



Hatchery production of the greater amberjack (*Seriola Dumerili*)

N. Papandroulakis, Tsalafouta, P. Anastasiadis, I. E. Papadakis, N. Mitrizakis (HCMR)
C.M. Hernández Cruz, A. La Barbera, H. Fernández-Palacios Barber, M. Izquierdo (FCPCT)
S. Jerez, M. Virginia Martín (IEO)
J. Perez, C. Rodríguez (ULL)
G. Iakovopoulos (GMF S.A.)



Hellenic Centre for Marine Research

Institute of Marine Biology, Biotechnology & Aquaculture



Fundación Parque Científico Tecnológico
Universidad de Las Palmas de Gran Canaria



INSTITUTO
ESPAÑOL DE
OCEANOGRAFÍA



Universidad de La Laguna



GALAXIDI
Marine Farm S.A.

- Data on applicable rearing conditions in captivity is scarce
 - particularly for the early developmental stages

Aim:

- Define (some of) the parameters related to the larval rearing in order to optimize the applied methodologies
 - larvae are optical predators
 - adequate foraging of appropriate preys



■ **Objectives:** to obtain new information on the larval rearing parameters

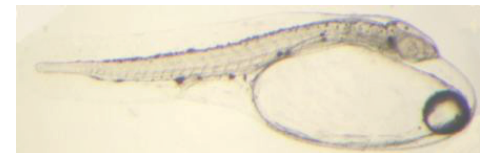
- Acquire basic biological information
 - Ontogeny of digestion and vision system
 - Ontogeny of Somatotropic axis
 - Skeletal Ontogeny

- Feeding regime - Prey enriching diet

- Husbandry parameters
 - tank type-shape,
 - stocking density,
 - light conditions



Ontogenesis of key biological systems



Mesocosm

- A semi-intensive technology that facilitates the rearing of several species providing the required “biological control”



Intensive rearing

- The “pseudo-green” water methodology – closed hydraulic circuit
- Controlled conditions
 - temperature
 - photophase
 - feed distribution
 - computer based application



Methodology (HCMR)

Mesocosm

- Water: filtered (5 μm) natural seawater (salinity 40 psu) UV treated
- T: 24 ± 0.7 °C
- pH: 8.0 to 8.2
- D.O. 5.8 to 6.8 mg l⁻¹.
- Photophase: 24L:00D from 3 to 25 dph then to 18L:06D.
- Light intensity: 500 lux to 1,000 lux, ~250 lux at night

- Feeding
 - 3 to 18 dph : Microalgae (*Chlorella sp*) and enriched rotifers (*Brachionus sp*)
 - 12 to 14 dph. *Artemia* AF A₀ nauplii and 14 to 30 dph enriched *Artemia* EG A₁ nauplii
 - 16 dph (MES) and 21 dph (INT) artificial feeds (grain size 200–300 μm ; and 300–500 μm)
 - Frozen gilthead sea bream eggs in MES after 20 dph
 - Mesocosm tanks developed naturally zooplankton (harpacticoida copepods)

Intensive

- Water: filtered borehole 35 psu-water.
- T: 22 ± 0.5 °C until mouth opening
 24 ± 0.5 °C afterwards.
- pH: 8.0 to 8.2
- D.O. 6.8 to 7.2 mg l⁻¹.
- Photophase: 24L:00D from 3 to 25 dph then 18L:06D
- Light intensity: 200 - 800 lux
~200 lux at night

Methodology (FCPCT)

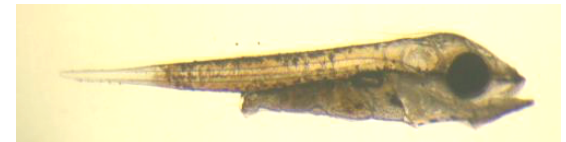
- Two different tanks types: 40,000 l and 2,000 l
 - Photoperiod was natural (14:10 h, L:D),
 - S=37 psu, and
 - T=25-27°C
 - DO=6.78 ± 0.5 ppm
 - Feeding: live phytoplankton (*Nannochloropsis sp.*), enriched rotifers (1-30 dph), enriched *Artemia* (12-30 dph), microdiets of 75, 150 and 300 µm (13-30 dph)



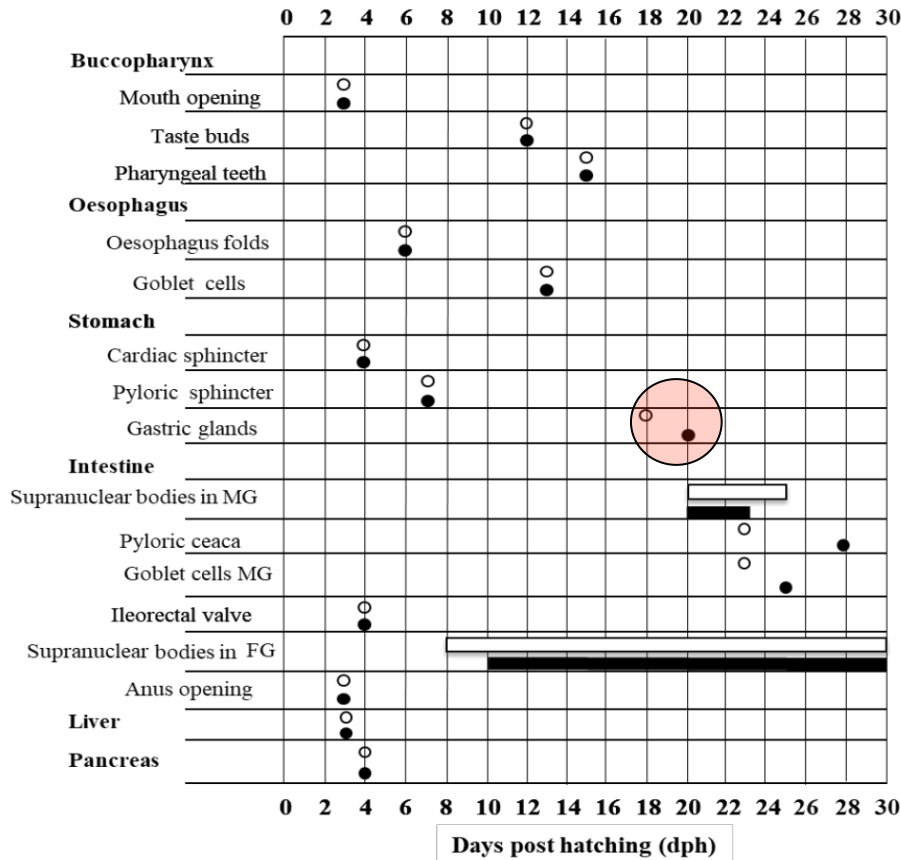
Ontogeny of digestive and vision systems

■ The digestive system

- enables the fish to capture, ingest, digest and absorb nutrients from the food
- is of special relevance to establish the larval feeding protocol
- ontogenesis of related organs and digestive enzymes



The *digestive system* ontogenesis is a rapid process



The **gastric glands** mark the transition from larval to juvenile and the development of a functional stomach

- in greater amberjack appeared after 5.5 mm of TL in all the rearing systems (16-20 dph).



