



Parasitic infections in greater amberjack in Greece



Pantelis Katharios, Nikos Seimenis Hellenic Centre for Marine Research Barcelona, 17 Jan 2017

Parasites

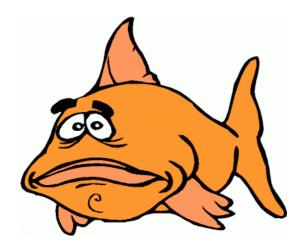


Parasitism

In biology/ecology, parasitism is a non-mutual symbiotic relationship between species, where one species, the parasite, benefits at the expense of the other, the host.

The "expense"

- In nature the expense is usually low
- In aquaculture it is BIG !!!



The cost of parasitic diseases

SPECIAL ISSUE ARTICLE

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Economic costs of protistan and metazoan parasites to global mariculture

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There are many ways to categorize parasites

- Macroparasites/microparasites
- Metazoa/protozoa
- Extracellular/intracellular
- Endoparasites/Ectoparasites
- Obligate/facultative parasites
- Generalists/Specialists
- Direct/Indirect

Evolution of parasites

- Parasites co-evolve with their host
- A successful parasite does not want to kill its host
- Less evolved parasites (eg Cryptocaryon irritans) are more pathogenic
- More evolved parasites are less harmful to their host
- They evolve strategies to "escape" host immune system
- They evolve strategies to disperse
- Some of these strategies are very resourceful and really fascinating

Aquaculture: Treatment or management?

We need to know:

- Life cycle
- Biology
- Ecology

Successful management = break the life cycle

Most important parasites in the Greek aquaculture

- Protozoa (Cryptocaryon irritans, Amyloodinium ocellatum)
- Myxosporea (Enteromyxum leei, Sphaerospora testicularis)
- Monogeneans (Sparicotyle chrysophrii, Diplectanum aequans)
- Digenean (Cardicola aurata)
- Crustacean (Lernanthropus kroyeri, Ceratothoa oestroides)

Main parasites of greater amberjack

Europe

- Zeuxapta seriolae
- Heteraxine heterocerca
- Neobenedenia sp.
- Paradeontacylix sp.
- Amyloodinium ocellatum

Greece

- Zeuxapta seriolae
- Paradeontacylix sp.
- Cryptocaryon irritans

Zeuxapta seriolae

- Monogenean worm
- Gill parasite
- Direct life cycle
- Highly host specific
- Causes anemia
- High mortality in cultured amberjacks

