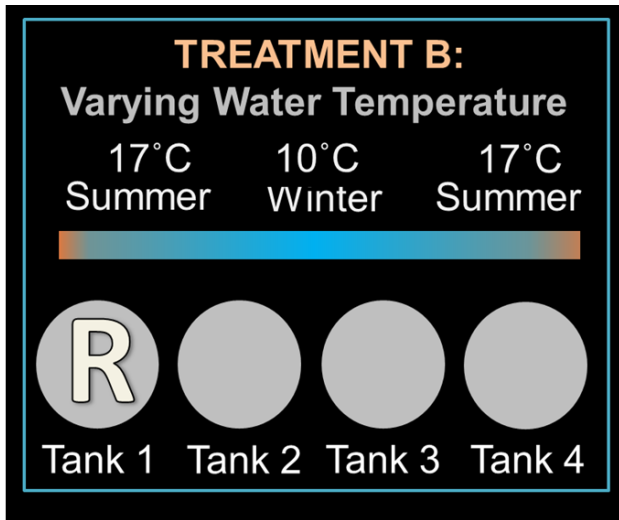
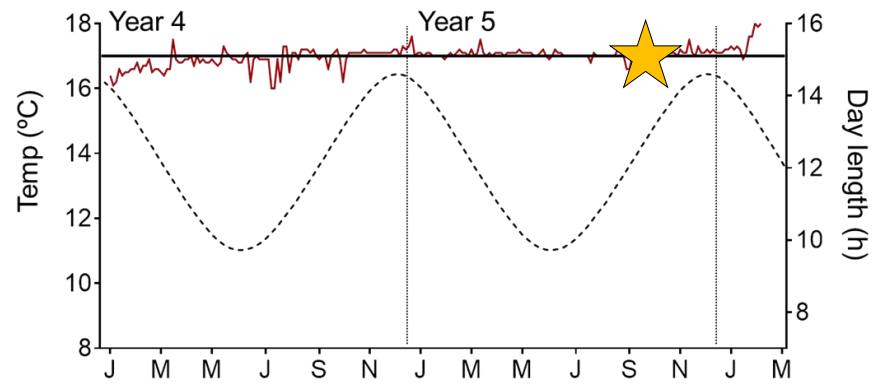
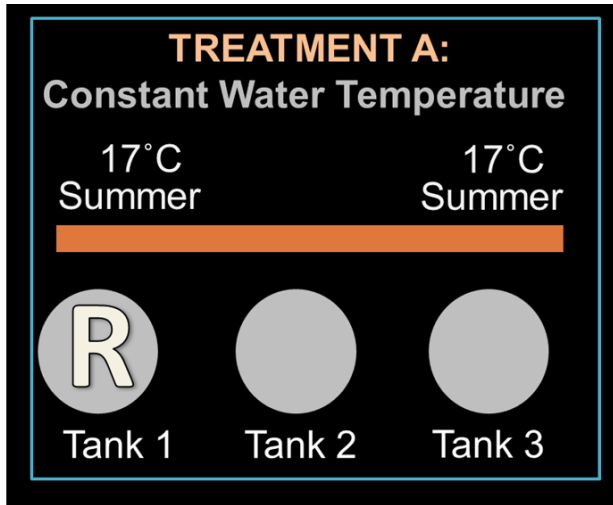
- A gonad biopsy and blood sample - repeatedly sampled every 8 weeks from fish in one tank from each treatment





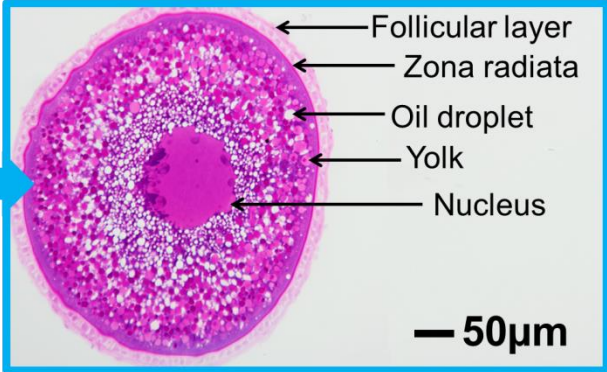
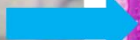
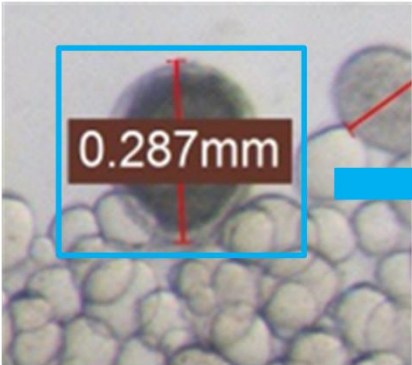
— Target temp
— Actual temp
- - - Simulated photoperiod

- Prior to the predicted spawning season all fish were biopsied in October



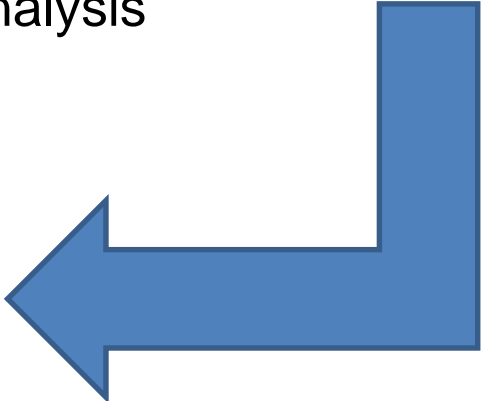
- Determine the percentage of females reaching spawning condition