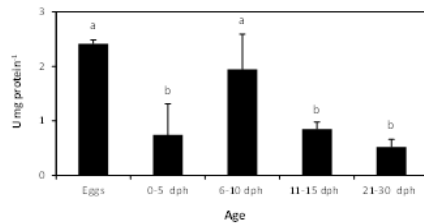
	0	1	2	3	4	5	6	7	8	9	12	15	17	20	23	25	28	30
INT	3.52	3.72	3.73	3.81	3.96	4.01	3.94	3.98	4.23	4.23	4.55	5.26	5.19	6.01	6.35	7.77	8.47	8.20
MES	3.52	3.67	3.75	3.83	3.99	4.05	4.03	4.02	4.05	4.09	4.53	5.00	5.69	6.49	7.2	9.23	8.54	8.74

Total length in (mm)

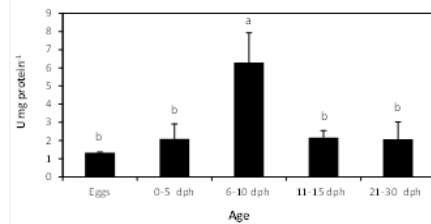
	Growth rate (exp)	Survival
Mesocosm	0.065 d ⁻¹	13.9±0.8%
Intensive	0.047 d ⁻¹	8.2±3.1%

Ontogenesis of *digestive enzymes*

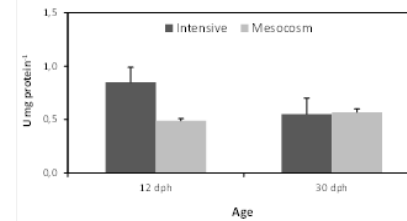
a) Amylase



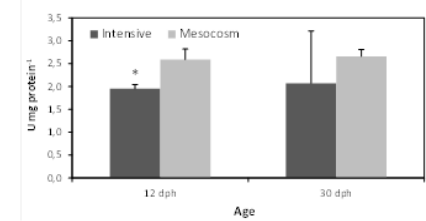
b) Lipase



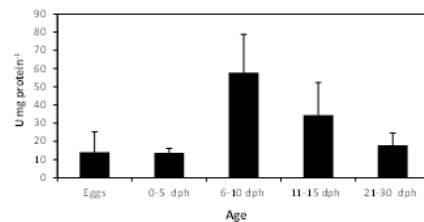
a) Amylase



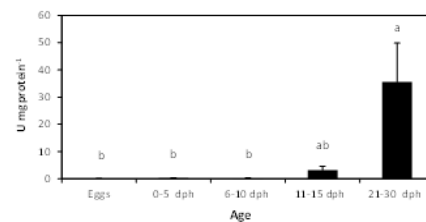
b) Lipase



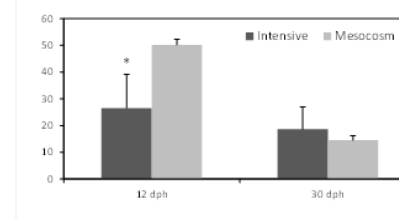
c) Alkaline protease



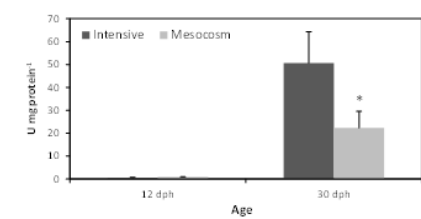
d) Pepsin



c) Alkaline protease

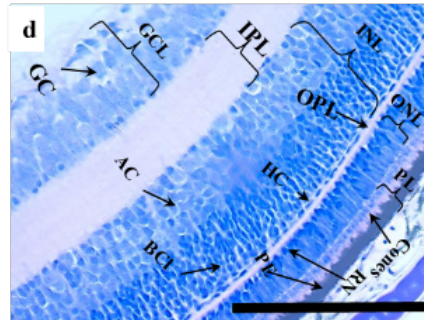
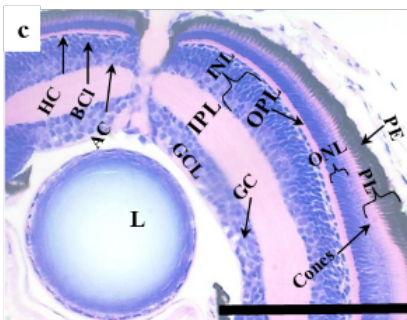
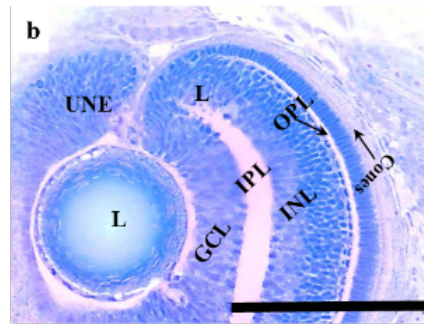
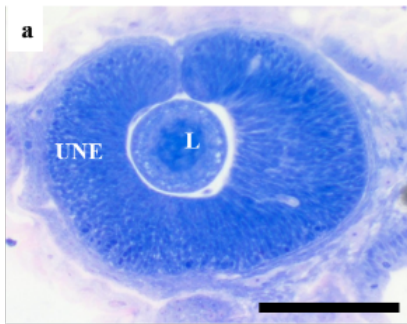


d) Pepsin



- 0-3 dph: enzymatic activities before onset of exogenous feeding
importance of egg glycogen as energy during the embryonic development
- 3-5 dph: higher activity of alkaline protease than lipase
proteins the main energy source
- 5-10 dph: period of rotifer feeding
proper nutritional condition of the larvae
- 10-15 dph: in both systems malnutrition
difficult acceptance of Artemia

