

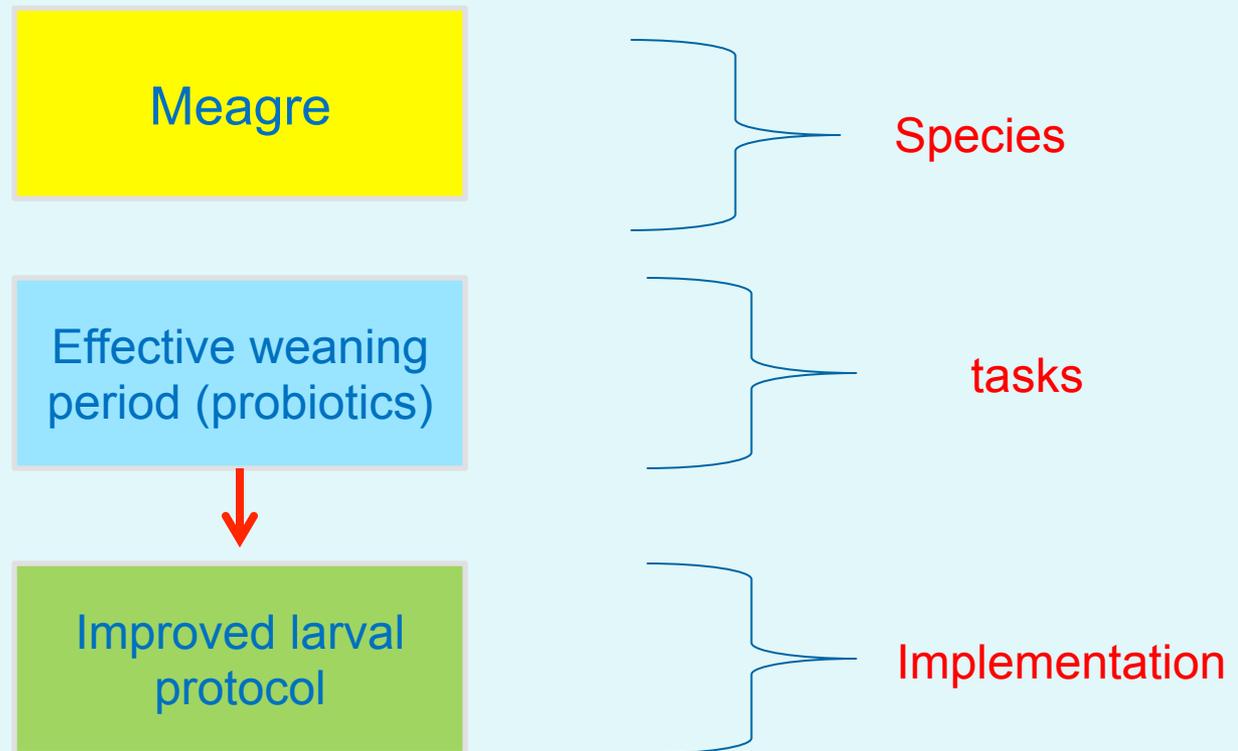


# Larval Husbandry (WP14-19)

**RTDs:** IOLR, IRTA,ULL, HCMR, FCPCT, IEO, DTU, IMR, NIFES,

**SMEs:** FORKYS, ASIALOR, SWH,CMRM, MC2, DOR

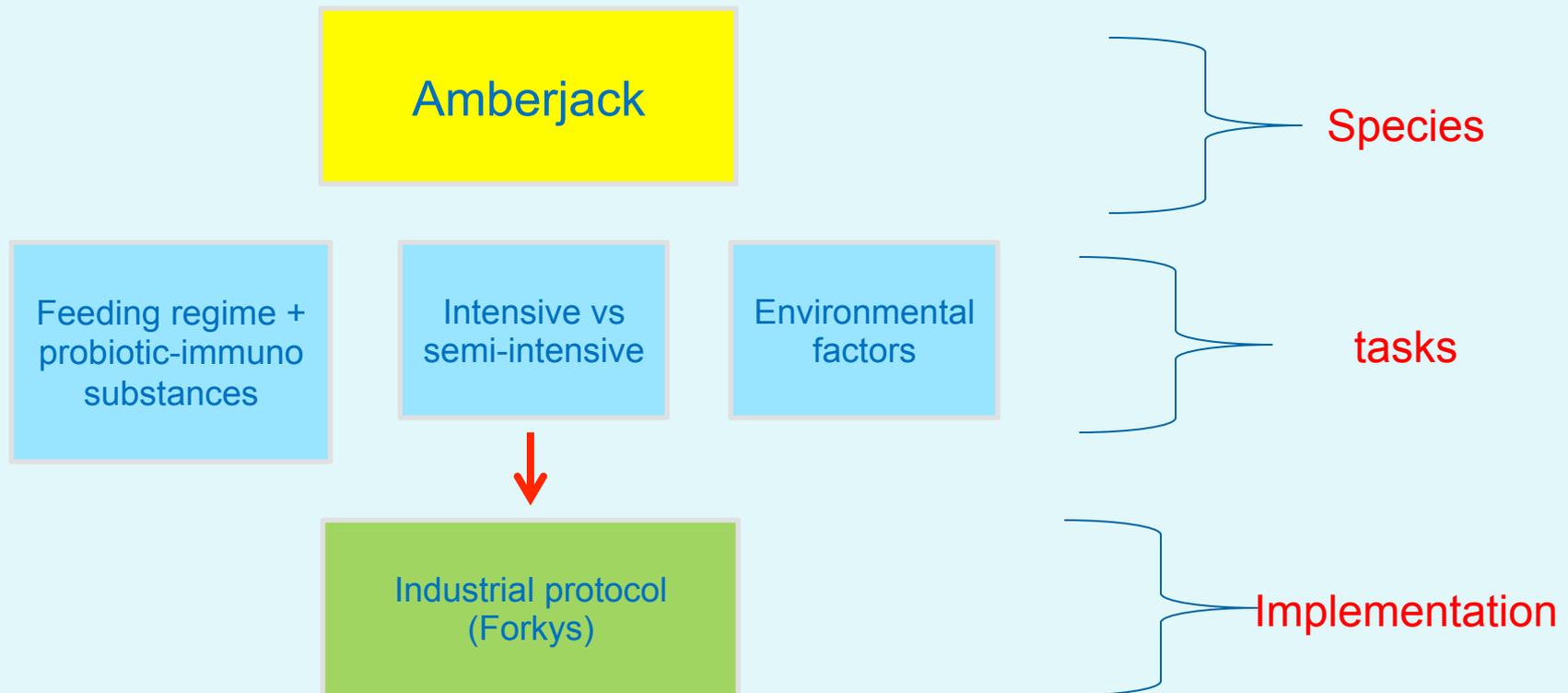
# Meagre strategy





<b>WP14 Meagre</b>			
<b>Tasks</b>	<b>Evaluation</b>	<b>Sub-tasks</b>	<b>Deliverable</b>
14.1 (IRTA)- effective weaning period (with or without probiotics)	•Growth, survival, size dispersion, j. quality, composition, pancreatic and intestinal enzymes (ULL)		<u>D14.1</u> Improved larval protocol