Sample collection



Ovarian biopsies collected
~ every 8 weeks for two
years

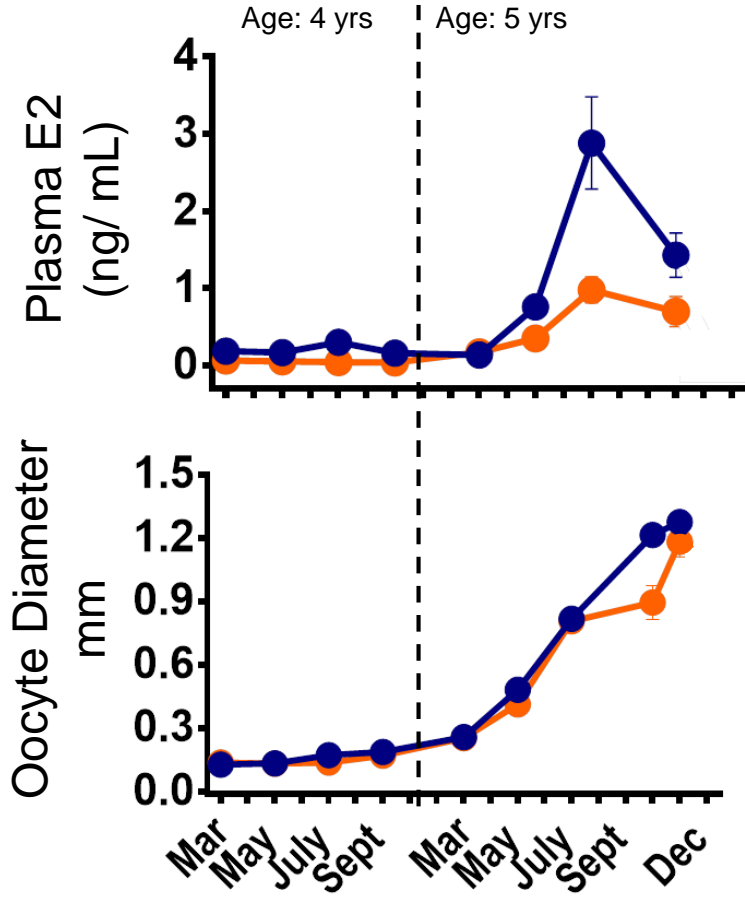
Oocyte diameters measured by light
microscopy. Samples processed for
histological analysis



RT-PCR of genes linked to oogenesis
(gonadotropin receptors: *fshr* and *lhr*)