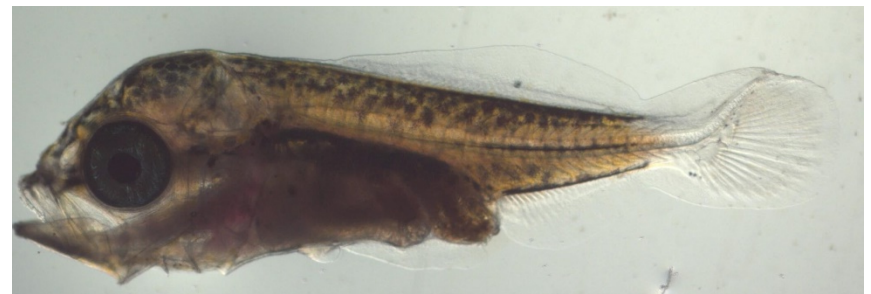
Ontogenesis of *Vision system*



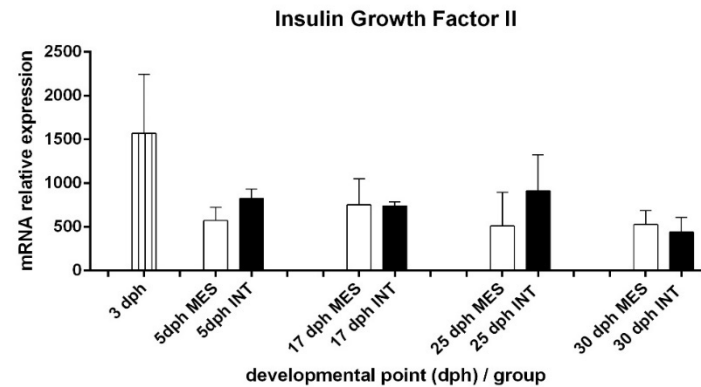
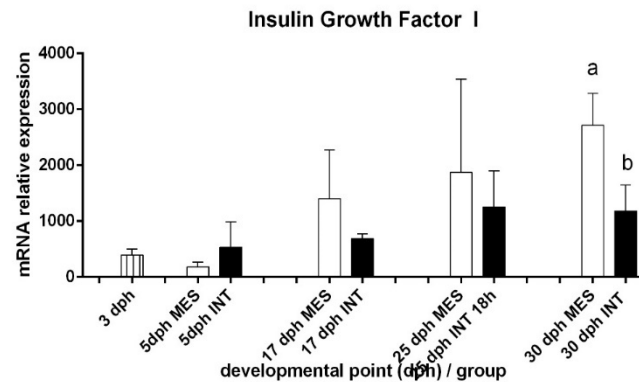
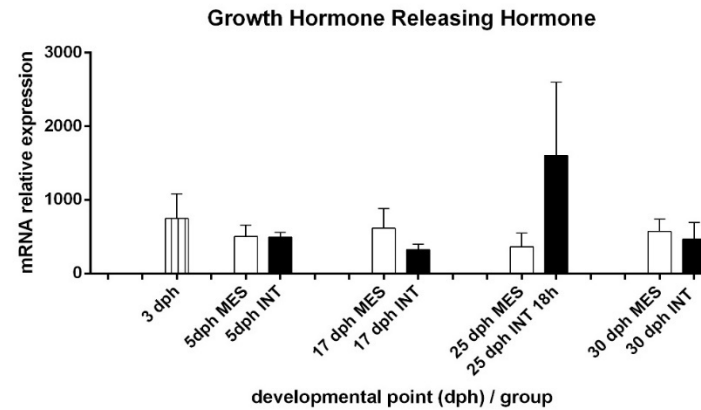
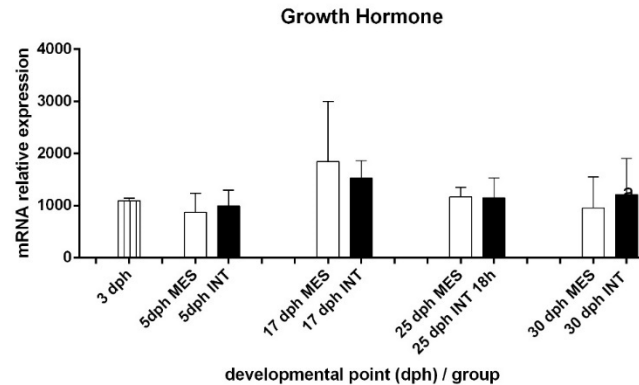
- 0 dph: retina an undifferentiated neural epithelium (UNE)
- 1-2 dph the first structures appeared
 - ganglia cell layer (GCL), the inner plexiform layer (IPL), the inner nuclear layer (INL), the outer plexiform layer (OPL), the outer nuclear layer (ONL) and the photoreceptor layer (PL).
- 3 dph- onwards: the pigment epithelium PE appeared on the external area of the retina
 - the nucleus of the cone cells appeared in the outer nuclear layer
- Rods first appeared at TL 5.0 ± 0.2 mm

Lessons learned

- Transitions to juveniles after 5.5 mm
- Protein main energy source since first feeding
- Night vision after 5.0 mm
- Rotifer appropriate as diet
- Difficult adaptation to Artemia



Ontogenesis of the somatotrophic axis

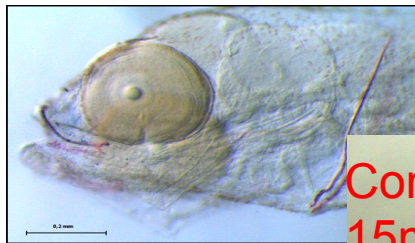


IGF Binding Proteins 1, 2, 3, 4, 5

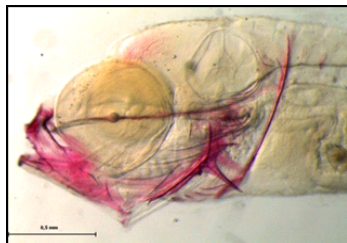
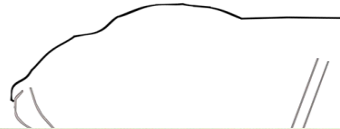
Available for the first time information on the regulation of the various components of the IGF signaling pathway in greater amberjack