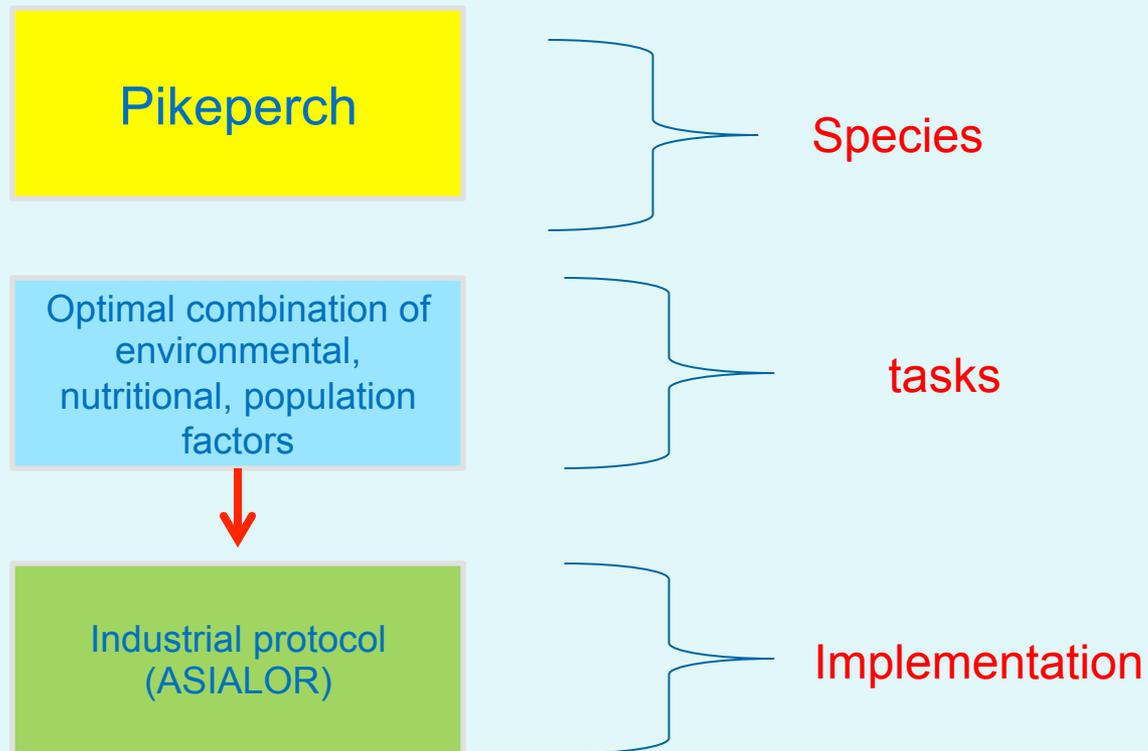
# Greater amberjack strategy



## WP15 Greater Amberjack

Tasks	Evaluation	Sub-tasks	Deliverable
<p>15.1(IEO) Effect of feeding regime (conc. and freq.) and probiotics</p>	<ul style="list-style-type: none"> <li>•Survival, growth, development, deform., nutr. Cond (IEO)</li> <li>•oxid. Stress, antiox. Defense enzymes , immune syst.(IEO).</li> <li>•Ontogeny of DT enzymes( ULL)</li> </ul>		<p><u>D15.2</u> feeding protocol and use of immune modulators</p>
<p>15.2 (HCMR) intensive vs semi-intensive systems</p>	<ul style="list-style-type: none"> <li>•Ontogeny of visual system, oxid. Stress, somatotropic axis, antiox. Defense enzymes , immune syst. (HCMR)</li> </ul>	<p>15.2.1 (HCMR) RAS vs Mesocosm</p> <p>15.2.2 (FCPCT) Stocking density</p> <p>15.2.3 (ULL) Ontogeny of D.T.</p>	<p><u>D15.1</u> Effective stocking densities</p> <p><u>D15.4</u> Ontogeny of visual and digestive system</p>
<p>15.3 (FCPCT) Environmental parameters</p>	<ul style="list-style-type: none"> <li>•Growth, survival, histology, b. comp, deform, stress+skeletal gene exp.,J. quality, size distribution, somatotropic axis (FCPCT)</li> </ul>	<p>15.3.1 (FCPCT) Tank hydrodynamics</p> <p>15.3.2 (HCMR) Light intensity and duration</p>	<p><u>D15.3</u> Optimum hydrodynamics and light conditions.</p>
<p>15.4 (IEO) Development of Industrial protocol</p>	<ul style="list-style-type: none"> <li>•Survival, growth, development, deform., nutr. Cond.</li> <li>•oxid. Stress , antiox. Defense enzymes, immune syst.</li> <li>Interval sampling for ossification patterns and skeletal deformities.</li> </ul>	<p>15.4.1 (IEO) Development of industrial protocol from results</p> <p>15.4.2 (FCPCT) Ossification patterns and skeletal deform. at different levels of intensification</p> <p>15.4.3 (FORKYS) validation at SME</p>	<p><u>D15.5</u> Industrial protocol</p>