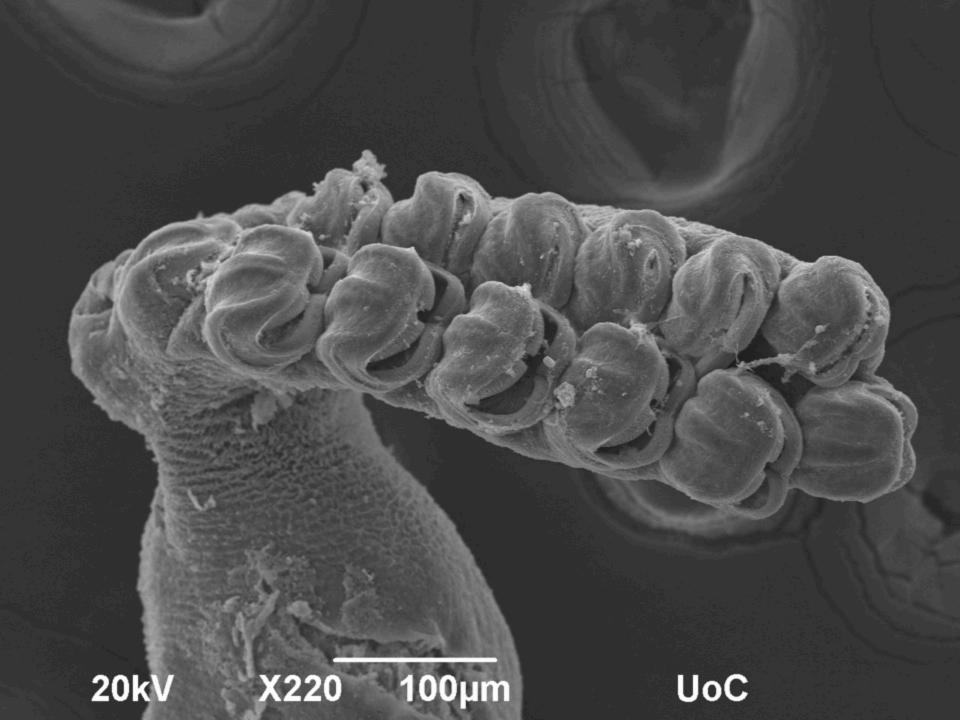


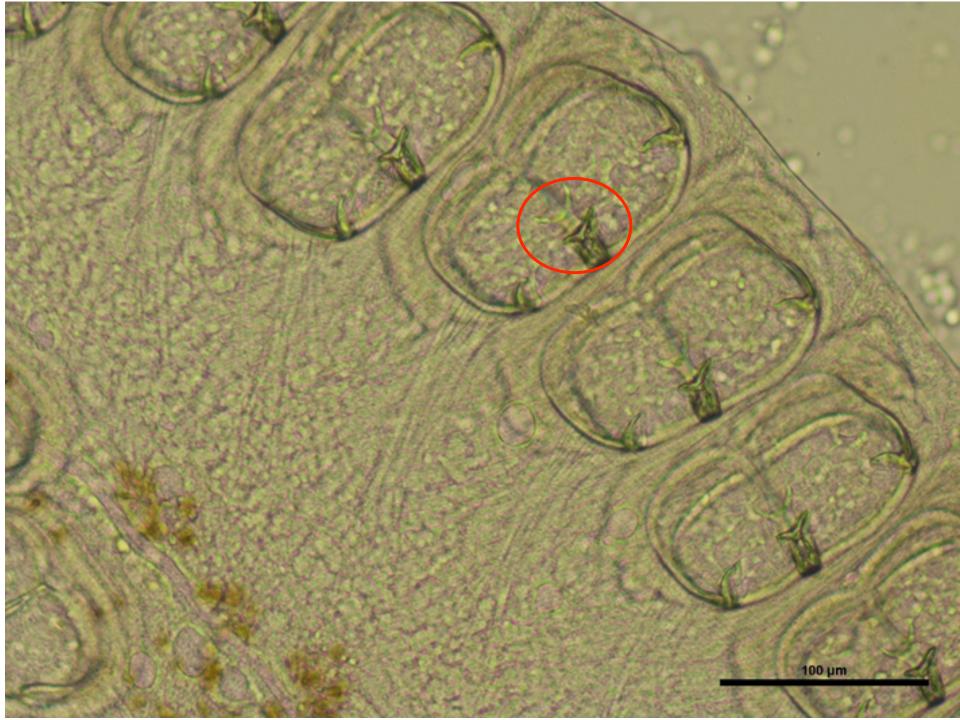
Identification

- Host-specificity
- Morphology of the parasite
- PCR (COI, 28s)