Skeletal Ontogeny

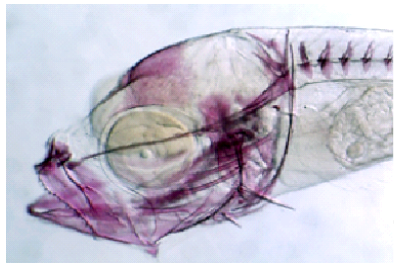
Cranial development



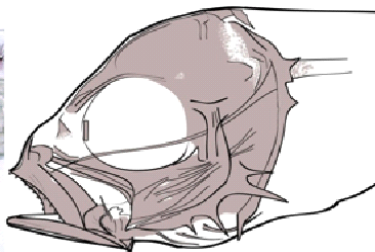
TL: 3.38 ± 0.15 mm



TL: 4.49 ± 0.14 mm



TL: 10.15 ± 1.86 mm



Vertebral column and fin ontogeny

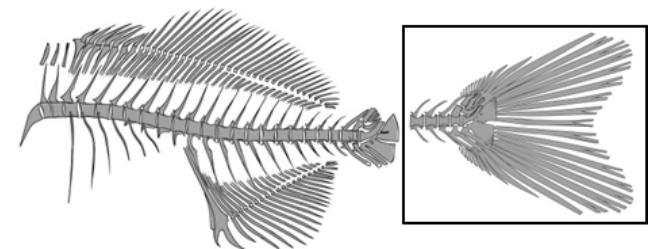
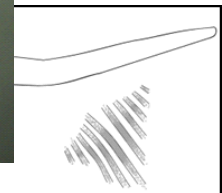


Completed bone mineralization of the vertebral column at 15mm



mm

TL: 5.36 ± 0.2 mm



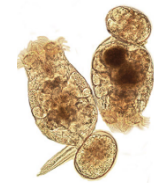
TL: 13.03 ± 0.09 mm

Effect of feeding regime and probiotics

- Effect of
 - prey concentration and supply frequency
 - use of immune modulators substances during the rotifer administration

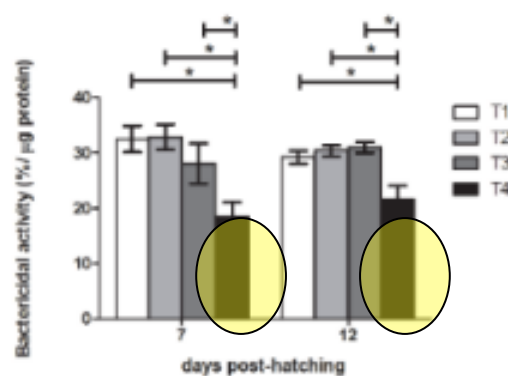
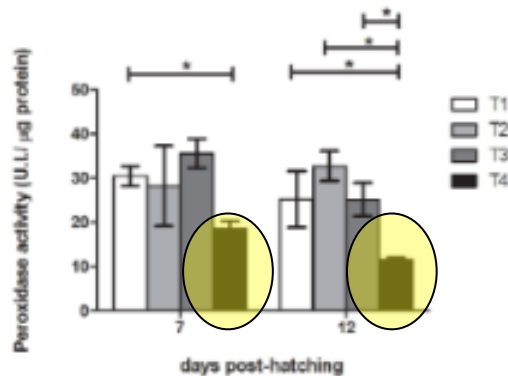
- Rotifers
 - Density: 5 vs 10 rot ml⁻¹
 - Distribution 2 vs 3 times d⁻¹

- Four enriching diets
 - A commercial diet (T1)
 - LC60/20:4n-6/10ppm carotenoids basic emulsion (T2),
 - LC60/20:4n-6/10ppm carotenoids basic emulsion with 20% *Echium* oil (T3)
 - LC60/20:4n-6/10ppm carotenoids basic emulsion with 20% black cumin oil (T4)

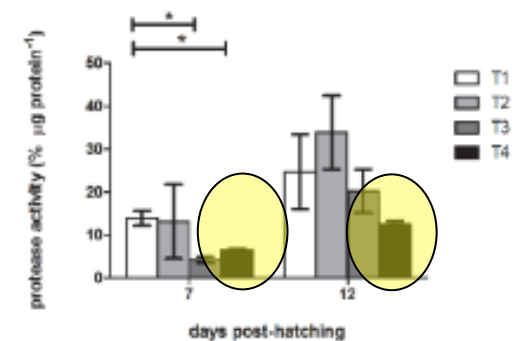
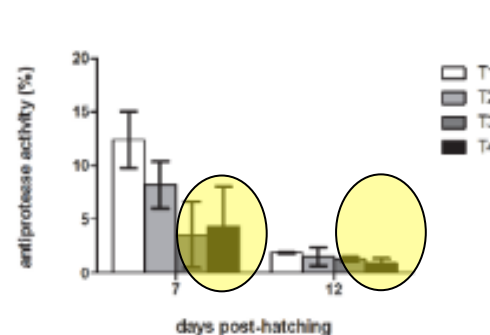


Larval growth and survival

- Similar regardless rotifer density
 - slightly better at 5 rot ml⁻¹
- Better for feeding 3 times day⁻¹
- Better with T4