# Pikeperch strategy

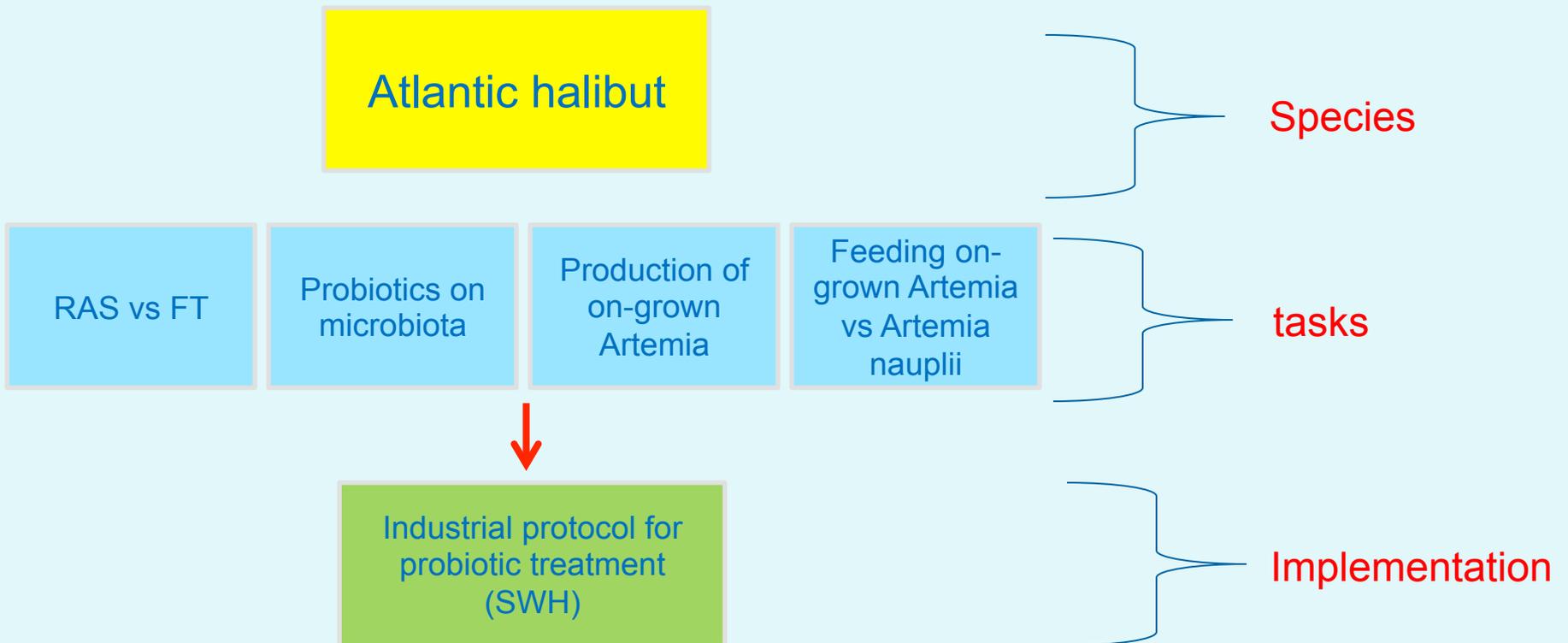




## WP16 pikeperch

Tasks	Evaluation	Sub-tasks	Deliverable
<p>16.1 Optimal combination of factors for improved larval rearing</p> <p>16.2 Development of industrial protocol</p>	<p>(UL, DTU) Effects of multifactorial designed experiments to test various factors (environ+nutritional+population), singly, in combination and interaction in terms of</p> <ul style="list-style-type: none"><li>• cannibalistic behavior, larval morphogenesis (DTU),</li><li>• ontogeny of skeleton, skeletal deform (IRTA).,</li><li>• gene expression of transcription factors, signaling molecules, ECM of skeletal system, digestive hormones (?)</li></ul> <p>Tested by SME ASIALOR</p>		<p><u>D16.1</u> Effect of environmental factors on larval rearing</p> <p><u>D16.2</u> Effect of nutritional factors on larval rearing</p> <p>D16.3 Effect of population factors on larval rearing</p> <p><u>16.4</u> Identification of optimal combinations of factors</p> <p><u>16.4</u> Evaluation of selected combinations under farm conditions.</p> <p><u>16.5</u> Industrial protocol for pikeperch larval rearing</p>