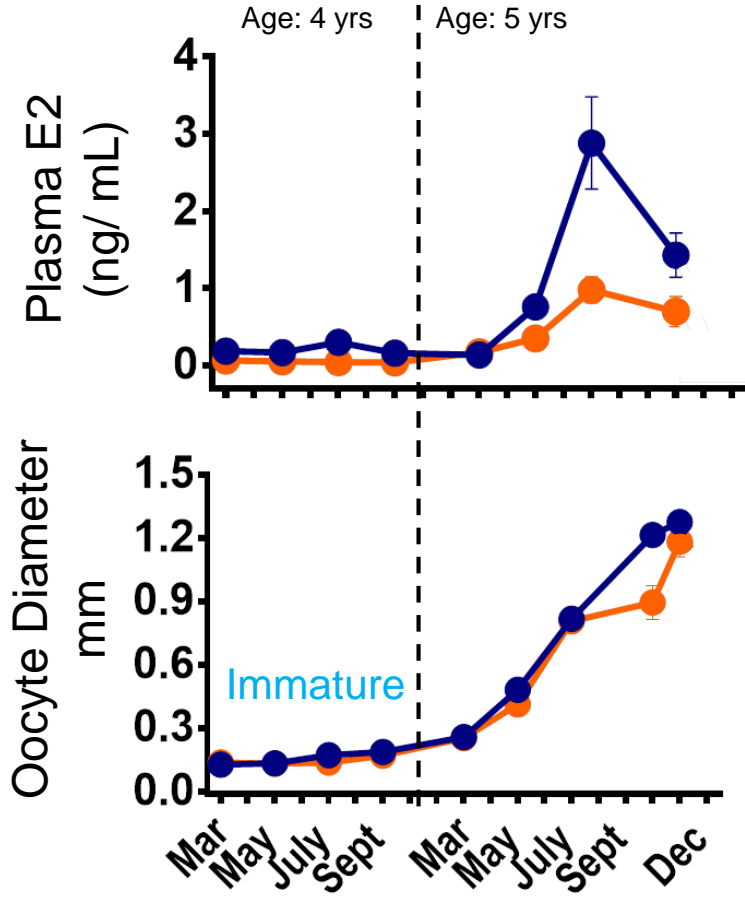
Oogenesis and the reproductive cycle

● Constant Temperature ● Varying Temperature



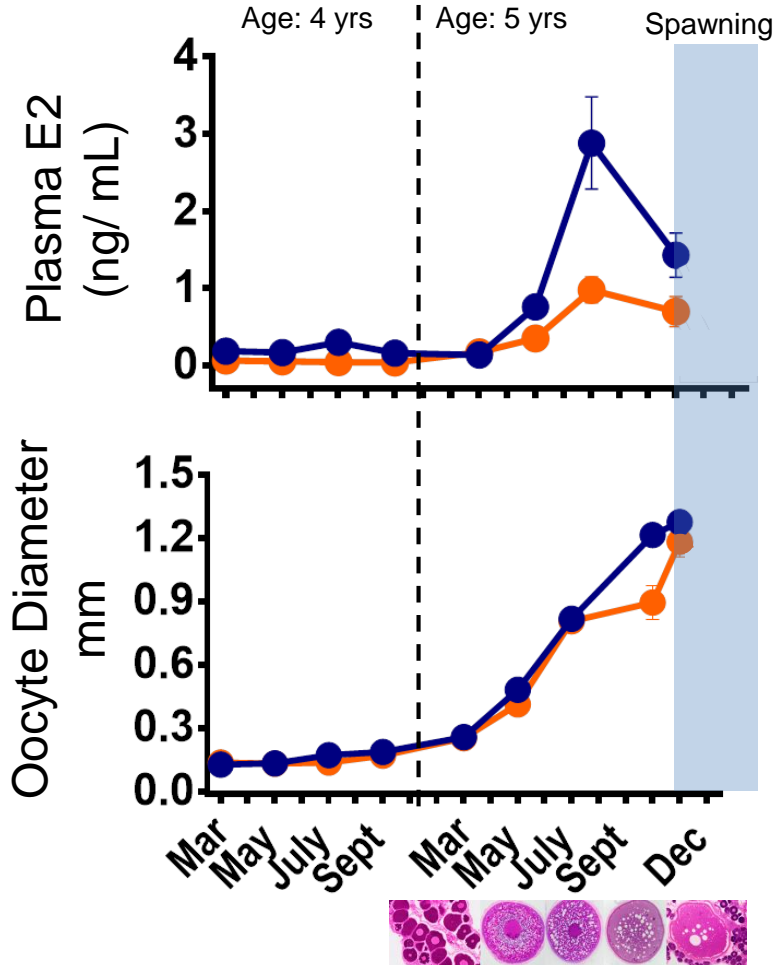
Oogenesis and the reproductive cycle

● Constant Temperature ● Varying Temperature



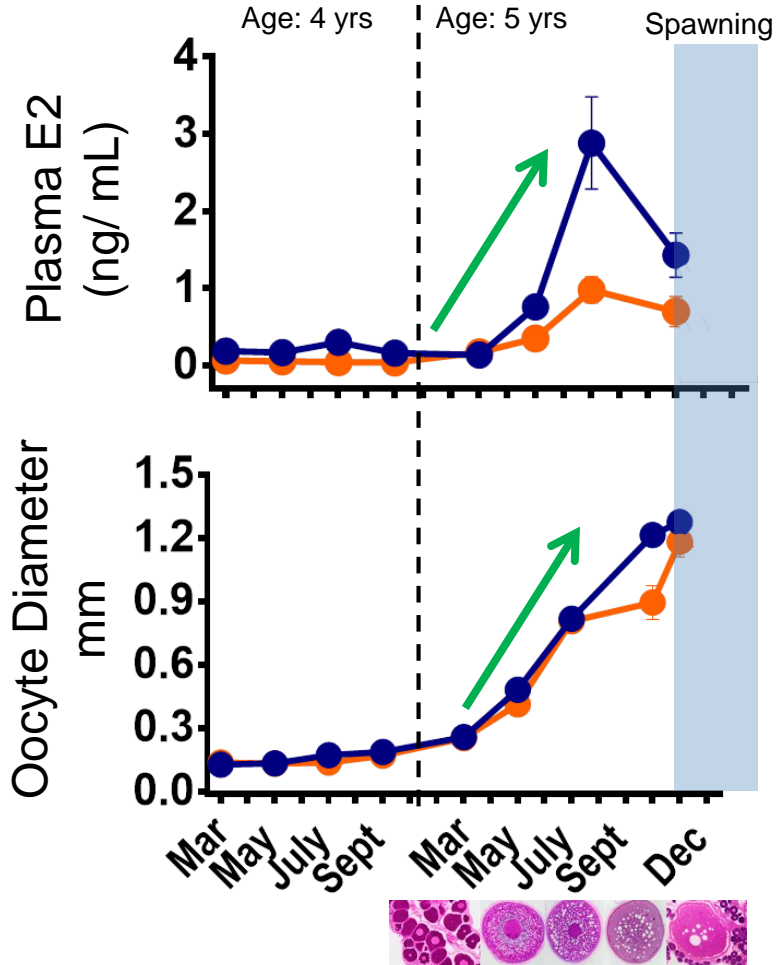