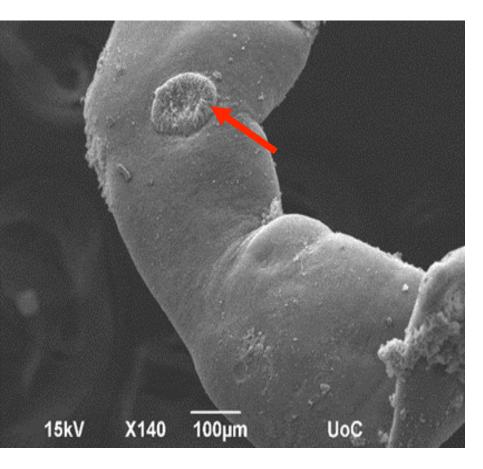
Morphology

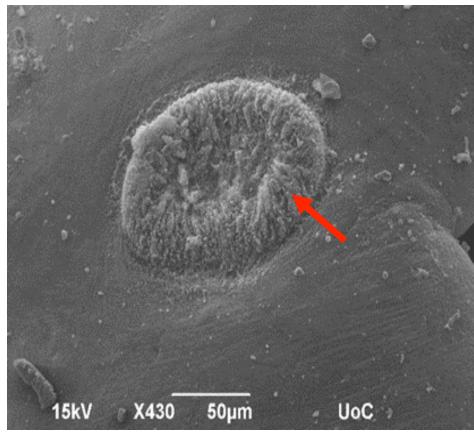








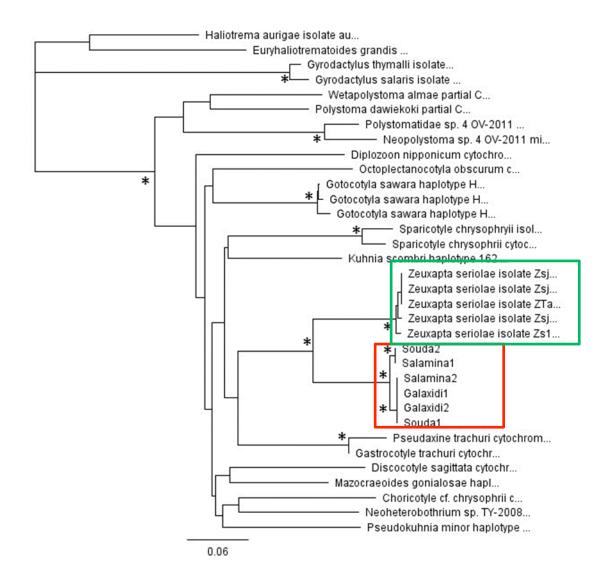




Molecular characterization BLAST results

Description	Max score		Query		Ident	Accession
Zeuxapta seriolae isolate Zsjf3aJF13 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	814	814	98%	0.0	84%	KP119287.1
Zeuxapta seriolae isolate Zsjf3aJF13 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial			93%	0.0	84%	KP119287.1
Zeuxapta seriolae isolate Zsjf3aJF13 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	823	823	98%	0.0	85%	KP119287.1

Phylogeny (COI)



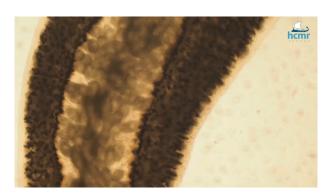
Life cycle

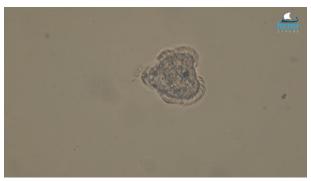






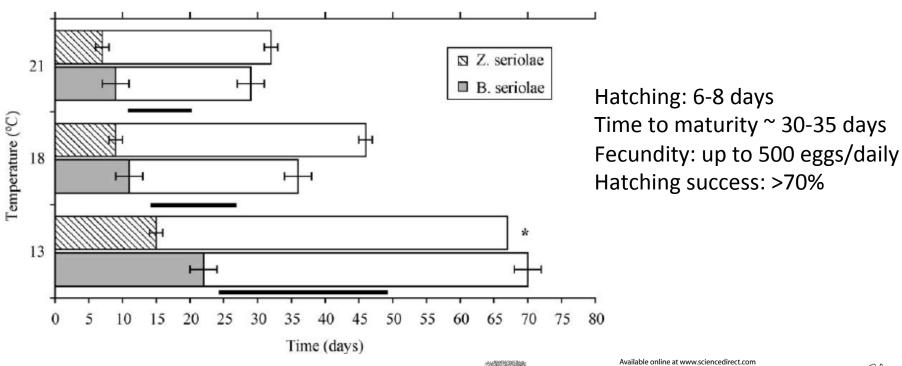








Life cycle 2





Available online at www.sciencedirect.com



International Journal for Parasitology 35 (2005) 315-327

Effects of temperature on fecundity in vitro, egg hatching and reproductive development of *Benedenia seriolae* and *Zeuxapta seriolae* (Monogenea) parasitic on yellowtail kingfish *Seriola lalandi*

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*School of Biological Sciences, University of Auckland, Private Bag 92019, P.O. Box 109 695, Auckland, New Zealand bNational Institute of Water and Atmospheric Research (NIWA), P.O. Box 109 695, Auckland, New Zealand Received 30 August 2004; received in revised form 12 November 2004; accepted 15 November 2004