# Atlantic halibut strategy

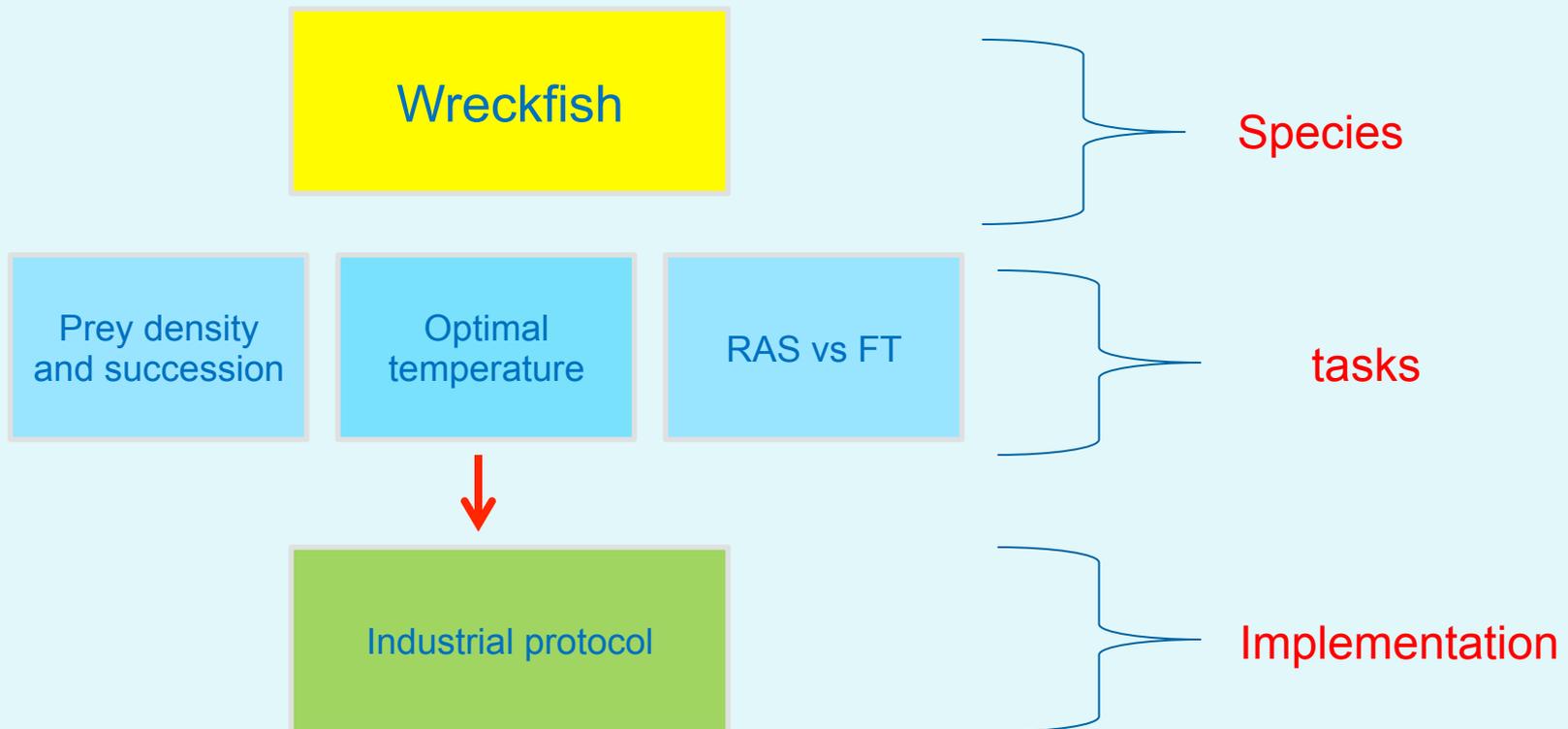




## WP17 Atlantic halibut

Tasks	Evaluation	Sub-tasks	Deliverable
<p><b>17.1</b> Recirculation vs Flow through (IMR)</p> <p><b>17.2</b> (IMR) Effects of probiotics on larval microbiota, survival and development of industrial protocol</p> <p><b>17.3</b> (IMR, SWH) Production of on-grown Artemia</p> <p><b>17.4</b> (NIFES, ULL) On grown Artemia vs Artemia nauplii until end of metamorphosis</p>	<ul style="list-style-type: none"> <li>•Growth, survival, pigmentation, eye migration (IMR, SWH),</li> <li>•Gut morphology (NIFES)</li> <li>•Digestive physiology (ULL-WP11)</li> <li>•Bacterial flora (IMR).</li> <li>•Nutrient analysis (NIFES)</li> <li>•Feeding, washing, disinfection of Artemia (IMR, SWH)</li> </ul>	<p>Based on results from 17.1, 17.2, full scale trial performed and industrial protocol for probiotic use.</p>	<p><u>D17.1</u> Production protocol of on-grown <i>Artemia</i>.</p> <p><u>D17.2</u> Determine if RAS more effective than FT</p> <p><u>D17.3</u> Effect of probiotics on larval microbiota and survival</p> <p><u>D17.4</u> Comparison of feeding on-grown <i>Artemia</i> vs <i>Artemia</i> nauplii on larval performance</p> <p><u>D17.5</u> Industrial protocol for probiotic treatment of larvae.</p>