Oogenesis and the reproductive cycle

● Constant Temperature ● Varying Temperature



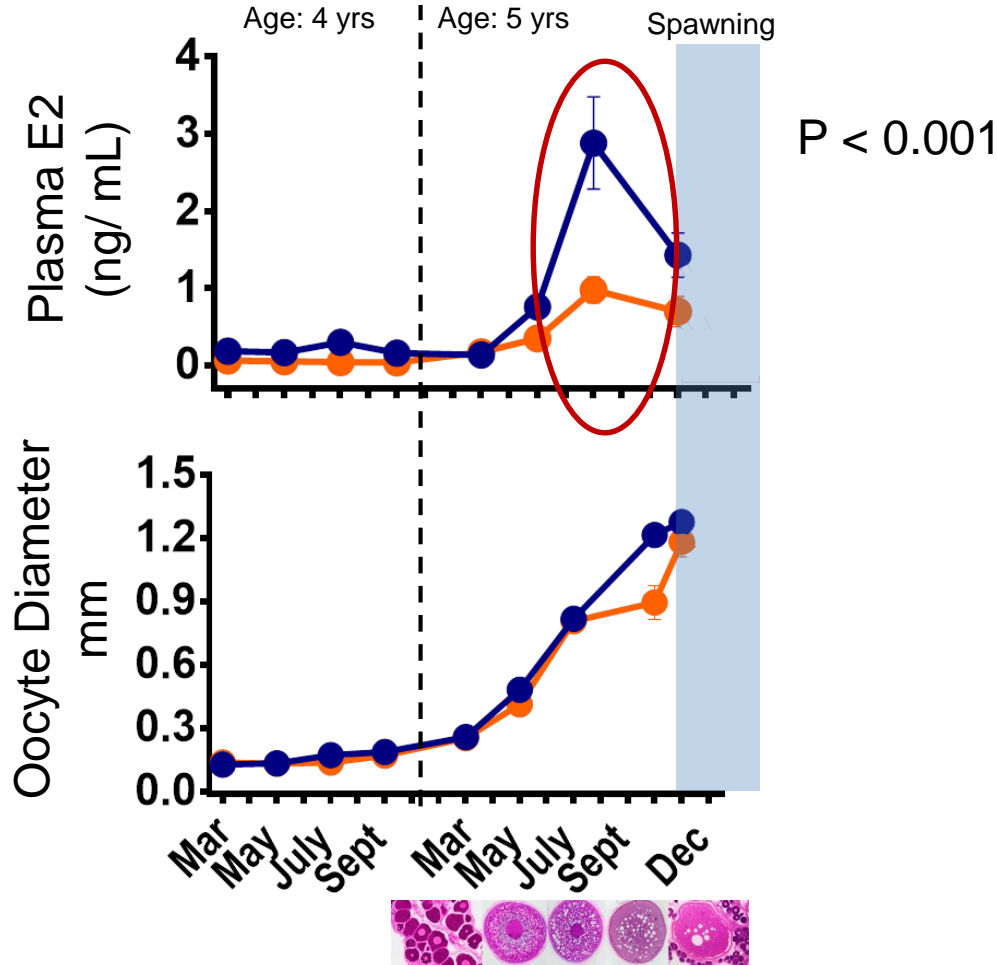
Oogenesis and the reproductive cycle

● Constant Temperature ● Varying Temperature

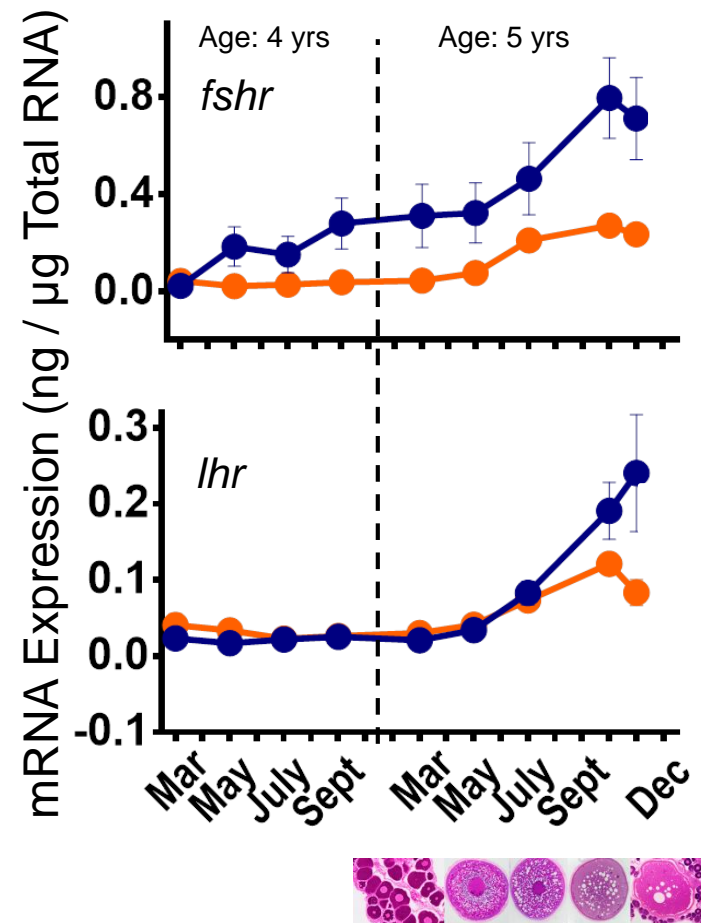
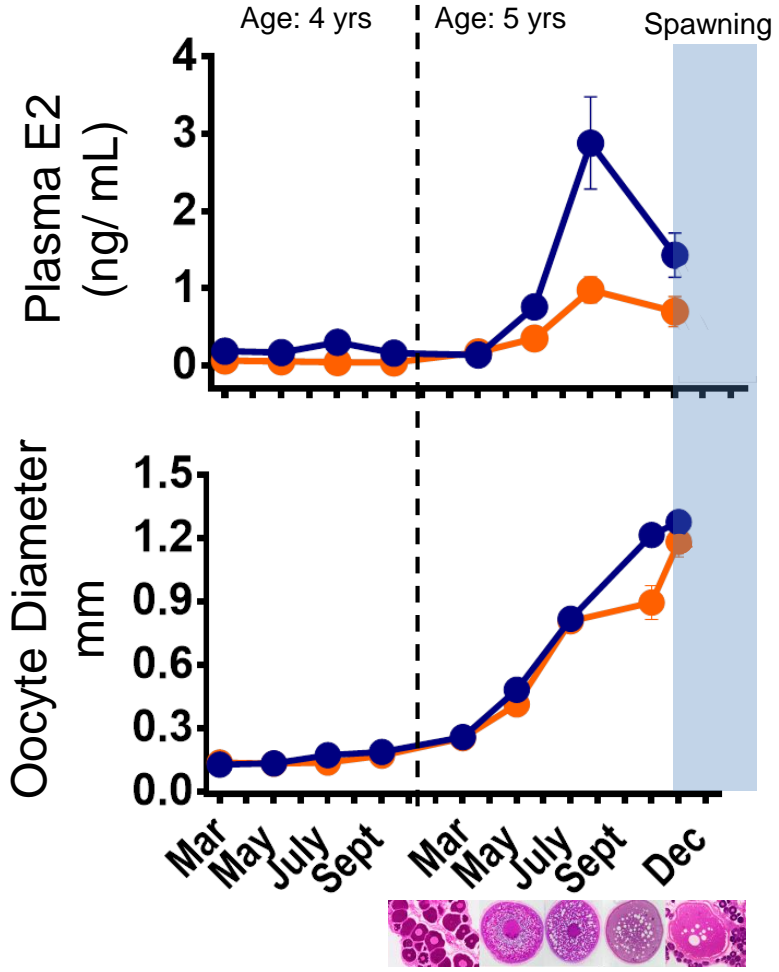