Some simple (naïve) calculations

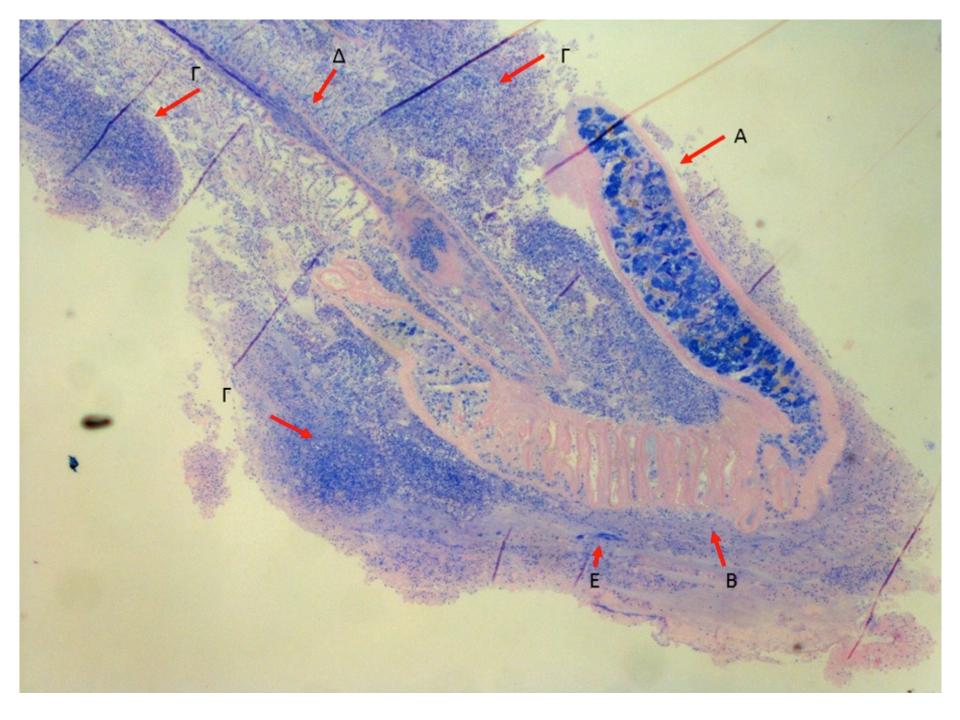
	# fish	parasite abundance	fecundity	spawning days	hatching success %	% attaching host	# parasites	new parasite abundance
gen 1	1000	5	500	5	70,00%	10,00%	875.000	875
gen 2	1000	875	500	5	70,00%	1,00%	15.312.500	15.313