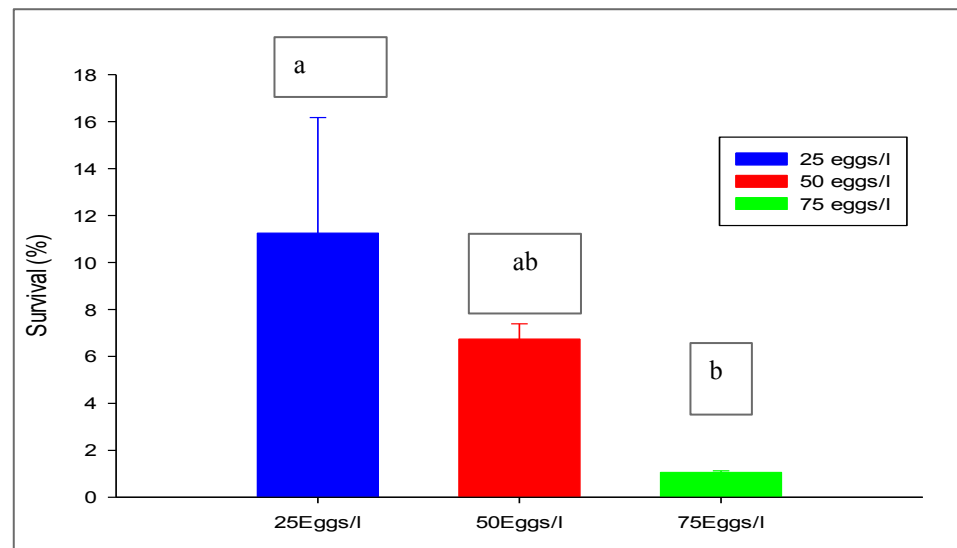
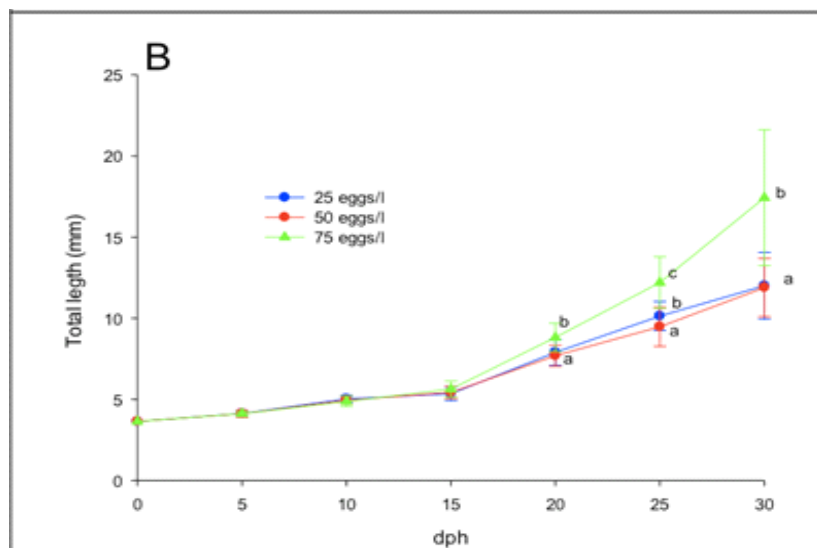


Humoral innate immune activities
7 dph and 12 dph larvae



Effect of larvae density

- 3 densities: 25, 50 and 75 eggs l⁻¹
 - Two different tanks types: 40,000 l and 2,000 l tested for 30 days
 - Photoperiod natural (14:10 h, L:D),
 - S: 37 psu,
 - T: 25-27°C
 - DO: 6.78 ± 0.5 ppm
 - Feeding: live phytoplankton (*Nannochloropsis sp.*), enriched rotifers (1-30 dph), enriched *Artemia* (12-30 dph), microdiets of 75, 150 and 300 μm (13-30 dph)



Deformities observed

A: Lordosis

B, G: Vertebral fusion

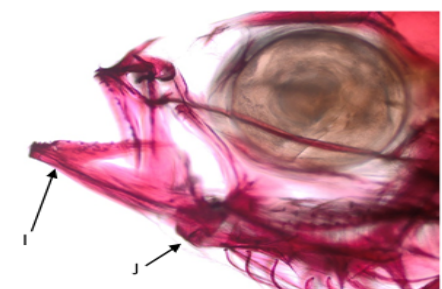
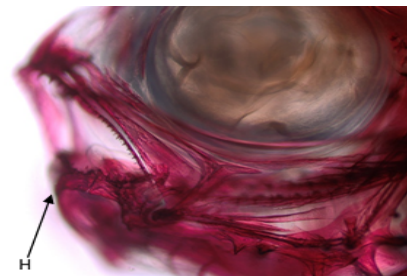
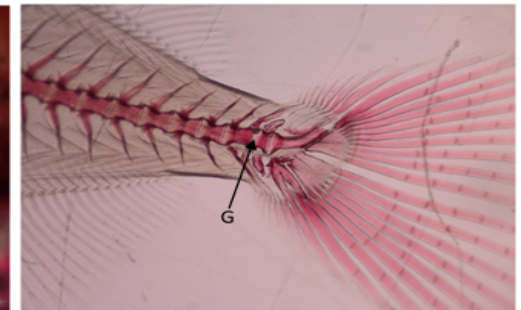
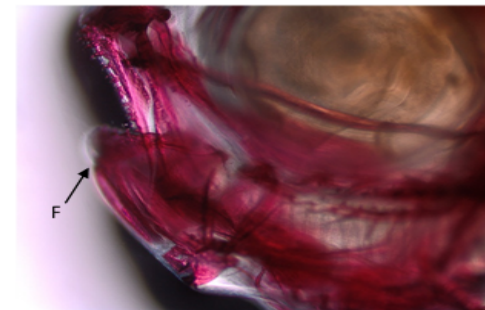
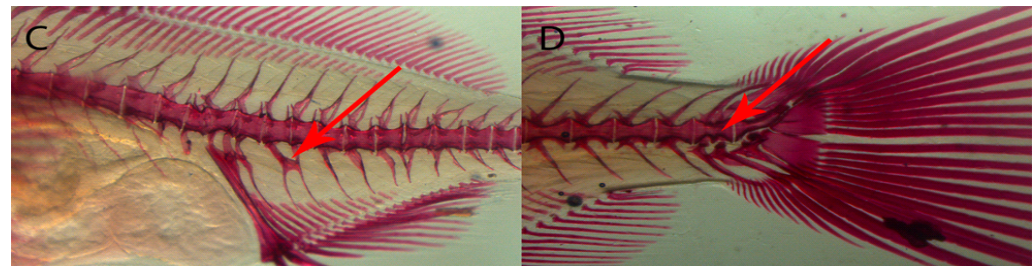
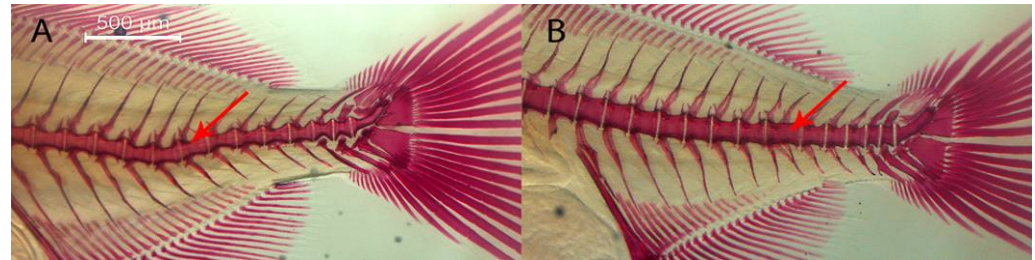
C: Fusion of neural arch and spines