Oogenesis and the reproductive cycle

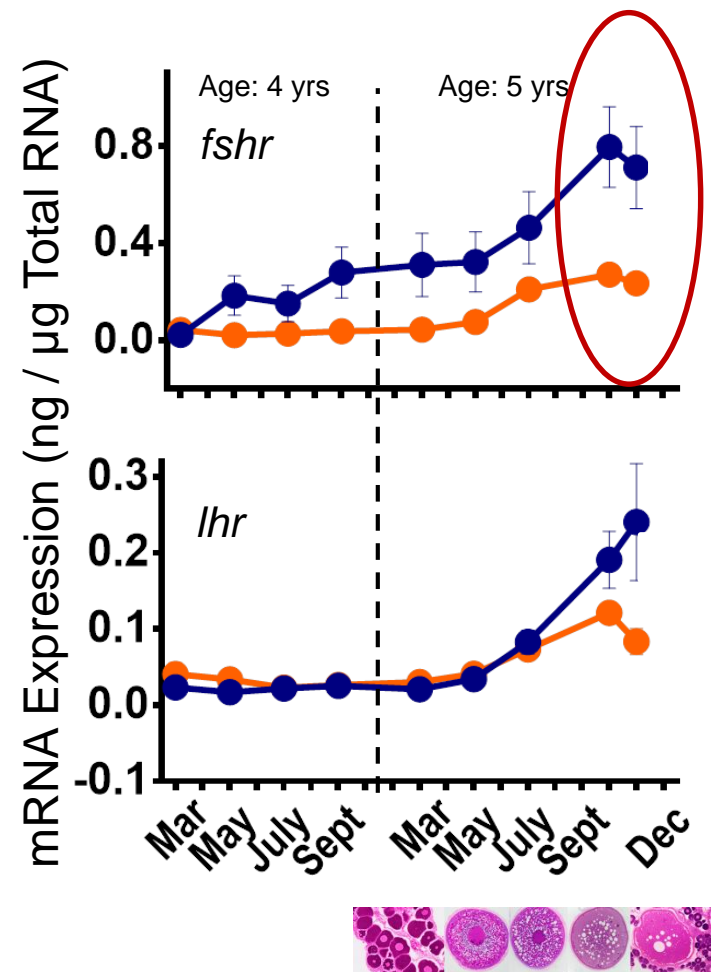
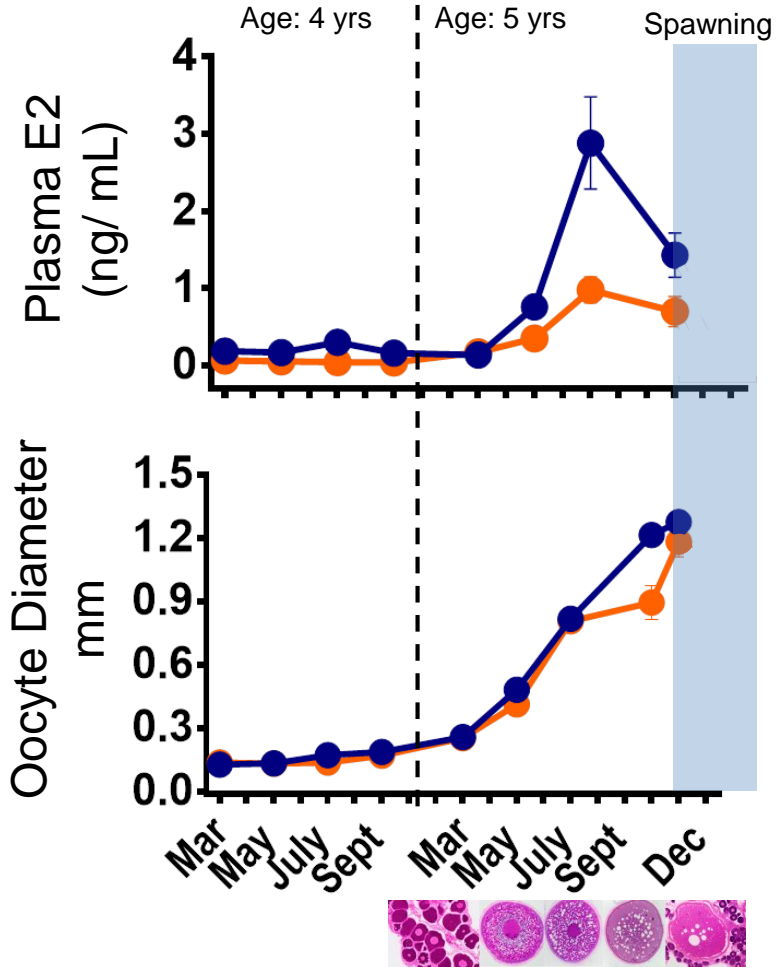
● Constant Temperature ● Varying Temperature