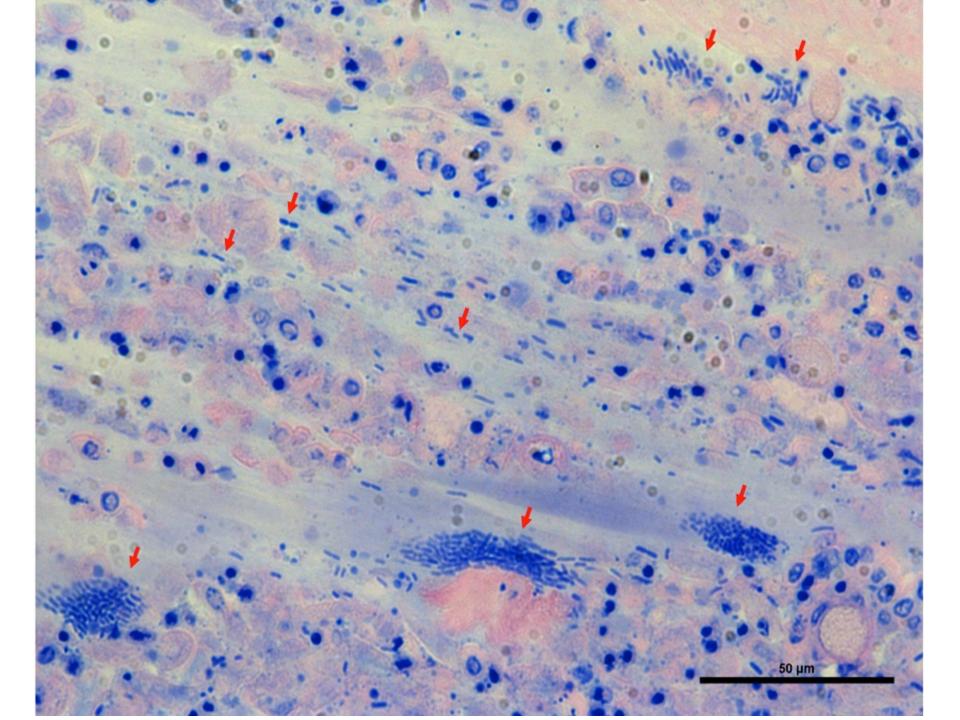
30 days generation time

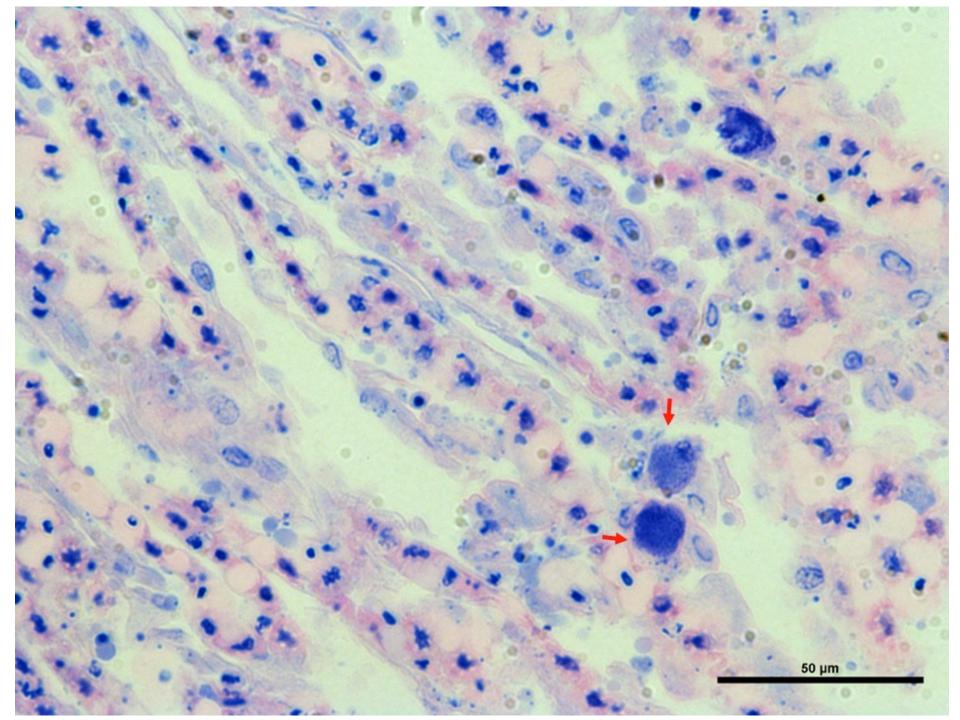
Pathology







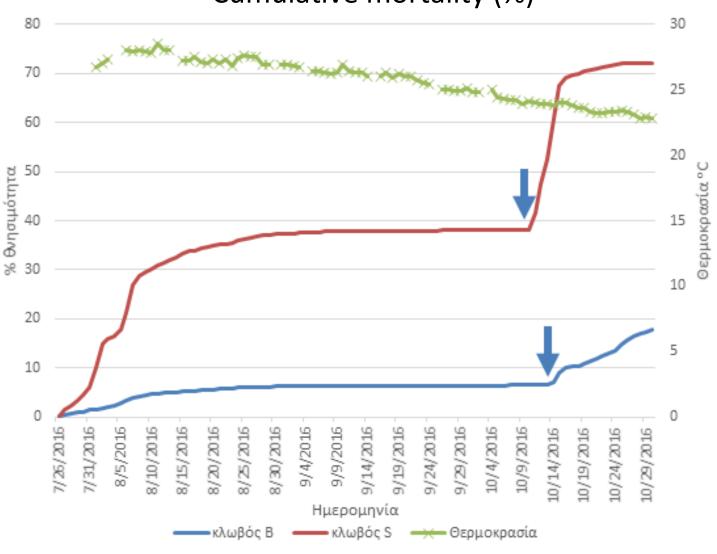




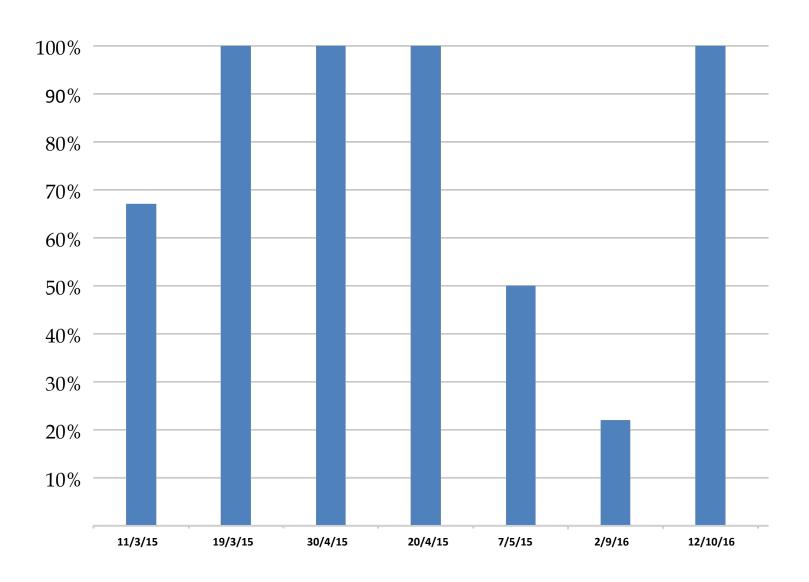
Haematology

Fish code	Hematocrit (%)	Hemoglobin (g/dl)	Weight (g)	Length (cm)
1	8	0.697	146	27
2	32	2.574	198	25
3	8	2.169	223	23
4	30	1.802	192	23
5	31	3.523	121	22
6	29	7.207	398	30
7	14	6.71	385	29
8	16	2.169	100	25
9	25	2.941	161	22
10	35	8.31	283	28

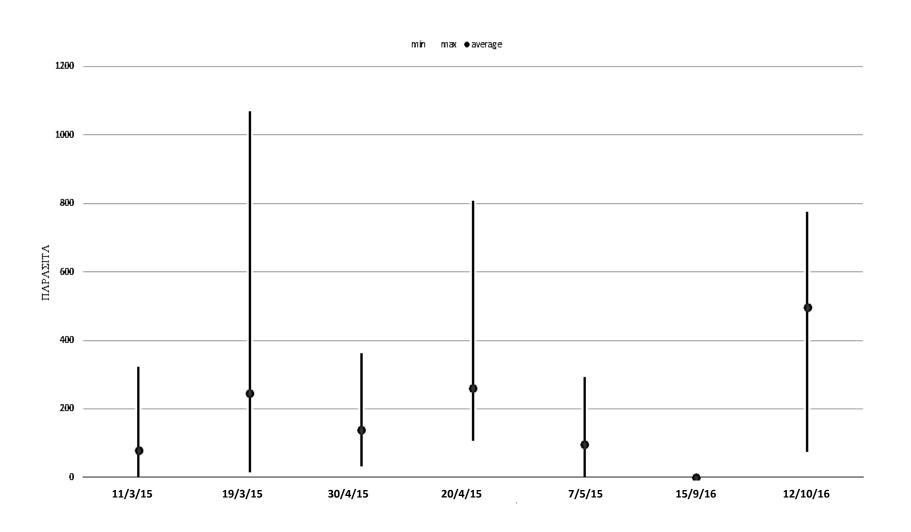




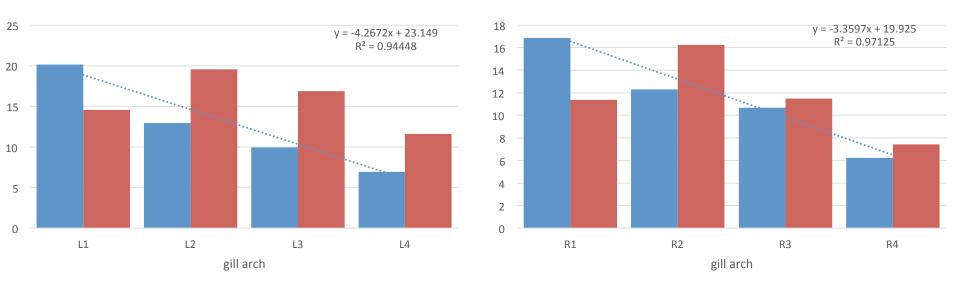
Prevalence



Mean abundance



Gill arch preference



Blue: Mature parasites Red: Juvenile parasites

Results

- 13745 parasites counted on 67 fish
- Prevalence 100% after March
- Maximum intensity >1000 parasites per fish
- Preference for the first gill arch
- Competition between parasite's age classes
- Yearlings most affected
- Continuous source of infection

Source of infection

- Wild populations
- Age class overlap in the same farm



Treatment

- Praziquantel in feed (150-200 mg/kg biomass 2-3 consecutive days – cycle should be repeated 2-3 times with a week interval). Palatability issues. Success depends on intensity of infection.
- Praziquantel baths (10-15 ppm for 30-60') very effective. Twothree baths with a week interval. Dissolve in ethanol or DMSO
- Ivermectin injection for bigger fish (150 µg/kg)
- Ivermectin baths 150 µg/L dissolved in DMSO for 1h. Very effective. Careful in dosage, it can be highly toxic!
- Thyme essential oil very effective in experimental trial (however it killed the fish as well !!!)

Management

- Clean nets
- Monitor fish on monthly basis. If parasites are found....treat!
- Age class overlap. If it cannot be avoided, try to orient the cages in a way that will make parasite disperse harder