D: Fusion of caudal vertebrae

F, H, I: Anomalous dentary

G: Partial vertebral fusion in caudal vertebrae

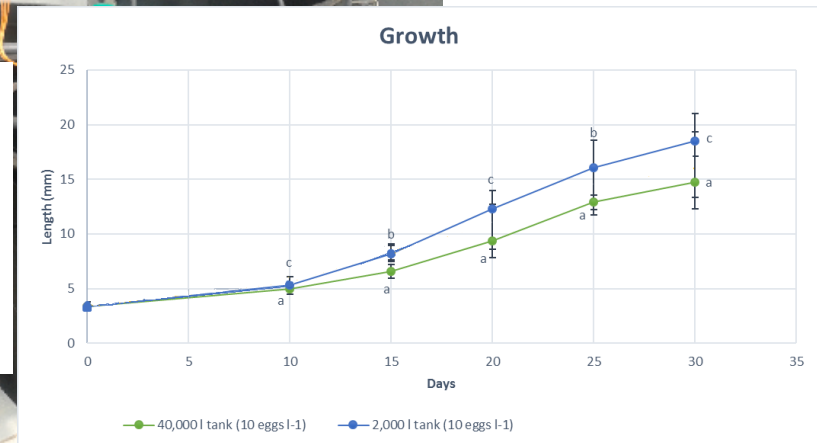
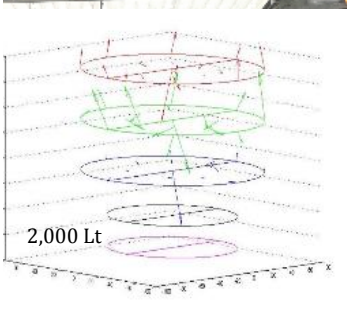
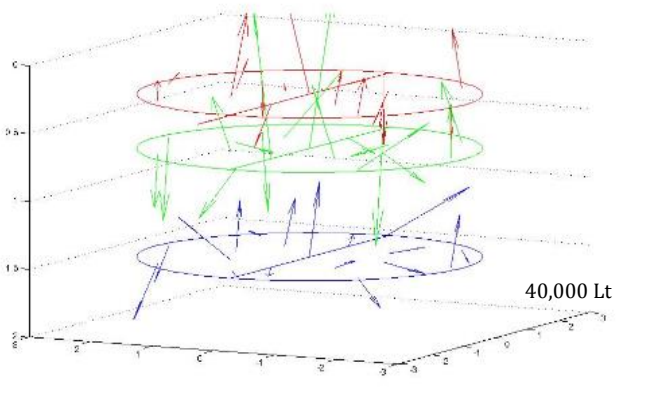
J: Cephalic anomalies (glossohyal)



Effect of environmental parameters

Tank hydrodynamics

- Two different tanks types: 40,000 l and 2,000 l were tested
 - eggs from natural spawning (10 eggs l⁻¹)
 - photoperiod natural (14:10 h, L:D),
 - S: 37 psu,
 - T: 25-27°C.
 - DO: 5–8 g l⁻¹.
 - Feeding: live phytoplankton (*Nannochloropsis sp.*), enriched rotifers (1-30 dph), enriched *Artemia* (12-30 dph), microdiets of 75, 150 and 300 μm (13-30 dph)



Differences in current profiles (generally higher in the 2,000 l tanks)

Survival low: 2-3%

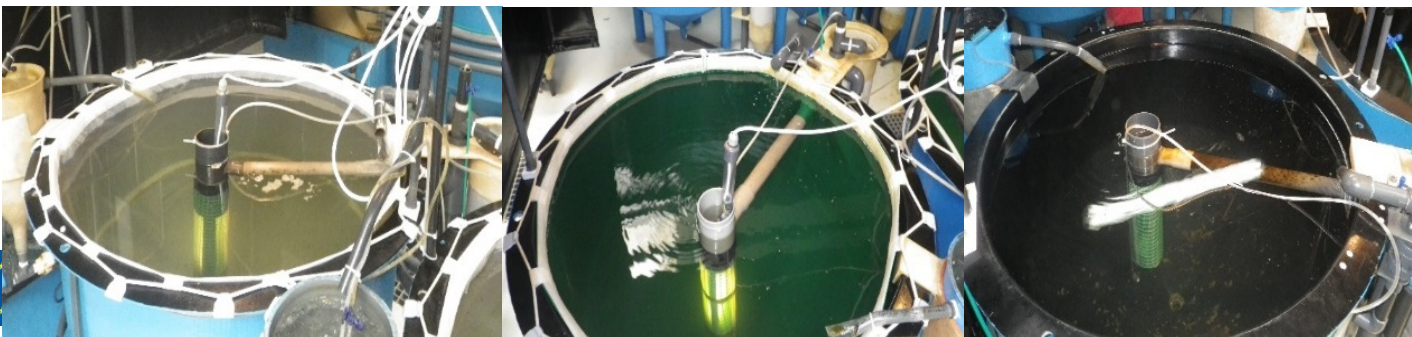
The effect of light (intensity and duration)

- Photophase 18L:06D vs 24L:00D)
- Background color (black, green, white)