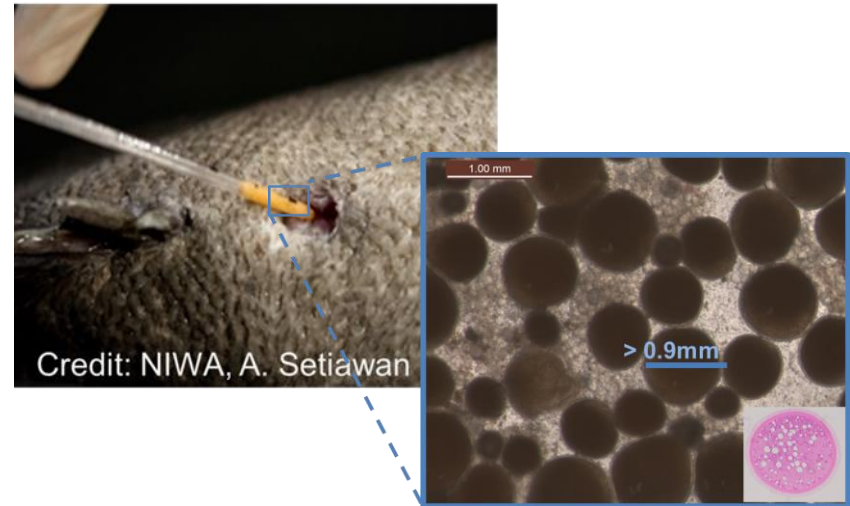
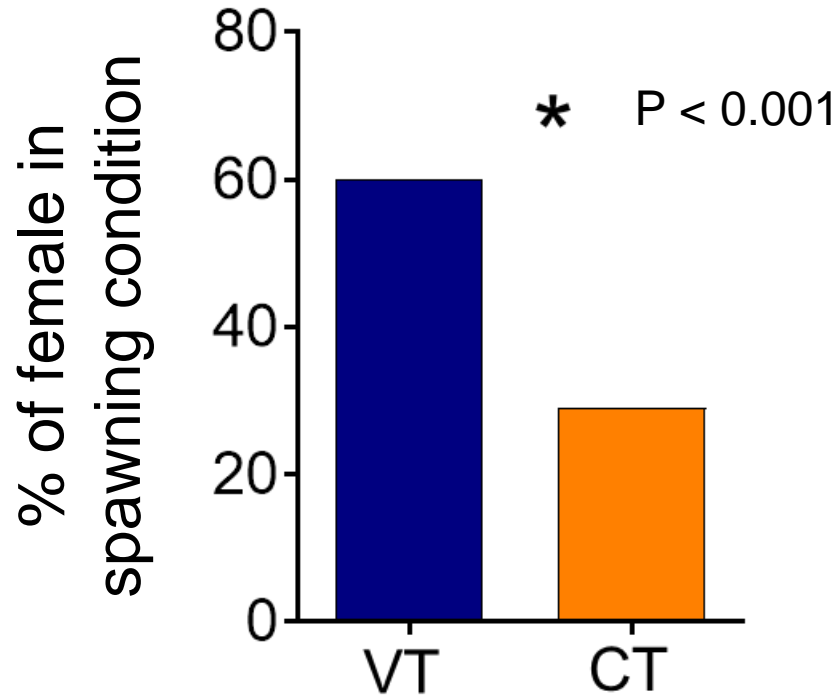
Oogenesis and the reproductive cycle



Oogenesis and the reproductive cycle

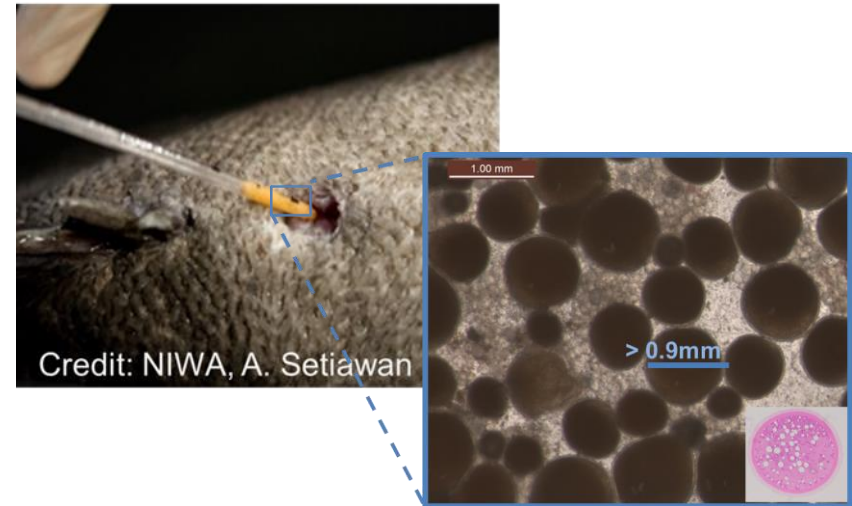
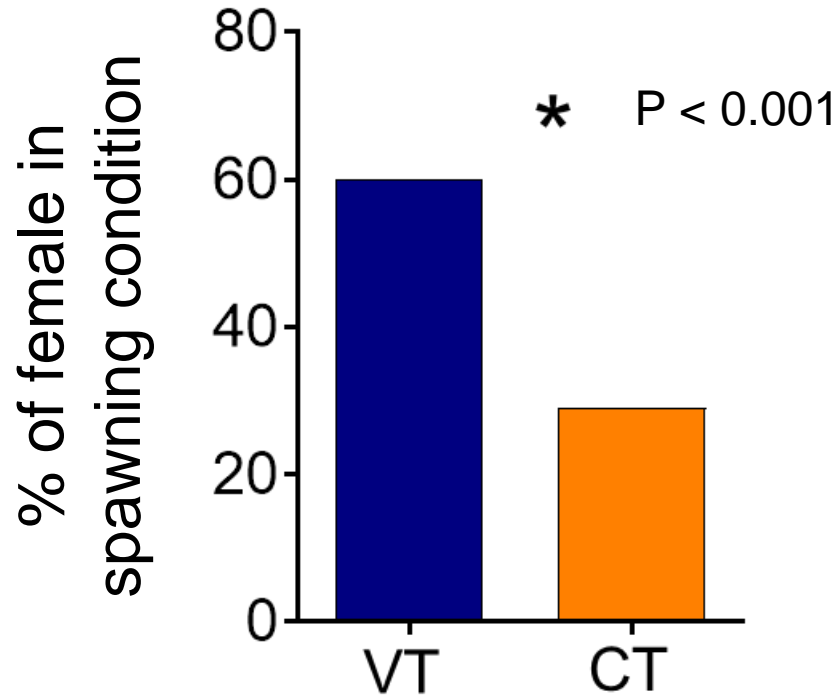