# Wreckfish strategy

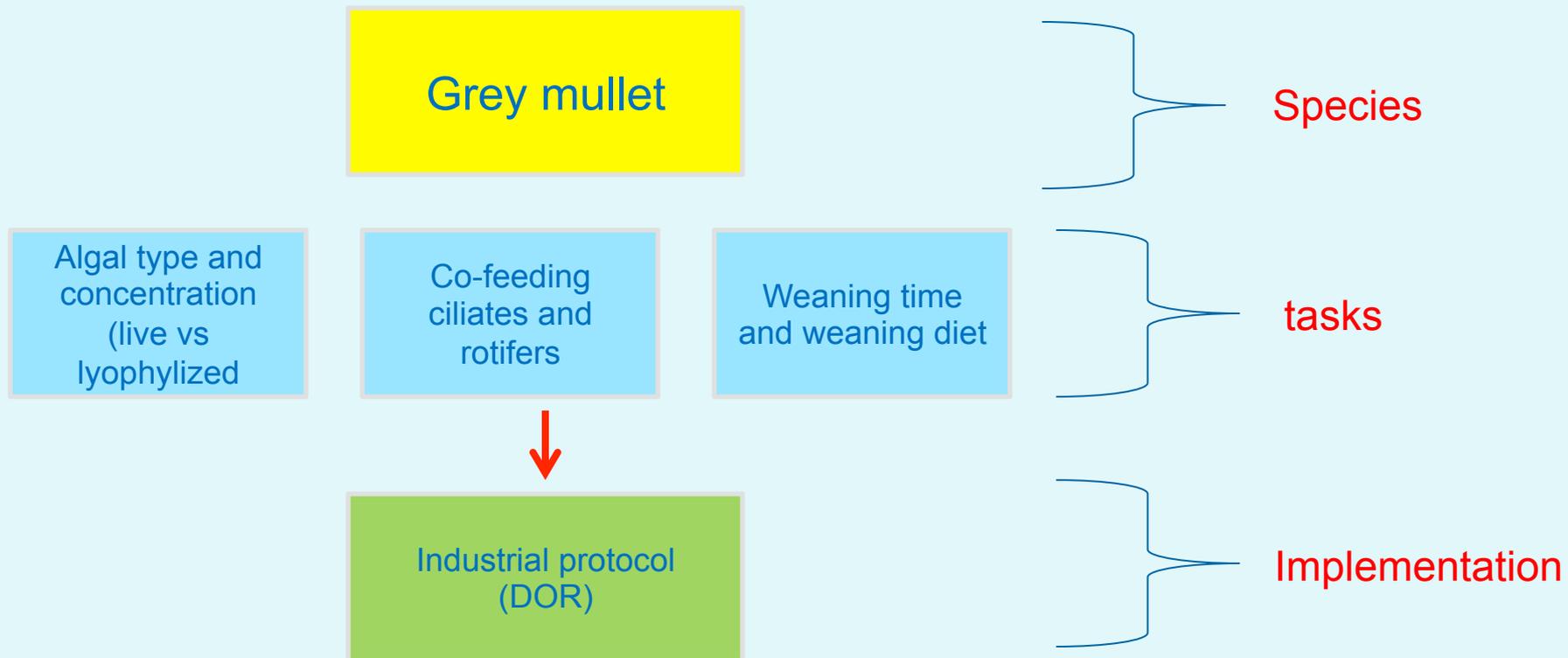




## WP18 Wreckfish

Tasks	Evaluation	Sub-tasks	Deliverable
<p><b>18.1</b> Development of feeding methodology (HCMR)- prey densities and succession of prey type</p> <p><b>18.2</b> Defining optimum conditions for larval rearing (IEO, MC2)</p>	<ul style="list-style-type: none"> <li>•Ontogeny of larval DT and visual system.</li> <li>•Ontogeny of DT enzymes</li> <li>•Growth, larval quality, (HCMR, IEO)</li> <li>•Growth survival, larval quality, size(IEO, MC2)</li> <li>•Larval biochemical profile (protein, lipids, EFA), biometric analysis, survival. (IEO, CMRM)</li> </ul>	<p><b>18.2.1</b> Testing the effect of two temperature ranges (14-17 and 19-22 °C</p> <p><b>18.2.2</b> Test RAS (CMRM) vs FT (IEO)</p>	<p><u>D18.1</u> Development of the DT of larval wreckfish</p> <p><u>D18.2</u> Determine optimal temperature conditions for rearing larval wreckfish</p> <p><u>D18.3</u> Develop a feeding protocol for wreckfish larvae</p> <p><u>D18.4</u> Determine the most effective culture system (RAS vs FT)</p>

# Grey mullet strategy



## WP 19 Grey mullet

Tasks	Evaluation	Sub-tasks	Deliverable
19.1 Effect of algal type and concentration on larval performance (IOLR)	<ul style="list-style-type: none"> <li>•Ingest. rate, growth survival, body comp., metamorphic synchrony, ontogeny of DT (IOLR,?)</li> </ul>	19.1.1 Determine algal type (Nan. Vs Iso) and concentr. in larval rearing 19.1.2 benefit of algal addition due to back-lighting or other factors 19.1.3 weaning diets varying in protein, cho, lipid at suggested weaning time and weaning diet change	D19.1 Determine the most effective algal type and concentration