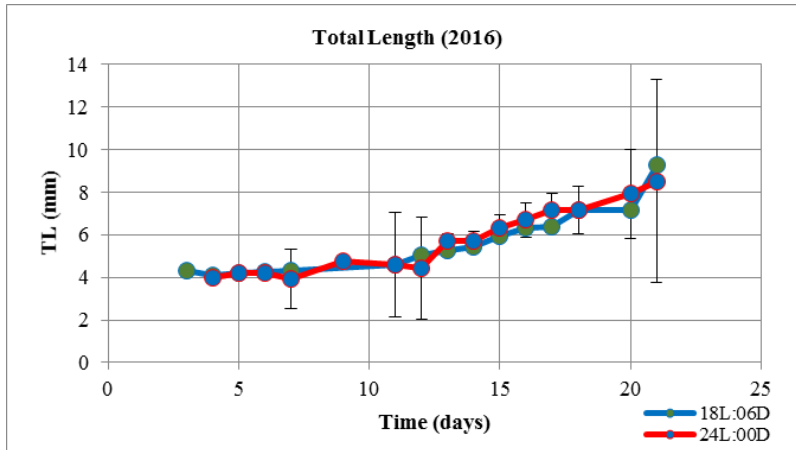
Intensive rearing



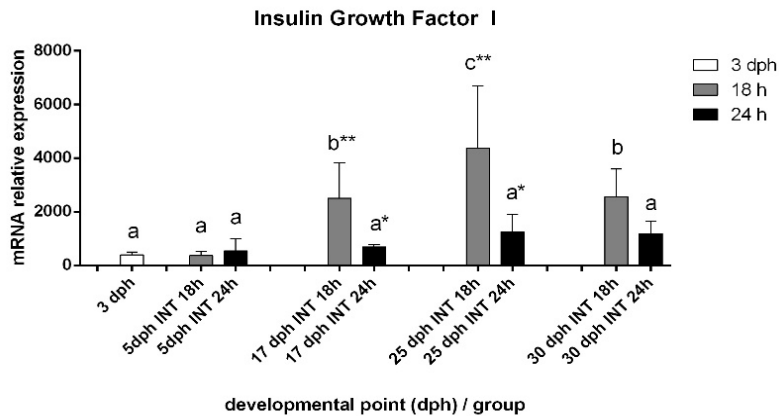
- The “pseudo-green” water methodology – closed hydraulic circuit
 - Water: filtered borehole 35 psu-water.
 - T: $22 \pm 0.5^{\circ}\text{C}$ until mouth opening
 $24 \pm 0.5^{\circ}\text{C}$ afterwards.
 - pH: 8.0 to 8.2
 - D.O. 6.8 to 7.2 mg l⁻¹.
 - Photophase: 24L:00D from 3 to 25 dph then 18L:06D
 - Light intensity: 200 - 800 lux
~200 lux at night



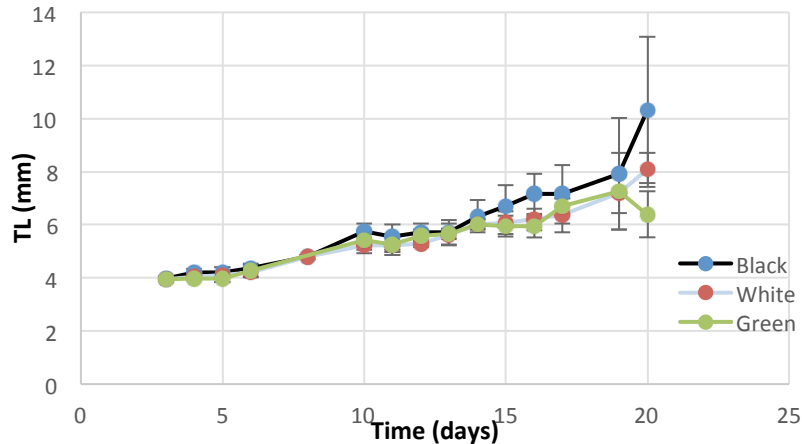
Results - Photophase



	Growth rate (exp)	Survival
18L:06D	0.031 d ⁻¹	10.6±4.2%
24L:00D	0.031 d ⁻¹	8.2±3.1%

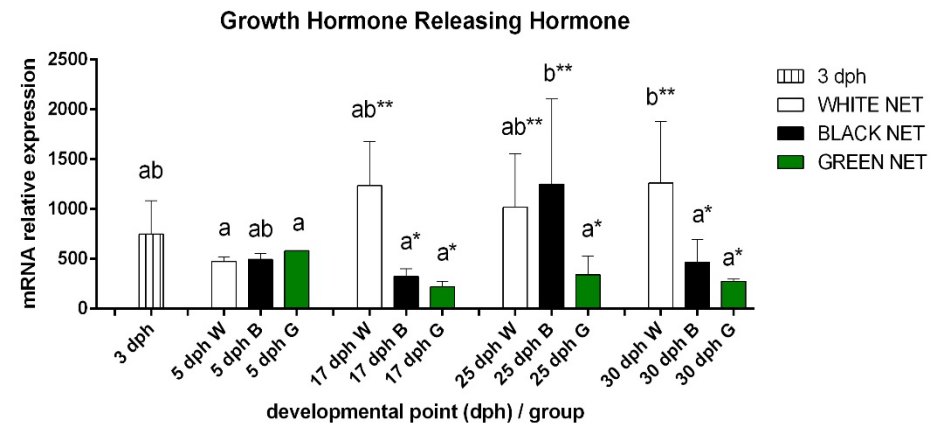
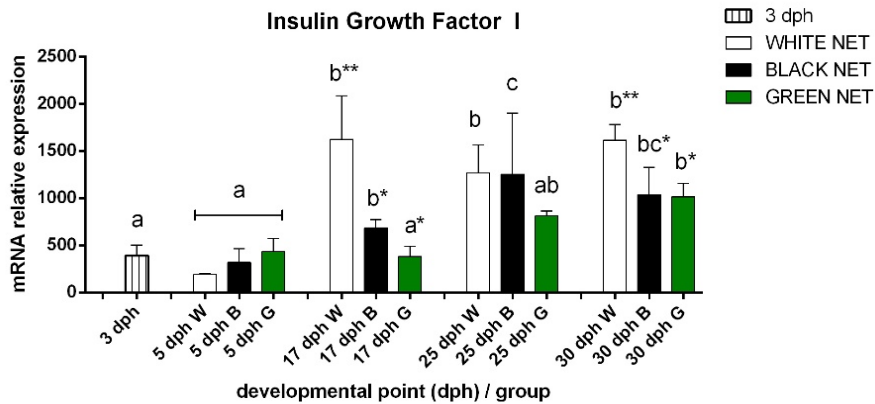


Results - Background color



Growth rate (exp): White: 0.0393 d⁻¹
 Black: 0.0481 d⁻¹
 Green: 0.0355 d⁻¹

SURVIVAL: WHITE: 22,2%
 BLACK: 8,2%
 GREEN: 16,5%



- Different expression of genes IGF-I, IGF-II and IGF Binding Proteins 2,3 & 4
- higher levels for **WHITE** background

- Max light intensity in the water column at 30 cm ~50 lux
- 3-4x higher than without light

Table 3. Measured light intensity (Lux) at several points in both tanks of each treatment. Values are means \pm standard deviation.