Percentage reaching spawning condition



- Fewer (~50 % less) females reached pre-spawning condition in tanks maintained at a constant 17 degrees

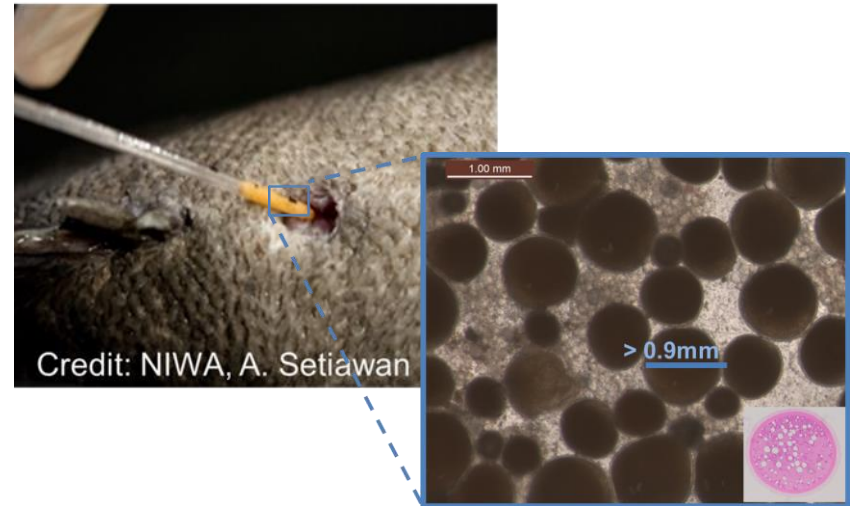
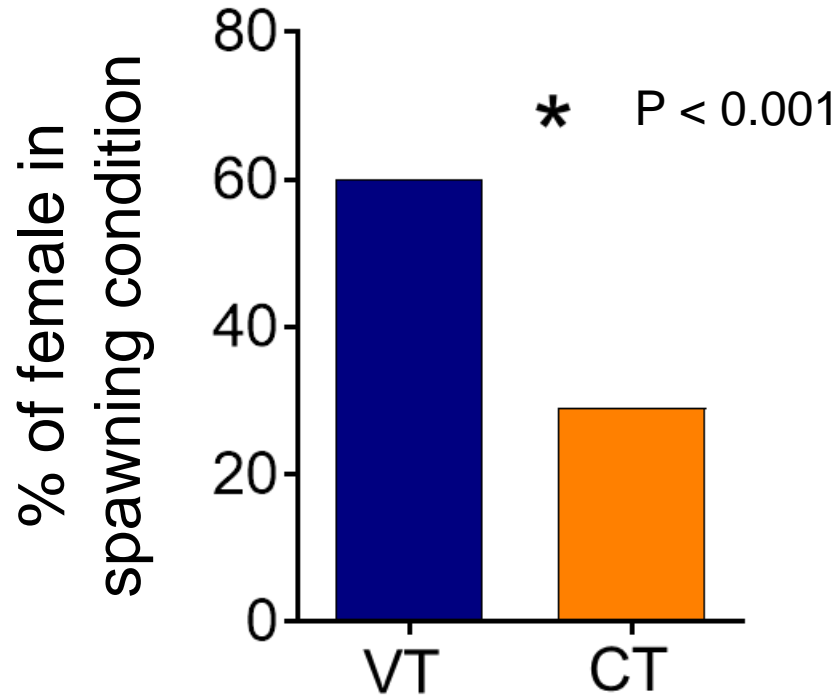
Percentage reaching spawning condition



- Fewer (~50 % less) females reached pre-spawning condition in tanks maintained at a constant 17 degrees

Exposure to a decreased temperature is important for appropriate progression of oogenesis

Percentage reaching spawning condition



- Fewer (~50 % less) females reached pre-spawning condition in tanks maintained at a constant 17 degrees

F1 broodstock also failed to spawn naturally

Progress to date:

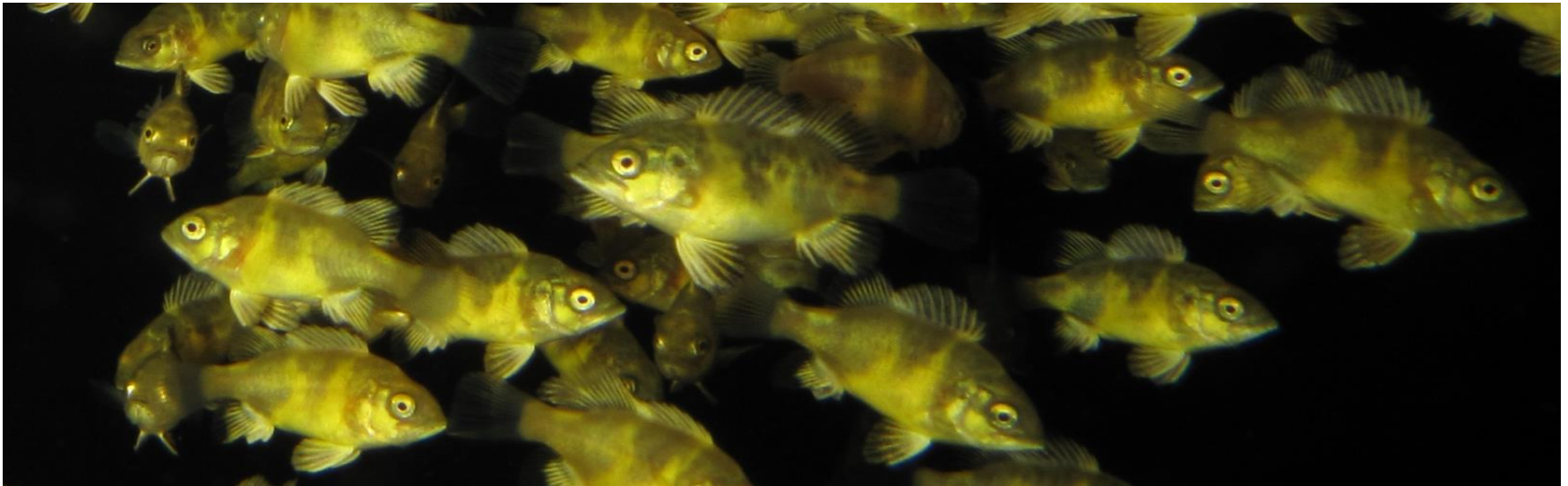
- ❑ Ovarian development of 'first-time spawning' hapuku has been described
 - ❑ Cool water temperatures are important for the appropriate progression of oogenesis
 - ❑ Sexual maturity occurs at 5 years in captivity
 - ❑ Life cycle completed in 2013

- ❑ Hormonal manipulations induced spawning
 - ❑ Some F1 broodstock now spawn naturally at 7 years old (handling-dependent)

- ❑ Larval survival (F2) improved to 7.2 % at 28 dph
 - ❑ Continued F2 survival through to fingerling at 5.5%
 - ❑ Improvements based on a many factors
 - ❑ Changes to broodstock rearing (i.e density, reduced handling)
 - ❑ Improved larval rearing conditions (i.e hygiene)

Concluding remarks:

- ❑ **Captive-bred hāpuku are a viable and sustainable source of high quality gametes with fingerling survival exceeding that of wild-caught broodstock**
- ❑ **Reliable production levels are not achieved until 7 years of age**
- ❑ **Minimal handling and focus on early growth may hasten reproductive competence**



Acknowledgements



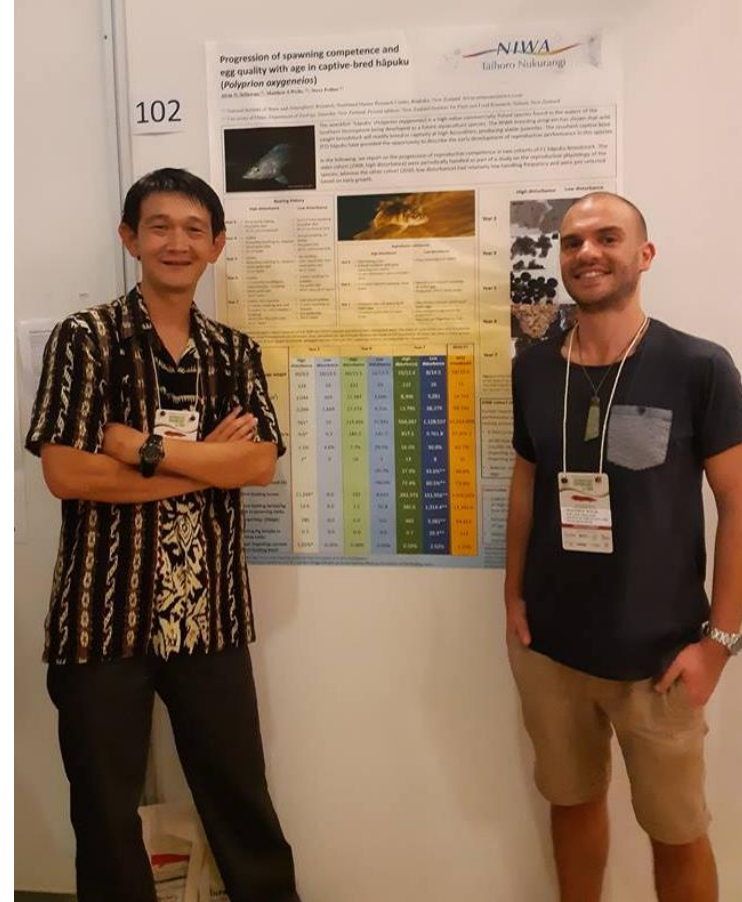
Assoc. Prof.
Mark Lokman



Dr. Jane
Symonds



Prof. Abigail
Elizur



Dr. Alvin
Setiawan



**Technical and Management Staff at NIWA
Northland Marine Research Centre**

www.niwa.co.nz



Questions?