19.2 Comparing selected algal protocol with lyophilized algae (IRTA)	<ul style="list-style-type: none"> <li>•Growth, survival, ingest. Rate, body comp., matur. of DT, skeletal deformities (IRTA)</li> </ul>		19.2 Evaluate effectiveness of replacing live algae with lyophilized algae
19.3 Determine the effect of co-feeding ciliates and rotifers on DT maturation and enzyme production (IOLR, Zoopt)	<ul style="list-style-type: none"> <li>•Gene expression of alk. Phosphatase and PepT1, growth, survival, FAA, BAA, FA (IOLR)</li> </ul>		19.3 Effect of co-feeding rotifers and ciliates
19.4 Determine weaning time and weaning diet based on shift from carnivorous to herbivorous feeding	<ul style="list-style-type: none"> <li>•Gene expression of Alk. Ph., PepT1,growth, survival, FA (IOLR)</li> <li>•Alk. Prot., amylase, lipase, trypsin, pepsin, alk. Ph., cytosol enz. (leu-ala pept.) (IRTA)</li> </ul>		19.4 Determine weaning time and weaning feed according to shift from carnivorous to omnivorous feeding.  19.5 Evaluate an improved grey mullet larval rearing protocol in commercial hatchery
19.5 Testing improved larval rearing protocol in commercial hatchery (DOR)	<ul style="list-style-type: none"> <li>•Growth, survival</li> </ul>		

# Common Themes in Larval Husbandry GWP

- Intensive vs extensive system (mesocosm)
- Tank greening and algal selection
- Improving live food performance
- Prey densities and feeding frequencies
- Effect of probiotic-immunostimulant substances
- RAS vs FT
- Environmental conditions on larval rearing
- Advancing weaning period