Measurement point		Green-no light	White-no light	Green-light	White-light	Black
Under surface	Centre (n=2)	334 \pm 4	303 \pm 14	255 \pm 11	234 \pm 1.1	295 \pm 24.6
	Side (n=8)	86 \pm 102	78 \pm 98	64 \pm 10	66 \pm 82	68 \pm 11
Depth ~30cm	Angle 45° (n=8)	15 \pm 7.4	13 \pm 6.2	11 \pm 4.5	11 \pm 7.7	12 \pm 2
	Towards centre (n=8)	11 \pm 1.7 ^a	12 \pm 1.1 ^a	31 \pm 1.7 ^b	27 \pm 4.8 ^b	9 \pm 3

- This study showed a catalytic effect of the light conditions on larval survival

- may reflect the effect of the species

- The presence of light in the rearing environment may affect any previous results

- Indicate a need for the rearing of larvae



of the

itude of

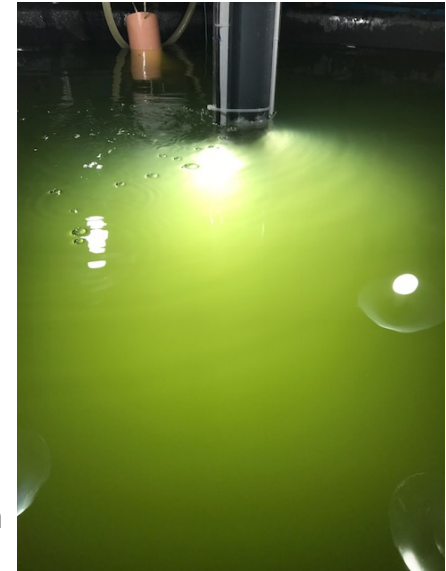
larval

Towards an industrial protocol

Validation in Commercial hatcheries

■ Hatchery 1

- Larvae density 75 ind l⁻¹
- Light intensity
 - 800 lux on 3-6 dph, 1200 lux on 6-12 dph 1000 lux 15 dph 500 lux 20 dph
- Photophase: 24L:00D from 3-20 dph then 18L:06D
- Feeding: phytoplankton, enriched rotifers, Artemia and dry feeds. Frozen eggs after 20dph
- Size selection during weaning
- In 2017
 - 15.000 juveniles classified in 4 size-classes between 0.3 and 2.5 g to nursery
- In 2018 similar method
 - until 20 dph survival ~10%
 - between 20-30 dph: no sorting → mortality >65% due to cannibalism
 - 30 dph 15.000 individuals were transferred to nursery



Validation in Commercial hatcheries (2)

■ Hatchery 2

- Direct incubation of eggs in the larval rearing tanks
- T: 24.5 to 25.0 °C.
- pH: 7.8
- Light conditions : modified in some tanks
 - intensity on the surface >1000 lux
- Photophase: 24L:00D from 3-20 dph then 18L:06D
- Feeding: enriched rotifers, instar I (TL >5mm) and enriched instar II (TL >6mm) Artemia nauplii and artificial diets (TL >7mm).
- Tanks with high light intensity performed better
- Sorting in size from 20 dph, improves significantly survival
 - 48.300 juveniles of 25-50 g transferred in cages



Recommendations (1)

■ Rearing parameters

- large tank improves growth performance and survival
- egg stocking densities ~ 25 eggs l⁻¹
- DO: 5.0 to 8. mg l⁻¹, preferably >6.0 mg l⁻¹
- S: 37 - 40 psu
- pH: 7.8-8.5
- T: 22 to 27°C, preferably between 23.5 and 25.0°C
- Photophase 18L:06D from 1 to 25 dph
- Light intensity: Defused light of 800-1200lux, 3-12 dph
gradually 800 lux until 20 dph

■ Feeding

- coordinate rearing conditions and larval development
- live microalgae at 150-300 x 10³ cell ml⁻¹ since 2 dph
- enriched rotifers min 2x d⁻¹, 3-15 dph, at 3-7 rot ml⁻¹; Artemia AF (TL>5mm), for 2-3 days; enriched Instar II (TL> 6.0 mm); dry diet (200-800 µm) (TL>6.5 mm)
- immune modulators (*Echium* and black cumin oil) together with optimized enriching emulsion and astaxanthin as carotenoid improve larval performance

Recommendations (2)

■ Husbandry

- high size variability (at 20-30 dph) in all rearing systems tested until today (unknown reasons)
 - early sorting to appropriate size classes
 - standard methods and equipment available in all hatcheries
 - significant higher survival compared to unsorted groups although the losses
 - mortality for unsorted groups >90%, for sorted groups 10-15%.
- Transportation of individuals
 - larvae with TL <15 mm do not tolerate netting, transfer performed with care.
 - larvae with TL >20 mm can be netted normally
- individuals >0.5-1 gr are handled easier
 - light anesthesia may help



2mm

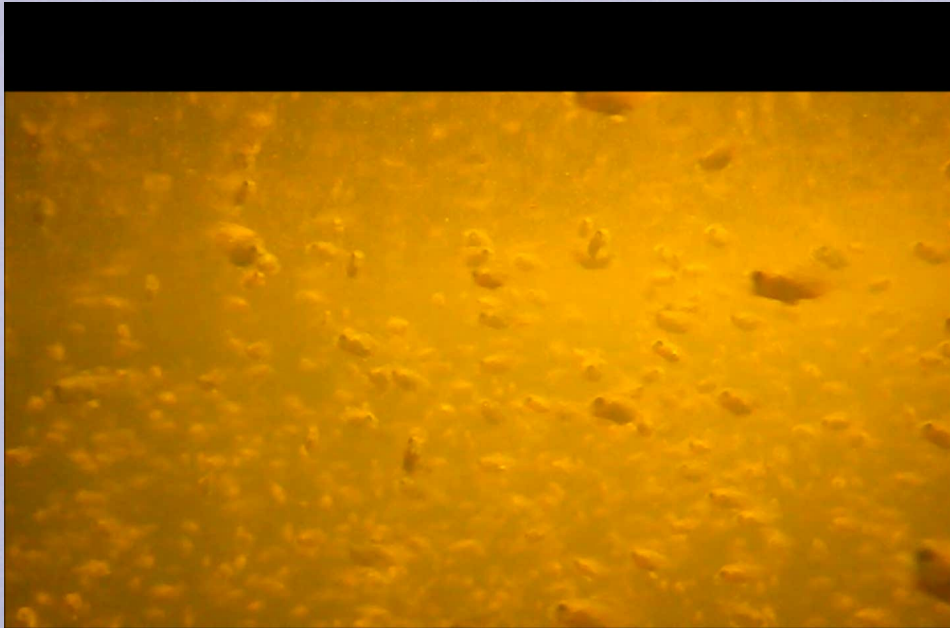


5mm



20mm

Thank you!



Greater Amberjack larval rearing, HCMR 2016



Hellenic Centre for Marine Research
Institute of Marine Biology, Biotechnology
& Aquaculture



Fundación Parque Científico Tecnológico
Universidad de Las Palmas de Gran Canaria



INSTITUTO
ESPAÑOL DE
OCEANOGRAFÍA

